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(57) Abstract <p>The present invention is drawn to pesticidal strains and proteins. <i>Bacillus</i> strains which are capable of producing pesticidal proteins and auxiliary proteins during vegetative growth are provided. Also provided are the purified proteins, nucleotide sequences encoding the proteins and methods for using the strains, proteins and genes for controlling pests.</p>			

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NOVEL PESTICIDAL PROTEINS AND STRAINS

The present invention is drawn to methods and compositions for controlling plant and non-plant pests. Particularly, new pesticidal proteins are disclosed which are isolatable from the vegetative growth stage of *Bacillus*. *Bacillus* strains, proteins, and genes encoding the proteins are provided. The methods and compositions of the invention may be used in a variety of systems for controlling plant and non-plant pests.

Insect pests are a major factor in the loss of the world's commercially important agricultural crops. Broad spectrum chemical pesticides have been used extensively to control or eradicate pests of agricultural importance. There is, however, substantial interest in developing effective alternative pesticides.

Microbial pesticides have played an important role as alternatives to chemical pest control. The most extensively used microbial product is based on the bacterium *Bacillus thuringiensis* (Bt). Bt is a gram-positive spore forming *Bacillus* which produces an insecticidal crystal protein (ICP) during sporulation.

Numerous varieties of Bt are known that produce more than 25 different but related ICP's. The majority of ICP's made by Bt are toxic to larvae of certain insects in the orders *Lepidoptera*, *Diptera* and *Coleoptera*. In general, when an ICP is ingested by a susceptible insect the crystal is solubilized and transformed into a toxic moiety by the insect gut proteases. None of the ICP's active against coleopteran larvae such as Colorado potato beetle (*Leptinotarsa decemlineata*) or Yellow mealworm (*Tenebrio molitor*) have demonstrated significant effects on members of the genus *Diabrotica* particularly *Diabrotica virgifera virgifera*, the western corn rootworm (WCRW) or *Diabrotica longicornis barberi*, the northern corn rootworm.

Bacillus cereus (Bc) is closely related to Bt. A major distinguishing characteristic is the absence of a parasporal crystal in Bc. Bc is a widely distributed bacterium that is commonly found in soil and has been isolated from a variety of foods and drugs. The organism has been implicated in the spoilage of food.

Although Bt has been very useful in controlling insect pests, there is a need to expand the number of potential biological control agents.

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Within the present invention compositions and methods for controlling plant pests are provided. In particular, novel pesticidal proteins are provided which are produced during vegetative growth of *Bacillus* strains. The proteins are useful as pesticidal agents.

More specifically, the present invention relates to a substantially purified *Bacillus* strain which produces a pesticidal protein during vegetative growth wherein said *Bacillus* is not *B. sphaericus* SSII-1. Preferred are a *Bacillus cereus* strain having Accession No. NRRL B-21058 and *Bacillus thuringiensis* strain having Accession No. NRRL B-21060. Also preferred is a *Bacillus* strain selected from Accession Numbers NRRL B-21224, NRRL B-21225, NRRL B-21226, NRRL B-21227, NRRL B-21228, NRRL B-21229, NRRL B-21230, and NRRL B-21439.

The invention further relates to an insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp, but preferably of a *Bacillus thuringiensis* and *B. cereus* strain, and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1. The insect-specific protein of the invention is preferably toxic to Coleoptera or Lepidoptera insects and has a molecular weight of about 30 kDa or greater, preferably of about 60 to about 100 kDa, and more preferably of about 80 kDa.

More particularly, the insect-specific protein of the invention has a spectrum of insecticidal activity that includes an activity against *Agrotis* and/or *Spodoptera* species, but preferably a black cutworm [*Agrotis ipsilon* ; BCW] and/or fall armyworm [*Spodoptera frugiperda*] and/or beet armyworm [*Spodoptera exigua*] and/or tobacco budworm and/or corn earworm [*Helicoverpa zea*] activity.

The insect-specific protein of the invention can preferably be isolated, for example, from *Bacillus cereus* having Accession No. NRRL B-21058, or from *Bacillus thuringiensis* having Accession No. NRRL B-21060.

The insect-specific protein of the invention can also preferably be isolated from a *Bacillus* spp strain selected from Accession Numbers NRRL B-21224, NRRL B-21225, NRRL B-21226, NRRL B-21227, NRRL B-21228, NRRL B-21229, NRRL B-21230, and NRRL B-21439.

The present invention especially encompasses an insect-specific protein that has the amino acid sequence selected from the group consisting of SEQ ID NO:5 and

SEQ ID NO:7, including any proteins that are structurally and/or functionally homologous thereto.

Further preferred is an insect-specific protein, wherein said protein has the sequence selected from the group consisting of SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:29 SEQ ID NO:32 and SEQ ID NO:2, including any proteins that are structurally and/or functionally homologous thereto.

Especially preferred is an insect-specific protein, wherein said protein has the sequence selected from the group consisting of SEQ ID NO:29 and SEQ ID NO:32, including any proteins that are structurally and/or functionally homologous thereto.

A further preferred embodiment of the invention comprises an insect-specific protein of the invention, wherein the sequences representing the secretion signal have been removed or inactivated.

The present invention further encompasses auxiliary proteins which enhance the insect-specific activity of an insect-specific protein. The said auxiliary proteins preferably have a molecular weight of about 50 kDa and can be isolated, for example, from the vegetative growth phase of a *Bacillus cereus* strain, but especially of *Bacillus cereus* strain AB78.

A preferred embodiment of the invention relates to an auxiliary protein, wherein the sequences representing the secretion signal have been removed or inactivated.

The present invention further relates to multimeric pesticidal proteins, which comprise more than one polypeptide chain and wherein at least one of the said polypeptide chains represents an insect-specific protein of the invention and at least one of the said polypeptide chains represents an auxiliary protein of the invention, which activates or enhances the pesticidal activity of the said insect-specific protein.

The multimeric pesticidal proteins according to the invention preferably have a molecular weight of about 50 kDa to about 200 kDa.

The invention especially encompasses a multimeric pesticidal protein, which comprises an insect-specific protein of the invention and an auxiliary protein according to the invention, which activates or enhances the pesticidal activity of the said insect-specific protein.

The present invention further relates to fusion proteins comprising several protein domains including at least an insect-specific protein of the invention and/or an auxiliary protein according to the invention produced by in frame genetic fusions,

which, when translated by ribosomes, produce a fusion protein with at least the combined attributes of the insect-specific protein of the invention and/or an auxiliary protein according to the invention and, optionally, of the other components used in the fusion.

A specific embodiment of the invention relates to a fusion protein comprising a ribonuclease S-protein, an insect-specific protein of the invention and an auxiliary protein according to the invention.

A further specific embodiment of the invention relates to a fusion protein comprising an insect-specific protein according to the invention and an auxiliary protein according to the invention having either the insect-specific protein or the auxiliary protein at the N-terminal end of the said fusion protein.

Preferred is a fusion protein, which comprises an insect-specific protein as given in SEQ ID NO:5 and an auxiliary protein as given in SEQ ID NO: 2 resulting in the protein given in SEQ ID NO: 23, including any proteins that are structurally and/or functionally homologous thereto.

Also preferred is a fusion protein, which comprises an insect-specific protein as given in SEQ ID NO:35 and an auxiliary protein as given in SEQ ID NO: 27 resulting in the protein given in SEQ ID NO: 50, including any proteins that are structurally and/or functionally homologous thereto.

The invention further relates to a fusion protein comprising an insect-specific protein of the invention and/or an auxiliary protein according to the invention fused to a signal sequence, preferably a secretion signal sequence or a targeting sequence that directs the transgene product to a specific organelle or cell compartment, which signal sequence is of heterologous origin with respect to the recipient protein.

Especially preferred within this invention is a fusion protein wherein the said protein has a sequence as given in SEQ ID NO: 43, or in SEQ ID NO: 46, including any proteins that are structurally and/or functionally homologous thereto.

As used in the present application, substantial sequence homology means close structural relationship between sequences of amino acids. For example, substantially homologous proteins may be 40% homologous, preferably 50% and most preferably 60% or 80% homologous, or more. Homology also includes a relationship wherein one or several subsequences of amino acids are missing, or subsequences with additional amino acids are interdispersed.

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A further aspect of the invention relates to a DNA molecule comprising a nucleotide sequence which encodes an insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1. In particular, the present invention relates to a DNA molecule comprising a nucleotide sequence which encodes an insect-specific protein wherein the spectrum of insecticidal activity includes an activity against *Agrotis* and/or *Spodoptera* species, but preferably a black cutworm [*Agrotis ipsilon*; BCW] and/or fall armyworm [*Spodoptera frugiperda*] and/or beet armyworm [*Spodoptera exigua*] and/or tobacco budworm and/or corn earworm [*Helicoverpa zea*] activity.

Preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ, ID NO: 4, or SEQ ID NO: 6, including any DNA molecules that are structurally and/or functionally homologous thereto.

Also preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given SEQ ID NO:19, SEQ ID NO:28, SEQ ID NO:31, or SEQ ID NO:1, including any DNA molecules that are structurally and/or functionally homologous thereto.

The invention further relates to a DNA molecule comprising a nucleotide sequence which encodes an auxiliary protein according to the invention which enhances the insect-specific activity of an insect-specific protein.

Preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given SEQ ID NO:19, including any DNA molecules that are structurally and/or functionally homologous thereto.

A further embodiment of the invention relates to a DNA molecule comprising a nucleotide sequence which encodes an insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1, which nucleotide sequence has been optimized for expression in a microorganism or a plant.

Preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:17 or SEQ ID NO:18, including any DNA molecules that are structurally and/or functionally homologous thereto.

Also preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:27, or

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SEQ ID NO:30, including any DNA molecules that are structurally and/or functionally homologous thereto.

The invention further relates to a DNA molecule which comprises a nucleotide sequence encoding a multimeric pesticidal protein, which comprises more than one polypeptide chains and wherein at least one of the said polypeptide chains represents an insect-specific protein of the invention and at least one of the said polypeptide chains represents an auxiliary protein according to the invention, which activates or enhances the pesticidal activity of the said insect-specific protein.

Preferred is a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein of the invention and an auxiliary protein according to the invention, which activates or enhances the pesticidal activity of the said insect-specific protein.

Especially preferred is a DNA molecule, wherein said molecule comprises a nucleotide sequence as given in SEQ ID NO:1 or SEQ ID NO:19, including any nucleotide sequences that are structurally and/or functionally homologous thereto. A further embodiment of the invention relates to a DNA molecule which comprises a nucleotide sequence encoding a fusion protein comprising several protein domains including at least an insect-specific protein of the invention and/or an auxiliary protein according to the invention produced by in frame genetic fusions, which, when translated by ribosomes, produce a fusion protein with at least the combined attributes of the insect-specific protein of the invention and/or an auxiliary protein according to the invention and, optionally, of the other components used in the fusion.

Preferred within the invention is a DNA molecule which comprises a nucleotide sequence encoding a fusion protein comprising an insect-specific protein according to the invention and an auxiliary protein according to the invention having either the insect-specific protein or the auxiliary protein at the N-terminal end of the said fusion protein. Especially preferred is a DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:22, including any DNA molecules that are structurally and/or functionally homologous thereto.

The invention further relates to a DNA molecule which comprises a nucleotide sequence encoding a fusion protein comprising an insect-specific protein of the invention and/or an auxiliary protein of the invention fused to a signal sequence, preferably a secretion signal sequence or a targeting sequence that directs the

transgene product to a specific organelle or cell compartment, which signal sequence is of heterologous origin with respect to the recipient DNA.

The present invention further encompasses a DNA molecule comprising a nucleotide sequence encoding a fusion protein or a multimeric protein according to the invention that has been optimized for expression in a microorganism or plant.

Preferred is an optimized DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:42, SEQ ID NO:45, or SEQ ID NO:49, including any DNA molecules that are structurally and/or functionally homologous thereto.

The invention further relates to an optimized DNA molecule, wherein the sequences encoding the secretion signal have been removed from its 5' end, but especially to an optimized DNA molecule, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO: 35 or SEQ ID NO:39, including any DNA molecules that are structurally and/or functionally homologous thereto.

As used in the present application, substantial sequence homology means close structural relationship between sequences of nucleotides. For example, substantially homologous DNA molecules may be 60% homologous, preferably 80% and most preferably 90% or 95% homologous, or more. Homology also includes a relationship wherein one or several subsequences of nucleotides or amino acids are missing, or subsequences with additional nucleotides or amino acids are interdispersed.

Also comprised by the present invention are DNA molecules which hybridizes to a DNA molecule according to the invention as defined hereinbefore, but preferably to an oligonucleotide probe obtainable from said DNA molecule comprising a contiguous portion of the coding sequence for the said insect-specific protein at least 10 nucleotides in length, under moderately stringent conditions and which molecules have insect-specific activity and also the insect-specific proteins being encoded by the said DNA molecules.

Preferred are DNA molecules, wherein hybridization occurs at 65°C in a buffer comprising 7% SDS and 0.5 M sodium phosphate.

Especially preferred is a DNA molecule comprising a nucleotide sequence which encodes an insect-specific protein according to the invention obtainable by a process comprising

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- (a) obtaining a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein; and
- (b) hybridizing said DNA molecule with an oligonucleotide probe according to claim 107 obtained from a DNA molecule comprising a nucleotide sequence as given in SEQ ID NO: 28, SEQ ID NO: 30, or SEQ ID NO: 31; and
- (c) isolating said hybridized DNA.

The invention further relates to an insect-specific protein, wherein the said protein is encoded by a DNA molecule according to the invention.

Also encompassed by the invention is an expression cassette comprising a DNA molecule according to the invention operably linked to expression sequences including the transcriptional and translational regulatory signals necessary for expression of the associated DNA constructs in a host organism, preferably a microorganism or a plant, and optionally further regulatory sequences.

The invention further relates to a vector molecule comprising an expression cassette according to the invention.

The expression cassette and/or the vector molecule according to the invention are preferably part of the plant genome.

A further embodiment of the invention relates to a host organism, preferably a host organism selected from the group consisting of plant and insect cells, bacteria, yeast, baculoviruses, protozoa, nematodes and algae, comprising a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the genome of the host organism.

The invention further relates to a transgenic plant, but preferably a maize plant, including parts as well as progeny and seed thereof comprising a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the plant genome.

Preferred is a transgenic plant including parts as well as progeny and seed thereof which has been stably transformed with a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette.

Also preferred is a transgenic plant including parts as well as progeny and seed thereof which expresses an insect-specific protein according to the invention.

The invention further relates to a transgenic plant, preferably a maize plant, according to the invention as defined hereinbefore, which further expresses a second distinct insect control principle, but preferably a *Bt* δ-endotoxin. The said plant is preferably a hybrid plant.

Parts of transgenic plants are to be understood within the scope of the invention to comprise, for example, plant cells, protoplasts, tissues, callus, embryos as well as flowers, stems, fruits, leaves, roots originating in transgenic plants or their progeny previously transformed with a DNA molecule according to the invention and therefore consisting at least in part of transgenic cells, are also an object of the present invention.

The invention further relates to plant propagating material of a plant according to the invention, which is treated with a seed protectant coating.

The invention further encompasses a microorganism transformed with a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, wherein the said microorganism is preferably a microorganism that multiply on plants and more preferably a root colonizing bacterium.

A further embodiment of the invention relates to an encapsulated insect-specific protein which comprises a microorganism comprising an insect specific protein according to the invention.

The invention also relates to an entomocidal composition comprising a host organism of the invention, but preferably a purified *Bacillus* strain, in an insecticidally-effective amount together with a suitable carrier.

Further comprised by the invention is an entomocidal composition comprising an isolated protein molecule according to the invention, alone or in combination with a host organism of the invention and/or an encapsulated insect-specific protein according to the invention, in an insecticidally-effective amount, together with a suitable carrier.

A further embodiment of the invention relates to a method of obtaining a purified insect-specific protein according to the invention, said method comprising applying a

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solution comprising said insect-specific protein to a NAD column and eluting bound protein.

Also comprised is a method for identifying insect activity of an insect-specific protein according to the invention, said method comprising:

- growing a *Bacillus* strain in a culture;
- obtaining supernatant from said culture;
- allowing insect larvae to feed on diet with said supernatant; and,
- determining mortality.

Another aspect of the invention relates to a method for isolating an insect-specific protein according to the invention, said method comprising:

- growing a *Bacillus* strain in a culture;
- obtaining supernatant from said culture; and,
- isolating said insect-specific protein from said supernatant.

The invention also encompasses a method for isolating a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein exhibiting the insecticidal activity of the proteins according to the invention, said method comprising:

- obtaining a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein; and
- hybridizing said DNA molecule with DNA obtained from a *Bacillus* species; and
- isolating said hybridized DNA.

The invention further relates to a method of increasing insect target range by using an insect specific protein according to the invention in combination with at least one second insecticidal protein that is different from the insect specific protein according to the invention, but preferably with an insecticidal protein selected from the group consisting of *Bt* δ-endotoxins, protease inhibitors, lectins, α-amylases and peroxidases.

Preferred is a method for increasing insect target range within a plant by expressing within the said plant a insect specific protein according to the invention in combination with at least one second insecticidal protein that is different from the insect specific protein according to the invention, but preferably with an insecticidal protein selected from the group consisting of *Bt* δ-endotoxins, protease inhibitors, lectins, α-amylases and peroxidases.

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Also comprised is a method of protecting plants against damage caused by an insect pest, but preferably by *Spodoptera* and/or *Agrotis* species, and more preferably by an insect pest selected from the group consisting of black cutworm [*Agrotis ipsilon* ; BCW], fall armyworm [*Spodoptera frugiperda*], beet armyworm [*Spodoptera exigua*], tobacco budworm and corn earworm [*Helicoverpa zea*] comprising applying to the plant or the growing area of the said plant an entomocidal composition or a toxin protein according to the invention.

The invention further relates to method of protecting plants against damage caused by an insect pest, but preferably by *Spodoptera* and/or *Agrotis* species, and more preferably by an insect pest selected from the group consisting of black cutworm [*Agrotis ipsilon* ; BCW], fall armyworm [*Spodoptera frugiperda*], beet armyworm [*Spodoptera exigua*], tobacco budworm and corn earworm [*Helicoverpa zea*] comprising planting a transgenic plant expressing a insect-specific protein according to the invention within an area where the said insect pest may occur.

The invention also encompasses a method of producing a host organism which comprises stably integrated into its genome a DNA molecule according to the invention and preferably expresses an insect-specific protein according to the invention comprising transforming the said host organism with a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette.

A further embodiment of the invention relates to a method of producing a transgenic plant or plant cell which comprises stably integrated into the plant genome a DNA molecule according to the invention and preferably expresses an insect-specific protein according to the invention comprising transforming the said plant and plant cell, respectively, with a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette.

The invention also relates to a method of producing an entomocidal composition comprising mixing an isolated *Bacillus* strain and/or a host organism and/or an isolated protein molecule, and/or an encapsulated protein according to the invention in an insecticidally-effective amount with a suitable carrier.

The invention also encompasses a method of producing transgenic progeny of a transgenic parent plant comprising stably incorporated into the plant genome a DNA

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molecule comprising a nucleotide sequence encoding an insect-specific protein according to the invention comprising transforming the said parent plant with a DNA molecule according to the invention, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette and transferring the pesticidal trait to the progeny of the said transgenic parent plant involving known plant breeding techniques.

Also encompassed by the invention is oligonucleotide probe capable of specifically hybridizing to a nucleotide sequence encoding a insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1, wherein said probe comprises a contiguous portion of the coding sequence for the said insect-specific protein at least 10 nucleotides in length and the use of the said oligonucleotide probe for screening of any *Bacillus* strain or other organisms to determine whether the insect-specific protein is naturally present or whether a particular transformed organism includes the said gene

The present invention recognizes that pesticidal proteins are produced during vegetative growth of *Bacillus* strains. Having recognized that such a class exists, the present invention embraces all vegetative insecticidal proteins, hereinafter referred to as VIPs, except for the mosquitocidal toxin from *B. sphaericus*.

The present VIPs are not abundant after sporulation and are particularly expressed during log phase growth before stationary phase. For the purpose of the present invention vegetative growth is defined as that period of time before the onset of sporulation. Genes encoding such VIPs can be isolated, cloned and transformed into various delivery vehicles for use in pest management programs.

For purposes of the present invention, pests include but are not limited to insects, fungi, bacteria, nematodes, mites, ticks, protozoan pathogens, animal-parasitic liver flukes, and the like. Insect pests include insects selected from the orders Coleoptera, Diptera, Hymenoptera, Lepidoptera, Mallophaga, Homoptera, Hemiptera, Orthoptera, Thysanoptera, Dermaptera, Isoptera, Anoplura, Siphonaptera, Trichoptera, etc., particularly Coleoptera and Lepidoptera.

Tables 1 - 10 gives a list of pests associated with major crop plants and pests of human and veterinary importance. Such pests are included within the scope of the present invention.

TABLE 1

Lepidoptera (Butterflies and Moth)

Maize

Ostrinia nubilalis, European corn borer
Agrotis ipsilon, black cutworm
Helicoverpa zea, corn earworm
Spodoptera frugiperda, fall armyworm
Diatraea grandiosella, southwestern corn borer
Elasmopalpus lignosellus, lesser cornstalk borer
Diatraea saccharalis, sugarcane borer

Sorghum

Chilo partellus, sorghum borer
Spodoptera frugiperda, fall armyworm
Helicoverpa zea, corn earworm
Elasmopalpus lignosellus, lesser cornstalk borer
Feltia subterranea, granulate cutworm

Wheat

Pseudaletia unipunctata, army worm
Spodoptera frugiperda, fall armyworm
Elasmopalpus lignosellus, lesser cornstalk borer
Agrotis orthogonia, pale western cutworm
Elasmopalpus lignosellus, lesser cornstalk borer

Sunflower

Suleima helianthana, sunflower bud moth
Homoeosoma electellum, sunflower moth

Cotton

Heliothis virescens, cotton boll worm
Helicoverpa zea, cotton bollworm
Spodoptera exigua, beet armyworm
Pectinophora gossypiella, pink bollworm

Rice

Diatraea saccharalis, sugarcane borer
Spodoptera frugiperda, fall armyworm
Helicoverpa zea, corn earworm

Soybean

Pseudoplusia includens, soybean looper
Anticarsia gemmatalis, velvetbean caterpillar
Plathypena scabra, green cloverworm
Ostrinia nubilalis, European corn borer
Agrotis ipsilon, black cutworm
Spodoptera exigua, beet armyworm
Heliothis virescens, cotton boll worm
Helicoverpa zea, cotton bollworm

Barley

Ostrinia nubilalis, European corn borer
Agrotis ipsilon, black cutworm

TABLE 2

Coleoptera (Beetles)**Maize**

Diabrotica virgifera virgifera, western corn rootworm
Diabrotica longicornis barberi, northern corn rootworm
Diabrotica undecimpunctata howardi, southern corn rootworm
Melanotus spp., wireworms
Cyclocephala borealis, northern masked chafer (white grub)
Cyclocephala immaculata, southern masked chafer (white grub)
Popillia japonica, Japanese beetle
Chaetocnema pulicaria, corn flea beetle
Sphenophorus maidis, maize billbug

Sorghum

Phyllophaga crinita, white grub
Eleodes, *Conoderus*, and *Aeolus spp.*, wireworms
Oulema melanopus, cereal leaf beetle
Chaetocnema pulicaria, corn flea beetle
Sphenophorus maidis, maize billbug

Wheat

Oulema melanopus, cereal leaf beetle
Hypera punctata, clover leaf weevil
Diabrotica undecimpunctata howardi, southern corn rootworm

Sunflower

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Zygogramma exclamationis, sunflower beetle
Bothyrus gibbosus, carrot beetle

Cotton

Anthonomus grandis, boll weevil

Rice

Colaspis brunnea, grape colaspis
Lissorhoptrus oryzophilus, rice water weevil
Sitophilus oryzae, rice weevil

Soybean

Epilachna varivestis, Mexican bean beetle

TABLE 3

Homoptera (Whiteflies, Aphids etc..)

Maize

Rhopalosiphum maidis, corn leaf aphid
Anuraphis maidiradicis, corn root aphid

Sorghum

Rhopalosiphum maidis, corn leaf aphid
Sipha flava, yellow sugarcane aphid

Wheat

Russian wheat aphid
Schizaphis graminum, greenbug
Macrosiphum avenae, English grain aphid

Cotton

Aphis gossypii, cotton aphid
Pseudatomoscelis seriatus, cotton fleahopper
Trialeurodes abutilonea, bandedwinged whitefly

Rice

Nephrotettix nigropictus, rice leafhopper

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Soybean

Myzus persicae, green peach aphid
Empoasca fabae, potato leafhopper

Barley

Schizaphis graminum, greenbug

Oil Seed Rape

Brevicoryne brassicae, cabbage aphid

TABLE 4

Hemiptera (Bugs)

Maize

Blissus leucopterus leucopterus, chinch bug

Sorghum

Blissus leucopterus leucopterus, chinch bug

Cotton

Lygus lineolaris, tarnished plant bug

Rice

Blissus leucopterus leucopterus, chinch bug
Acrosternum hilare, green stink bug

Soybean

Acrosternum hilare, green stink bug

Barley

Blissus leucopterus leucopterus, chinch bug
Acrosternum hilare, green stink bug
Euschistus servus, brown stink bug

TABLE 5

Orthoptera (Grasshoppers, Crickets, and Cockroaches)

Maize

Melanoplus femur-rubrum, redlegged grasshopper
Melanoplus sanguinipes, migratory grasshopper

Wheat

Melanoplus femur-rubrum, redlegged grasshopper
Melanoplus differentialis, differential grasshopper
Melanoplus sanguinipes, migratory grasshopper

Cotton

Melanoplus femur-rubrum, redlegged grasshopper
Melanoplus differentialis, differential grasshopper

Soybean

Melanoplus femur-rubrum, redlegged grasshopper
Melanoplus differentialis, differential grasshopper

Structural/Household

Periplaneta americana, American cockroach
Blattella germanica, German cockroach
Blatta orientalis, oriental cockroach

TABLE 6

Diptera (Flies and Mosquitoes)

Maize

Hylemya platura, seedcorn maggot
Agromyza parvicornis, corn blotch leafminer

Sorghum

Contarinia sorghicola, sorghum midge

Wheat

Mayetiola destructor, Hessian fly
Sitodiplosis mosellana, wheat midge
Meromyza americana, wheat stem maggot
Hylemya coarctata, wheat bulb fly

Sunflower

Neolasioptera murtfeldtiana, sunflower seed midge

Soybean

Hylemya platura, seedcorn maggot

Barley

Hylemya platura, seedcorn maggot
Mayetiola destructor, Hessian fly

Insects attacking humans and animals and disease carriers

Aedes aegypti, yellowfever mosquito
Aedes albopictus, forest day mosquito
Phlebotomus papatasii, sand fly
Musca domestica, house fly
Tabanus atratus, black horse fly
Cochliomyia hominivorax, screwworm fly

TABLE 7**Thysanoptera (Thrips)****Maize**

Anaphothrips obscurus, grass thrips

Wheat

Frankliniella fusca, tobacco thrips

Cotton

Thrips tabaci, onion thrips
Frankliniella fusca, tobacco thrips

Soybean

Sericothrips variabilis, soybean thrips
Thrips tabaci, onion thrips

TABLE 8**Hymenoptera (Sawflies, Ants, Wasps, etc.)****Maize**

Solenopsis milesta, thief ant

Wheat

Cephus cinctus, wheat stem sawfly

TABLE 9**Other Orders and Representative Species*****Dermoptera* (Earwigs)**

Forficula auricularia, European earwig

***Isoptera* (Termites)**

Reticulitermes flavipes, eastern subterranean termite

***Mallophaga* (Chewing Lice)**

Cuclotogaster heterographa, chicken head louse
Bovicola bovis, cattle biting louse

***Anoplura* (Sucking Lice)**

Pediculus humanus, head and body louse

***Siphonaptera* (Fleas)**

Ctenocephalides felis, cat flea

TABLE 10

Acari (Mites and Ticks)

Maize

Tetranychus urticae, twospotted spider mite

Sorghum

Tetranychus cinnabarinus, carmine spider mite
Tetranychus urticae, twospotted spider mite

Wheat

Aceria tulipae, wheat curl mite

Cotton

Tetranychus cinnabarinus, carmine spider mite
Tetranychus urticae, twospotted spider mite

Soybean

Tetranychus turkestanii, strawberry spider mite
Tetranychus urticae, twospotted spider mite

Barley

Petrobia latens, brown wheat mite

Important human and animal Acari

Demacentor variabilis, American dog tick
Argas persicus, fowl tick
Dermatophagoides farinae, American house dust mite
Dermatophagoides pteronyssinus, European house dust mite

Now that it has been recognized that pesticidal proteins can be isolated from the vegetative growth phase of *Bacillus*, other strains can be isolated by standard techniques and tested for activity against particular plant and non-plant pests. Generally *Bacillus* strains can be isolated from any environmental sample, including soil, plant, insect, grain elevator dust, and other sample material, etc., by methods

known in the art. See, for example, Travers *et al.* (1987) *Appl. Environ. Microbiol.* 53:1263-1266; Saleh *et al.* (1969) *Can J. Microbiol.* 15:1101-1104; DeLucca *et al.* (1981) *Can. J. Microbiol.* 27:865-870; and Norris, *et al.* (1981) "The genera *Bacillus* and *Sporolactobacillus*," In Starr *et al.* (eds.), *The Prokaryotes: A Handbook on Habitats, Isolation, and Identification of Bacteria*, Vol. II, Springer-Verlog Berlin Heidelberg. After isolation, strains can be tested for pesticidal activity during vegetative growth. In this manner, new pesticidal proteins and strains can be identified.

Such *Bacillus* microorganisms which find use in the invention include *Bacillus cereus* and *Bacillus thuringiensis*, as well as those *Bacillus* species listed in Table 11.

TABLE 11

List of *Bacillus* species

Morphological Group 1

- B. megaterium*
- B. cereus**
- B. cereus* var. *mycoides*
- B. thuringiensis**
- B. licheniformis*
- B. subtilis**
- B. pumilus*
- B. firmus**
- B. coagulans*

Morphological Group 2

- B. polymyxa*
- B. macerans*
- B. circulans*
- B. stearothermophilus*
- B. alvei**
- B. laterosporus**
- B. brevis*
- B. pulvifaciens*
- B. popilliae**
- B. lentimorbus**
- B. larvae**

Morphological Group 3

- B. sphaericus**
B. pasteurii

Unassigned Strains**Subgroup A**

- B. apiarus**
B. filicolicus
B. thiaminolyticus
B. alcalophilus

Subgroup B

- B. cirroflagellosum*
B. chitinosporus
B. latus

Subgroup C

- B. badius*
B. aneurinolyticus
B. macroides
B. freundreichii

Subgroup D

- B. pantothenicus*
B. epiphytus

Subgroup E1

- B. aminovorans*
B. globisporus
B. insolitus
B. psychrophilus

Subgroup E2

- B. psychrosaccharolyticus*
B. macquariensis

*=Those *Bacillus* strains that have been previously found associated with insects

Grouping according to Parry, J.M. et al. (1983) Color Atlas of *Bacillus* species, Wolfe Medical Publications, London.

In accordance with the present invention, the pesticidal proteins produced during vegetative growth can be isolated from *Bacillus*. In one embodiment, insecticidal proteins produced during vegetative growth, can be isolated. Methods for protein isolation are known in the art. Generally, proteins can be purified by conventional chromatography, including gel-filtration, ion-exchange, and immunoaffinity chromatography, by high-performance liquid chromatography, such as reversed-phase high-performance liquid chromatography, ion-exchange high-performance liquid chromatography, size-exclusion high-performance liquid chromatography, high-performance chromatofocusing and hydrophobic interaction chromatography, etc., by electrophoretic separation, such as one-dimensional gel electrophoresis, two-dimensional gel electrophoresis, etc. Such methods are known in the art. See for example Current Protocols in Molecular Biology, Vols. 1 and 2, Ausubel et al. (eds.), John Wiley & Sons, NY (1988). Additionally, antibodies can be prepared against substantially pure preparations of the protein. See, for example, Radka et al. (1983) J. Immunol. 128:2804; and Radka et al. (1984) Immunogenetics 19:63. Any combination of methods may be utilized to purify protein having pesticidal properties. As the protocol is being formulated, pesticidal activity is determined after each purification step.

Such purification steps will result in a substantially purified protein fraction. By "substantially purified" or "substantially pure" is intended protein which is substantially free of any compound normally associated with the protein in its natural state. "Substantially pure" preparations of protein can be assessed by the absence of other detectable protein bands following SDS-PAGE as determined visually or by densitometry scanning. Alternatively, the absence of other amino-terminal sequences or N-terminal residues in a purified preparation can indicate the level of purity. Purity can be verified by rechromatography of "pure" preparations showing the absence of other peaks by ion exchange, reverse phase or capillary electrophoresis. The terms "substantially pure" or "substantially purified" are not meant to exclude artificial or synthetic mixtures of the proteins with other compounds. The terms are also not meant to exclude the presence of minor impurities which do not interfere with the biological activity of the protein, and which may be present, for example, due to incomplete purification.

Once purified protein is isolated, the protein, or the polypeptides of which it is comprised, can be characterized and sequenced by standard methods known in the art. For example, the purified protein, or the polypeptides of which it is comprised, may be fragmented as with cyanogen bromide, or with proteases such as papain, chymotrypsin, trypsin, lysyl-C endopeptidase, etc. (Oike *et al.* (1982) J. Biol. Chem. 257:9751-9758; Liu *et al.* (1983) Int. J. Pept. Protein Res. 21:209-215). The resulting peptides are separated, preferably by HPLC, or by resolution of gels and electroblotting onto PVDF membranes, and subjected to amino acid sequencing. To accomplish this task, the peptides are preferably analyzed by automated sequenators. It is recognized that N-terminal, C-terminal, or internal amino acid sequences can be determined. From the amino acid sequence of the purified protein, a nucleotide sequence can be synthesized which can be used as a probe to aid in the isolation of the gene encoding the pesticidal protein.

It is recognized that the pesticidal proteins may be oligomeric and will vary in molecular weight, number of protomers, component peptides, activity against particular pests, and in other characteristics. However, by the methods set forth herein, proteins active against a variety of pests may be isolated and characterized.

Once the purified protein has been isolated and characterized it is recognized that it may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the pesticidal proteins can be prepared by mutations in the DNA. Such variants will possess the desired pesticidal activity. Obviously, the mutations that will be made in the DNA encoding the variant must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. See, EP Patent Application Publication No. 75,444.

In this manner, the present invention encompasses the pesticidal proteins as well as components and fragments thereof. That is, it is recognized that component protomers, polypeptides or fragments of the proteins may be produced which retain pesticidal activity. These fragments include truncated sequences, as well as N-terminal, C-terminal, internal and internally deleted amino acid sequences of the proteins.

Most deletions, insertions, and substitutions of the protein sequence are not expected to produce radical changes in the characteristics of the pesticidal protein. However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays.

The proteins or other component polypeptides described herein may be used alone or in combination. That is, several proteins may be used to control different insect pests.

Some proteins are single polypeptide chains while many proteins consist of more than one polypeptide chain, i.e., they are oligomeric. Additionally, some VIPs are pesticidally active as oligomers. In these instances, additional protomers are utilized to enhance the pesticidal activity or to activate pesticidal proteins. Those protomers which enhance or activate are referred to as auxiliary proteins. Auxiliary proteins activate or enhance a pesticidal protein by interacting with the pesticidal protein to form an oligomeric protein having increased pesticidal activity compared to that observed in the absence of the auxiliary protein.

Auxiliary proteins activate or increase the activity of pesticidal proteins such as the VIP1 protein from AB78. Such auxiliary proteins are exemplified by, but not limited to, the VIP2 protein from AB78. As demonstrated in the Experimental section of the application, auxiliary proteins can activate a number of pesticidal proteins. Thus, in one embodiment of the invention, a plant, Parent 1, can be transformed with an auxiliary protein. This Parent 1 can be crossed with a number of Parent 2 plants transformed with one or more pesticidal proteins whose pesticidal activities are activated by the auxiliary protein.

Amongst the pesticidal proteins of the invention a new class of insect-specific proteins could be surprisingly identified within the scope of the present invention. The said proteins, which are designated throughout this application as VIP3, can be obtained from *Bacillus spp* strains, but preferably from *Bacillus thuringiensis* strains and most preferably from *Bacillus thuringiensis* strains AB88 and AB424. The said VIPs are present mostly in the supernatants of *Bacillus* cultures amounting to at least 75% of the total in strain AB88. The VIP3 proteins are further characterized by their unique spectrum of insectical acitivity, which includes an activity against *Agrotis* and/or *Spodoptera* species, but especially a black cutworm [BCW] and/or fall

armyworm and/or beet armyworm and/or tobacco budworm and/or corn earworm activity.

Black cutworm is an agronomically important insect quite resistant to δ -endotoxins. MacIntosh et al (1990) J Invertebr Pathol 56, 258-266 report that the δ -endotoxins CryIA(b) and CryIA(c) possesses insecticidal properties against BCW with LC₅₀ of more than 80 μ g and 18 μ g/ml of diet respectively. The vip3A insecticidal proteins according to the invention provide >50% mortality when added in an amount of protein at least 10 to 500, preferably 50 to 350, and more preferably 200 to 300 fold lower than the amount of CryIA proteins needed to achieve just 50% mortality. Especially preferred within the invention are vip3A insecticidal proteins which provide 100% mortality when added in an amount of protein at least 260 fold lower than the amount of CryIA proteins needed to achieve just 50% mortality.

The vip3 insecticidal proteins according to the invention are present mostly in the supernatants of the cultures and are therefore are to be classified as secreted proteins. They preferably contain in the N-terminal sequence a number of positively charged residues followed by a hydrophobic core region and are not N-terminally processed during export.

As the other pesticidal proteins reported hereto within the scope of the invention, the VIP3 proteins can be detected in growth stages prior to sporulation establishing a further clear distinction from other proteins that belong to the δ -endotoxin family. Preferably, expression of the insect-specific protein starts during mid-log phase and continues during sporulation. Owing to the specific expression pattern in combination with the high stability of the VIP3 proteins, large amounts of the VIP3 proteins can be found in supernatants of sporulating cultures. Especially preferred are the VIP3 proteins identified in SEQ ID NO:29 and SEQ ID NO:32 and the corresponding DNA molecules comprising nucleotide sequences encoding the said proteins, but especially those DNA molecules comprising the nucleotide sequences given in SEQ ID NO:28, SEQ ID NO:30 and SEQ ID NO:31.

The pesticidal proteins of the invention can be used in combination with Bt endotoxins or other insecticidal proteins to increase insect target range. Furthermore, the use of the VIPs of the present invention in combination with Bt δ -endotoxins or other insecticidal principles of a distinct nature has particular utility for the prevention and/or management of insect resistance. Other insecticidal principles include

protease inhibitors (both serine and cysteine types), lectins, α -amylase and peroxidase. In one preferred embodiment, expression of VIPs in a transgenic plant is accompanied by the expression of one or more Bt δ -endotoxins. This co-expression of more than one insecticidal principle in the same transgenic plant can be achieved by genetically engineering a plant to contain and express all the genes necessary. Alternatively, a plant, Parent 1, can be genetically engineered for the expression of VIPs. A second plant, Parent 2, can be genetically engineered for the expression of Bt δ -endotoxin. By crossing Parent 1 with Parent 2, progeny plants are obtained which express all the genes introduced into Parents 1 and 2. Particularly preferred Bt δ -endotoxins are those disclosed in EP-A 0618976, herein incorporated by reference.

A substantial number of cytotoxic proteins, though not all, are binary in action. Binary toxins typically consist of two protein domains, one called the A domain and the other called the B domain (see Sourcebook of Bacterial Protein Toxins, J. E. Alouf and J. H. Freer eds.(1991) Academic Press). The A domain possesses a potent cytotoxic activity. The B domain binds an external cell surface receptor before being internalized. Typically, the cytotoxic A domain must be escorted to the cytoplasm by a translocation domain. Often the A and B domains are separate polypeptides or protomers, which are associated by a protein-protein interaction or a di-sulfide bond. However, the toxin can be a single polypeptide which is proteolytically processed within the cell into two domains as in the case for *Pseudomonas* exotoxin A. In summary binary toxins typically have three important domains, a cytotoxic A domain, a receptor binding B domain and a translocation domain. The A and B domain are often associated by protein-protein interacting domains.

The receptor binding domains of the present invention are useful for delivering any protein, toxin, enzyme, transcription factor, nucleic acid, chemical or any other factor into target insects having a receptor recognized by the receptor binding domain of the binary toxins described in this patent. Similarly, since binary toxins have translocation domains which penetrate phospholipid bilayer membranes and escort cytotoxins across those membranes, such translocation domains may be useful in escorting any protein, toxin, enzyme, transcription factor, nucleic acid, chemical or any other factor across a phospholipid bilayer such as the plasma membrane or a vesicle membrane. The translocation domain may itself perforate membranes, thus having toxic or insecticidal properties. Further, all binary toxins have cytotoxic domains; such a

cytotoxic domain may be useful as a lethal protein, either alone or when delivered into any target cell(s) by any means.

Finally, since binary toxins comprised of two polypeptides often form a complex, it is likely that there are protein-protein interacting regions within the components of the binary toxins of the invention. These protein-protein interacting domains may be useful in forming associations between any combination of toxins, enzymes, transcription factors, nucleic acids, antibodies, cell binding moieties, or any other chemicals, factors, proteins or protein domains.

Toxins, enzymes, transcription factors, antibodies, cell binding moieties or other protein domains can be fused to pesticidal or auxiliary proteins by producing in frame genetic fusions which, when translated by ribosomes, would produce a fusion protein with the combined attributes of the VIP and the other component used in the fusion. Furthermore, if the protein domain fused to the VIP has an affinity for another protein, nucleic acid, carbohydrate, lipid, or other chemical or factor, then a three-component complex can be formed. This complex will have the attributes of all of its components. A similar rationale can be used for producing four or more component complexes. These complexes are useful as insecticidal toxins, pharmaceuticals, laboratory reagents, and diagnostic reagents, etc. Examples where such complexes are currently used are fusion toxins for potential cancer therapies, reagents in ELISA assays and immunoblot analysis.

One strategy of altering pesticidal or auxiliary proteins is to fuse a 15-amino-acid "S-tag" to the protein without destroying the insect cell binding domain(s), translocation domains or protein-protein interacting domains of the proteins. The S-tag has a high affinity ($K_d = 10^{-9}$ M) for a ribonuclease S-protein, which, when bound to the S-tag, forms an active ribonuclease (See F. M. Richards and H. W. Wyckoff (1971) in "The Enzymes", Vol. IV (Boyer, P.D. ed.), pp. 647-806. Academic Press, New York). The fusion can be made in such a way as to destroy or remove the cytotoxic activity of the pesticidal or auxiliary protein, thereby replacing the VIP cytotoxic activity with a new cytotoxic ribonuclease activity. The final toxin would be comprised of the S-protein, a pesticidal protein and an auxiliary protein, where either the pesticidal protein or the auxiliary protein is produced as translational fusions with the S-tag. Similar strategies can be used to fuse other potential cytotoxins to pesticidal or auxiliary proteins including (but not limited to) ribosome inactivating

proteins, insect hormones, hormone receptors, transcription factors, proteases, phosphatases, *Pseudomonas* exotoxin A, or any other protein or chemical factor that is lethal when delivered into cells. Similarly, proteins can be delivered into cells which are not lethal, but might alter cellular biochemistry or physiology.

The spectrum of toxicity toward different species can be altered by fusing domains to pesticidal or auxiliary proteins which recognize cell surface receptors from other species. Such domains might include (but are not limited to) antibodies, transferrin, hormones, or peptide sequences isolated from phage displayed affinity selectable libraries. Also, peptide sequences which are bound to nutrients, vitamins, hormones, or other chemicals that are transported into cells could be used to alter the spectrum of toxicity. Similarly, any other protein or chemical which binds a cell surface receptor or the membrane and could be internalized might be used to alter the spectrum of activity of VIP1 and VIP2.

The pesticidal proteins of the present invention are those proteins which confer a specific pesticidal property. Such proteins may vary in molecular weight, having component polypeptides at least a molecular weight of 30 kDa or greater, preferably about 50 kDa or greater.

The auxiliary proteins of the invention may vary in molecular weight, having at least a molecular weight of about 15 kDa or greater, preferably about 20 kDa or greater; more preferably, about 30 kDa or greater. The auxiliary proteins themselves may have component polypeptides.

It is possible that the pesticidal protein and the auxiliary protein may be components of a multimeric, pesticidal protein. Such a pesticidal protein which includes the auxiliary proteins as one or more of its component polypeptides may vary in molecular weight, having at least a molecular weight of 50 kDa up to at least 200 kDa, preferably about 100 kDa to 150 kDa.

An auxiliary protein may be used in combination with the pesticidal proteins of the invention to enhance activity or to activate the pesticidal protein. To determine whether the auxiliary protein will affect activity, the pesticidal protein can be expressed alone and in combination with the auxiliary protein and the respective activities compared in feeding assays for pesticidal activity.

It may be beneficial to screen strains for potential pesticidal activity by testing activity of the strain alone and in combination with the auxiliary protein. In some

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instances an auxiliary protein in combination with the native proteins of the strains yields pesticidal activity where none is seen in the absence of an auxiliary protein.

The auxiliary protein can be modified, as described above, by various methods known in the art. Therefore, for purposes of the invention, the term "Vegetative Insecticidal Protein" (VIP) encompasses those proteins produced during vegetative growth which alone or in combination can be used for pesticidal activity. This includes pesticidal proteins, auxiliary proteins and those proteins which demonstrate activity only in the presence of the auxiliary protein or the polypeptide components of these proteins.

It is recognized that there are alternative methods available to obtain the nucleotide and amino acid sequences of the present proteins. For example, to obtain the nucleotide sequence encoding the pesticidal protein, cosmid clones, which express the pesticidal protein, can be isolated from a genomic library. From larger active cosmid clones, smaller subclones can be made and tested for activity. In this manner, clones which express an active pesticidal protein can be sequenced to determine the nucleotide sequence of the gene. Then, an amino acid sequence can be deduced for the protein. For general molecular methods, see, for example, *Molecular Cloning, A Laboratory Manual, Second Edition, Vols. 1-3, Sambrook et al. (eds.) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989)*, and the references cited therein.

The present invention also encompasses nucleotide sequences from organisms other than *Bacillus*, where the nucleotide sequences are isolatable by hybridization with the *Bacillus* nucleotide sequences of the invention. Proteins encoded by such nucleotide sequences can be tested for pesticidal activity. The invention also encompasses the proteins encoded by the nucleotide sequences. Furthermore, the invention encompasses proteins obtained from organisms other than *Bacillus* wherein the protein cross-reacts with antibodies raised against the proteins of the invention. Again the isolated proteins can be assayed for pesticidal activity by the methods disclosed herein or others well-known in the art.

Once the nucleotide sequences encoding the pesticidal proteins of the invention have been isolated, they can be manipulated and used to express the protein in a variety of hosts including other organisms, including microorganisms and plants.

The pesticidal genes of the invention can be optimized for enhanced expression in plants. See, for example EP-A 0618976; EP-A 0359472; EP-A 0385962; WO 91/16432; Perlak *et al.* (1991) Proc. Natl. Acad. Sci. USA 88:3324-3328; and Murray *et al.* (1989) Nucleic Acids Research 17: 477-498. In this manner, the genes can be synthesized utilizing plant preferred codons. That is the preferred codon for a particular host is the single codon which most frequently encodes that amino acid in that host. The maize preferred codon, for example, for a particular amino acid may be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is found in Murray *et al.* (1989), Nucleic Acids Research 17:477-498, the disclosure of which is incorporated herein by reference. Synthetic genes can also be made based on the distribution of codons a particular host uses for a particular amino acid.

In this manner, the nucleotide sequences can be optimized for expression in any plant. It is recognized that all or any part of the gene sequence may be optimized or synthetic. That is, synthetic or partially optimized sequences may also be used.

In like manner, the nucleotide sequences can be optimized for expression in any microorganism. For *Bacillus* preferred codon usage, see, for example US Patent No. 5,024,837 and Johansen *et al.* (1988) Gene 65:293-304.

Methodologies for the construction of plant expression cassettes as well as the introduction of foreign DNA into plants are described in the art. Such expression cassettes may include promoters, terminators, enhancers, leader sequences, introns and other regulatory sequences operably linked to the pesticidal protein coding sequence. It is further recognized that promoters or terminators of the VIP genes can be used in expression cassettes.

Generally, for the introduction of foreign DNA into plants Ti plasmid vectors have been utilized for the delivery of foreign DNA as well as direct DNA uptake, liposomes, electroporation, micro-injection, and the use of microprojectiles. Such methods had been published in the art. See, for example, Guerche *et al.*, (1987) Plant Science 52:111-116; Neuhause *et al.*, (1987) Theor. Appl. Genet. 75:30-36; Klein *et al.*, (1987) Nature 327: 70-73; Howell *et al.*, (1980) Science 208:1265; Horsch *et al.*, (1985) Science 227: 1229-1231; DeBlock *et al.*, (1989) Plant Physiology 91:694-701; Methods for Plant Molecular Biology (Weissbach and Weissbach, eds.) Academic Press, Inc. (1988); and Methods in Plant Molecular Biology (Schuler and Zielinski,

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eds.) Academic Press, Inc. (1989). See also US patent application serial no. 08/008,374 herein incorporated by reference. See also, EP-A 0193259 and EP-A 0451878. It is understood that the method of transformation will depend upon the plant cell to be transformed.

It is further recognized that the components of the expression cassette may be modified to increase expression. For example, truncated sequences, nucleotide substitutions or other modifications may be employed. See, for example Perlak *et al.* (1991) Proc. Natl. Acad. Sci. USA 88:3324-3328; Murray *et al.*, (1989) Nucleic Acids Research 17:477-498; and WO 91/16432.

The construct may also include any other necessary regulators such as terminators, (Guerineau *et al.*, (1991), Mol. Gen. Genet., 226:141-144; Proudfoot, (1991), Cell, 64:671-674; Sanfacon *et al.*, (1991), Genes Dev., 5:141-149; Mogen *et al.*, (1990), Plant Cell, 2:1261-1272; Munroe *et al.*, (1990), Gene, 91:151-158; Ballas *et al et al.*, (1989), Nucleic Acids Res., 17:7891-7903; Joshi *et al.*, (1987), Nucleic Acid Res., 15:9627-9639); plant translational consensus sequences (Joshi, C.P., (1987), Nucleic Acids Research, 15:6643-6653), introns (Luehrs and Walbot, (1991), Mol. Gen. Genet., 225:81-93) and the like, operably linked to the nucleotide sequence. It may be beneficial to include 5' leader sequences in the expression cassette construct. Such leader sequences can act to enhance translation.

Translational leaders are known in the art and include:

Picornavirus leaders, for example, EMCV leader (encephalomyocarditis 5' noncoding region) (Elroy-Stein, O., Fuerst, T.R., and Moss, B. (1989) PNAS USA 86:6126-6130);

Potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison *et al.*, (1986); MDMV leader (Maize Dwarf Mosaic Virus); Virology, 154:9-20), and

Human immunoglobulin heavy-chain binding protein (BiP). (Macejak, D.G., and Sarnow, P., (1991), Nature, 353:90-94;

Untranslated leader from the coat protein mRNA of alfalfa mosaic virus (AMV RNA 4), (Jobling, S.A., and Gehrke, L., (1987), Nature, 325:622-625;

Tobacco mosaic virus leader (TMV), (Gallie, D.R. *et al.*, (1989), Molecular Biology of RNA, pages 237-256; and

Maize Chlorotic Mottle Virus leader (MCMV) (Lommel, S.A. *et al.*, (1991), Virology, 81:382-385. See also, Della-Cioppa *et al.*, (1987), Plant Physiology, 84:965-968.

A plant terminator may be utilized in the expression cassette. See, Rosenberg *et al.*, (1987), Gene, 56:125; Guerineau *et al.*, (1991), Mol. Gen. Genet., 226:141-144; Proudfoot, (1991), Cell, 64:671-674; Sanfacon *et al.*, (1991), Genes Dev., 5:141-149; Mogen *et al.*, (1990), Plant Cell, 2:1261-1272; Munroe *et al.*, (1990), Gene, 91:151-158; Ballas *et al.*, (1989), Nucleic Acids Res., 17:7891-7903; Joshi *et al.*, (1987), Nucleic Acid Res., 15:9627-9639.

For tissue specific expression, the nucleotide sequences of the invention can be operably linked to tissue specific promoters. See, for example, EP-A 0618976, herein incorporated by reference.

Further comprised within the scope of the present invention are transgenic plants, in particular transgenic fertile plants transformed by means of the aforescribed processes and their asexual and/or sexual progeny, which comprise and preferably also express the pesticidal protein according to the invention. Especially preferred are hybrid plants.

The transgenic plant according to the invention may be a dicotyledonous or a monocotyledonous plant. Preferred are monocotyledonous plants of the *Graminaceae* family involving Lolium, Zea, Triticum, Triticale, Sorghum, Saccharum, Bromus, Oryzae, Avena, Hordeum, Secale and Setaria plants.

Especially preferred are transgenic maize, wheat, barley, sorghum, rye, oats, turf grasses and rice.

Among the dicotyledonous plants soybean, cotton, tobacco, sugar beet, oilseed rape, and sunflower are especially preferred herein.

The expression 'progeny' is understood to embrace both, "asexually" and "sexually" generated progeny of transgenic plants. This definition is also meant to include all mutants and variants obtainable by means of known processes, such as for example cell fusion or mutant selection and which still exhibit the characteristic properties of the initially transformed parent plant, together with all crossing and fusion products of the transformed plant material.

Another object of the invention concerns the proliferation material of transgenic plants.

The proliferation material of transgenic plants is defined relative to the invention as any plant material that may be propagated sexually or asexually *in vivo* or *in vitro*. Particularly preferred within the scope of the present invention are protoplasts, cells,

calli, tissues, organs, seeds, embryos, pollen, egg cells, zygotes, together with any other propagating material obtained from transgenic plants.

Parts of plants, such as for example flowers, stems, fruits, leaves, roots originating in transgenic plants or their progeny previously transformed by means of the process of the invention and therefore consisting at least in part of transgenic cells, are also an object of the present invention.

Before the plant propagation material [fruit, tuber, grains, seed], but especially seed is sold as a commercial product, it is customarily treated with a protectant coating comprising herbicides, insecticides, fungicides, bactericides, nematicides, molluscicides or mixtures of several of these preparations, if desired together with further carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation to provide protection against damage caused by bacterial, fungal or animal pests.

In order to treat the seed, the protectant coating may be applied to the seeds either by impregnating the tubers or grains with a liquid formulation or by coating them with a combined wet or dry formulation. In addition, in special cases, other methods of application to plants are possible, eg treatment directed at the buds or the fruit.

The plant seed according to the invention comprising a DNA molecule comprising a nucleotide sequence encoding a pesticidal protein according to the invention may be treated with a seed protectant coating comprising a seed treatment compound, such as, for example, captan, carboxin, thiram (TMTD[®]), methalaxyl (Apron[®]) and pirimiphos-methyl (Actellic[®]) and others that are commonly used in seed treatment. Preferred within the scope of the invention are seed protectant coatings comprising an entomocidal composition according to the invention alone or in combination with one of the a seed protectant coating customarily used in seed treatment.

It is thus a further object of the present invention to provide plant propagation material for cultivated plants, but especially plant seed that is treated with a seed protectant coating as defined hereinbefore.

It is recognized that the genes encoding the pesticidal proteins can be used to transform insect pathogenic organisms. Such organisms include Baculoviruses, fungi, protozoa, bacteria and nematodes.

The *Bacillus* strains of the invention may be used for protecting agricultural crops and products from pests. Alternatively, a gene encoding the pesticide may be

introduced via a suitable vector into a microbial host, and said host applied to the environment or plants or animals. Microorganism hosts may be selected which are known to occupy the "phytosphere" (phyllloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

Such microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylius*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconostoc*, and *Alcaligenes*; fungi, particularly yeast, e.g., *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodotorula*, and *Aureobasidium*. Of particular interest are such phytosphere bacterial species as *Pseudomonas syringae*, *Pseudomonas fluorescens*, *Serratia marcescens*, *Acetobacter xylinum*, *Agrobacteria*, *Rhodopseudomonas sphaeroides*, *Xanthomonas campestris*, *Rhizobium meliloti*, *Alcaligenes entrophus*, *Clavibacter xyli* and *Azotobacter vinlandii*, and phytosphere yeast species such as *Rhodotorula rubra*, *R. glutinis*, *R. marina*, *R. aurantiaca*, *Cryptococcus albidus*, *C. diffluens*, *C. laurentii*, *Saccharomyces rosei*, *S. pretoriensis*, *S. cerevisiae*, *Sporobolomyces rosae*, *S. odorus*, *Kluyveromyces veronae*, and *Aureobasidium pollulans*. Of particular interest are the pigmented microorganisms.

A number of ways are available for introducing a gene expressing the pesticidal protein into the microorganism host under conditions which allow for stable maintenance and expression of the gene. For example, expression cassettes can be constructed which include the DNA constructs of interest operably linked with the transcriptional and translational regulatory signals for expression of the DNA constructs, and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system which is functional in the host, whereby integration or stable maintenance will occur.

Transcriptional and translational regulatory signals include but are not limited to promoter, transcriptional initiation start site, operators, activators, enhancers, other regulatory elements, ribosomal binding sites, an initiation codon, termination signals,

and the like. See, for example, US Patent 5,039,523; US Patent No. 4,853,331; EPO 0480762A2; Sambrook *et al.* supra; Molecular Cloning, a Laboratory Manual, Maniatis *et al.* (eds) Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982); Advanced Bacterial Genetics, Davis *et al.* (eds.) Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1980); and the references cited therein.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of the target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and -positive, include *Enterobacteriaceae*, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; *Bacillaceae*; *Rhizobiceae*, such as *Rhizobium*; *Spirillaceae*, such as *photobacterium*, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*, *Desulfovibrio*, *Spirillum*; *Lactobacillaceae*; *Pseudomonadaceae*, such as *Pseudomonas* and *Acetobacter*; *Azotobacteraceae* and *Nitrobacteraceae*. Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Saccharomyces* and *Schizosaccharomyces*; and *Basidiomycetes* yeast, such as *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, and the like.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the protein gene into the host, availability of expression systems, efficiency of expression, stability of the protein in the host, and the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as *Rhodotorula* sp., *Aureobasidium* sp., *Saccharomyces* sp., and *Sporobolomyces* sp.; phylloplane

organisms such as *Pseudomonas sp.*, *Erwinia sp.* and *Flavobacterium sp.*; or such other organisms as *Escherichia*, *LactoBacillus sp.*, *Bacillus sp.*, and the like. Specific organisms include *Pseudomonas aeurginosa*, *Pseudomonas fluorescens*, *Saccharomyces cerevisiae*, *Bacillus thuringiensis*, *Escherichia coli*, *Bacillus subtilis*, and the like.

VIP genes can be introduced into micro-organisms that multiply on plants (epiphytes) to deliver VIP proteins to potential target pests. Epiphytes can be gram-positive or gram-negative bacteria for example.

Root colonizing bacteria, for example, can be isolated from the plant of interest by methods known in the art. Specifically, a *Bacillus cereus* strain which colonizes roots could be isolated from roots of a plant (for example see J. Handelsman, S. Raffel, E. Mester, L. Wunderlich and C. Grau, Appl. Environ. Microbiol. 56:713-718, (1990)). VIP1 and/or VIP2 and/or VIP3 could be introduced into a root colonizing *Bacillus cereus* by standard methods known in the art.

Specifically, VIP1 and/or VIP2 derived from *Bacillus cereus* strain AB78 can be introduced into a root colonizing *Bacillus cereus* by means of conjugation using standard methods (J. Gonzalez, B. Brown and B. Carlton, Proc. Natl. Acad. Sci. 79:6951-6955, (1982)).

Also, VIP1 and/or VIP2 and/or VIP3 or other VIPs of the invention can be introduced into the root colonizing *Bacillus* by means of electro-transformation. Specifically, VIPs can be cloned into a shuttle vector, for example, pHT3101 (D. Lereclus et al., FEMS Microbiol. Letts., 60:211-218 (1989)) as described in Example 10. The shuttle vector pHT3101 containing the coding sequence for the particular VIP can then be transformed into the root colonizing *Bacillus* by means of electroporation (D. Lereclus et al. 1989, FEMS Microbiol. Letts. 60:211-218).

Expression systems can be designed so that VIP proteins are secreted outside the cytoplasm of gram negative bacteria, *E. coli*, for example. Advantages of having VIP proteins secreted are (1) it avoids potential toxic effects of VIP proteins expressed within the cytoplasm and (2) it can increase the level of VIP protein expressed and (3) can aid in efficient purification of VIP protein.

VIP proteins can be made to be secreted in *E. coli*, for example, by fusing an appropriate *E. coli* signal peptide to the amino-terminal end of the VIP signal peptide or replacing the VIP signal peptide with the *E. coli* signal peptide. Signal peptides

recognized by *E. coli* can be found in proteins already known to be secreted in *E. coli*, for example the OmpA protein (J. Ghrayeb, H. Kimura, M. Takahara, Y. Masui and M. Inouye, EMBO J., 3:2437-2442 (1984)). OmpA is a major protein of the *E. coli* outer membrane and thus its signal peptide is thought to be efficient in the translocation process. Also, the OmpA signal peptide does not need to be modified before processing as may be the case for other signal peptides, for example lipoprotein signal peptide

(G. Duffaud, P. March and M. Inouye, Methods in Enzymology, 153:492 (1987)).

Specifically, unique BamHI restriction sites can be introduced at the amino-terminal and carboxy-terminal ends of the VIP coding sequences using standard methods known in the art. These BamHI fragments can be cloned, in frame, into the vector pIN-III-ompA1, A2 or A3 (J. Ghrayeb, H. Kimura, M. Takahara, H. Hsiung, Y. Masui and M. Inouye, EMBO J., 3:2437-2442 (1984)) thereby creating ompA:VIP fusion gene which is secreted into the periplasmic space. The other restriction sites in the polylinker of pIN-III-ompA can be eliminated by standard methods known in the art so that the VIP amino-terminal amino acid coding sequence is directly after the ompA signal peptide cleavage site. Thus, the secreted VIP sequence in *E. coli* would then be identical to the native VIP sequence.

When the VIP native signal peptide is not needed for proper folding of the mature protein, such signal sequences can be removed and replaced with the ompA signal sequence. Unique BamHI restriction sites can be introduced at the amino-termini of the proprotein coding sequences directly after the signal peptide coding sequences of VIP and at the carboxy-termini of VIP coding sequence. These BamHI fragments can then be cloned into the pIN-III-ompA vectors as described above.

General methods for employing the strains of the invention in pesticide control or in engineering other organisms as pesticidal agents are known in the art. See, for example US Patent No. 5,039,523 and EP 0480762A2.

VIPs can be fermented in a bacterial host and the resulting bacteria processed and used as a microbial spray in the same manner that *Bacillus thuringiensis* strains have been used as insecticidal sprays. In the case of a VIP(s) which is secreted from *Bacillus*, the secretion signal is removed or mutated using procedures known in the art. Such mutations and/or deletions prevent secretion of the VIP protein(s) into the growth medium during the fermentation process. The VIPs are retained within the cell

and the cells are then processed to yield the encapsulated VIPs. Any suitable microorganism can be used for this purpose. *Psuedomonas* has been used to express *Bacillus thuringiensis* endotoxins as encapsulated proteins and the resulting cells processed and sprayed as an insecticide. (H. Gaertner *et al.* 1993, In Advanced Engineered Pesticides, L. Kim ed.)

Various strains of *Bacillus thuringiensis* are used in this manner. Such *Bt* strains produce endotoxin protein(s) as well as VIPs. Alternatively, such strains can produce only VIPs. A sporulation deficient strain of *Bacillus subtilis* has been shown to produce high levels of the CryIIIA endotoxin from *Bacillus thuringiensis* (Agaisse, H. and Lereclus, D., "Expression in *Bacillus subtilis* of the *Bacillus thuringiensis* CryIIIA toxin gene is not dependent on a sporulation-specific sigma factor and is increased in a *spoOA* mutant", *J. Bacteriol.*, 176:4734-4741 (1994)). A similar *spoOA* mutant can be prepared in *Bacillus thuringiensis* and used to produce encapsulated VIPs which are not secreted into the medium but are retained within the cell.

To have VIPs maintained within the *Bacillus* cell the signal peptide can be disarmed so that it no longer functions as a secretion signal. Specifically, the putative signal peptide for VIP1 encompasses the first 31 amino acids of the protein with the putative consensus cleavage site, Ala-X-Ala, at the C-terminal portion of this sequence (G. von Heijne , *J. Mol. Biol.* 184:99-105 (1989)) and the putative signal peptide for VIP2 encompasses the first 40 amino acids of the protein with the putative cleavage site after Ala40. The cleavage sites in either VIP1 or VIP2 can be mutated with methods known in the art to replace the cleavage site consensus sequence with alternative amino acids that are not recognized by the signal peptidases.

Alternatively, the signal peptides of VIP1, VIP2 and/or other VIPs of the invention can be eliminated from the sequence thereby making them unrecognizable as secretion proteins in *Bacillus*. Specifically, a methionine start site can be engineered in front of the proprotein sequence in VIP1, starting at Asp32, or the proprotein sequence in VIP2, starting at Glu41 using methods known in the art.

VIP genes can be introduced into micro-organisms that multiply on plants (epiphytes) to deliver VIP proteins to potential target pests. Epiphytes can be gram-positive or gram-negative bacteria for example.

The *Bacillus* strains of the invention or the microorganisms which have been genetically altered to contain the pesticidal gene and protein may be used for

protecting agricultural crops and products from pests. In one aspect of the invention, whole, i.e., unlysed, cells of a toxin (pesticide)-producing organism are treated with reagents that prolong the activity of the toxin produced in the cell when the cell is applied to the environment of target pest(s).

Alternatively, the pesticides are produced by introducing a heterologous gene into a cellular host. Expression of the heterologous gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. These cells are then treated under conditions that prolong the activity of the toxin produced in the cell when the cell is applied to the environment of target pest(s). The resulting product retains the toxicity of the toxin. These naturally encapsulated pesticides may then be formulated in accordance with conventional techniques for application to the environment hosting a target pest, e.g., soil, water, and foliage of plants. See, for example EPA 0192319, and the references cited therein.

The active ingredients of the present invention are normally applied in the form of compositions and can be applied to the crop area or plant to be treated, simultaneously or in succession, with other compounds. These compounds can be both fertilizers or micronutrient donors or other preparations that influence plant growth. They can also be selective herbicides, insecticides, fungicides, bactericides, nematicides, mollusicides or mixtures of several of these preparations, if desired, together with further agriculturally acceptable carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formulation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers.

Preferred methods of applying an active ingredient of the present invention or an agrochemical composition of the present invention which contains at least one of the insect-specific proteins produced by the bacterial strains of the present invention are leaf application, seed coating and soil application. The number of applications and the rate of application depend on the intensity of infestation by the corresponding pest.

The present invention thus further provides an entomocidal composition comprising as an active ingredient at least one of the novel insect-specific proteins

according to the invention and/or a recombinant microorganism containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, but especially a recombinant *Bacillus spp* strain, such as *Bacillus cereus* or *Bacillus thuringiensis*, containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, or a derivative or mutant thereof, together with an agricultural adjuvant such as a carrier, diluent, surfactant or application-promoting adjuvant. The composition may also contain a further biologically active compound. The said compound can be both a fertilizer or micronutrient donor or other preparations that influence plant growth. It can also be a selective herbicide, insecticide, fungicide, bactericide, nematicide, molluscide or mixtures of several of these preparations, if desired, together with further agriculturally acceptable carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formulation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers.

The composition may comprise from 0.1 to 99% by weight of the active ingredient, from 1 to 99.9% by weight of a solid or liquid adjuvant, and from 0 to 25% by weight of a surfactant. The active ingredient comprising at least one of the novel insect-specific proteins according to the invention or a recombinant microorganism containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, but especially a recombinant *Bacillus spp* strain, such as *Bacillus cereus* or *Bacillus thuringiensis* strain containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, or a derivative or mutant thereof, or the composition containing the said active ingredient, may be administered to the plants or crops to be protected together with certain other insecticides or chemicals (1993 Crop Protection Chemicals Reference, Chemical and Pharmaceutical Press, Canada) without loss of potency. It is compatible with most other commonly used agricultural spray materials but should not be used in extremely alkaline spray solutions. It may be administered as a dust, a suspension, a wettable powder or in any other material form suitable for agricultural application.

The invention further provides methods for controlling or inhibiting of insect pests by applying an active ingredient comprising at least one of the novel insect-specific proteins according to the invention or a recombinant microorganism containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form or a composition comprising the said active ingredient to (a) an environment in which the insect pest may occur, (b) a plant or plant part in order to protect said plant or plant part from damage caused by an insect pest, or (c) seed in order to protect a plant which develops from said seed from damage caused by an insect pest.

A preferred method of application in the area of plant protection is application to the foliage of the plants (foliar application), with the number of applications and the rate of application depending on the plant to be protected and the risk of infestation by the pest in question. However, the active ingredient may also penetrate the plants through the roots (systemic action) if the locus of the plants is impregnated with a liquid formulation or if the active ingredient is incorporated in solid form into the locus of the plants, for example into the soil, e.g. in granular form (soil application). In paddy rice crops, such granules may be applied in metered amounts to the flooded rice field.

The compositions according to the invention are also suitable for protecting plant propagating material, e.g. seed, such as fruit, tubers or grains, or plant cuttings, from insect pests. The propagation material can be treated with the formulation before planting: seed, for example, can be dressed before being sown. The active ingredient of the invention can also be applied to grains (coating), either by impregnating the grains with a liquid formulation or by coating them with a solid formulation. The formulation can also be applied to the planting site when the propagating material is being planted, for example to the seed furrow during sowing. The invention relates also to those methods of treating plant propagation material and to the plant propagation material thus treated.

The compositions according to the invention comprising as an active ingredient a recombinant microorganism containing at least one of the novel toxin genes in recombinant form, but especially a recombinant *Bacillus spp* strain, such as *Bacillus cereus* or *Bacillus thuringiensis* strain containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, or a derivative or mutant thereof may be applied in any method

known for treatment of seed or soil with bacterial strains. For example, see US Patent No.4,863,866. The strains are effective for biocontrol even if the microorganism is not living. Preferred is, however, the application of the living microorganism.

Target crops to be protected within the scope of the present invention comprise, e.g., the following species of plants:

cereals (wheat, barley, rye, oats, rice, sorghum and related crops), beet (sugar beet and fodder beet), forage grasses (orchardgrass, fescue, and the like), drupes, pomes and soft fruit (apples, pears, plums, peaches, almonds, cherries, strawberries, raspberries and blackberries), leguminous plants (beans, lentils, peas, soybeans), oil plants (rape, mustard, poppy, olives, sunflowers, coconuts, castor oil plants, cocoa beans, groundnuts), cucumber plants (cucumber, marrows, melons) fiber plants (cotton, flax, hemp, jute), citrus fruit (oranges, lemons, grapefruit, mandarins), vegetables (spinach, lettuce, asparagus, cabbages and other Brassicae, onions, tomatoes, potatoes, paprika), lauraceae (avocados, carrots, cinnamon, camphor), deciduous trees and conifers (e.g. linden-trees, yew-trees, oak-trees, alders, poplars, birch-trees, firs, larches, pines), or plants such as maize, tobacco, nuts, coffee, sugar cane, tea, vines, hops, bananas and natural rubber plants, as well as ornamentals (including composites).

A recombinant *Bacillus spp* strain, such as *Bacillus cereus* or *Bacillus thuringiensis* strain, containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form is normally applied in the form of entomocidal compositions and can be applied to the crop area or plant to be treated, simultaneously or in succession, with further biologically active compounds. These compounds may be both fertilizers or micronutrient donors or other preparations that influence plant growth. They may also be selective herbicides, insecticides, fungicides, bactericides, nematicides, molluscicides or mixtures of several of these preparations, if desired together with further carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation.

The active ingredient according to the invention may be used in unmodified form or together with any suitable agriculturally acceptable carrier. Such carriers are adjuvants conventionally employed in the art of agricultural formulation, and are therefore formulated in known manner to emulsifiable concentrates, coatable pastes, directly sprayable or dilutable solutions, dilute emulsions, wettable powders, soluble powders,

dusts, granulates, and also encapsulations, for example, in polymer substances. Like the nature of the compositions, the methods of application, such as spraying, atomizing, dusting, scattering or pouring, are chosen in accordance with the intended objective and the prevailing circumstances. Advantageous rates of application are normally from about 50 g to about 5 kg of active ingredient (a.i.) per hectare ("ha", approximately 2.471 acres), preferably from about 100 g to about 2kg a.i./ha. Important rates of application are about 200 g to about 1kg a.i./ha and 200g to 500g a.i./ha.

For seed dressing advantageous application rates are 0.5 g to 1000 g a.i.per 100 kg seed, preferably 3 g to 100 g a.i. per 100 kg seed or 10 g to 50 g a.i.per 100 kg seed.

Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formulation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers. The formulations, i.e. the entomocidal compositions, preparations or mixtures containing the recombinant *Bacillus spp strain*, such as *Bacillus cereus* or *Bacillus thuringiensis* strain containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form as an active ingredient or combinations thereof with other active ingredients, and, where appropriate, a solid or liquid adjuvant, are prepared in known manner, e.g., by homogeneously mixing and/or grinding the active ingredients with extenders, e.g., solvents, solid carriers, and in some cases surface-active compounds (surfactants).

Suitable solvents are: aromatic hydrocarbons, preferably the fractions containing 8 to 12 carbon atoms, e.g. xylene mixtures or substituted naphthalenes, phthalates such as dibutyl phthalate or dioctyl phthalate, aliphatic hydrocarbons such as cyclohexane or paraffins, alcohols and glycols and their ethers and esters, such as ethanol, ethylene glycol monomethyl or monoethyl ether, ketones such as cyclohexanone, strongly polar solvents such as N-methyl-2-pyrrolidone, dimethylsulfoxide or dimethylformamide, as well as vegetable oils or epoxidised vegetable oils such as epoxidised coconut oil or soybean oil; or water.

The solid carriers used, e.g., for dusts and dispersible powders, are normally natural mineral fillers such as calcite, talcum, kaolin, montmorillonite or attapulgite. In order to improve the physical properties it is also possible to add highly dispersed silicic acid or highly dispersed absorbent polymers. Suitable granulated adsorptive

carriers are porous types, for example pumice, broken brick, sepiolite or bentonite; and suitable nonsorbent carriers are materials such as calcite or sand. In addition, a great number of pregranulated materials of inorganic or organic nature can be used, e.g. especially dolomite or pulverized plant residues.

Depending on the nature of the active ingredients to be formulated, suitable surface-active compounds are non-ionic, cationic and/or anionic surfactants having good emulsifying, dispersing and wetting properties. The term "surfactants" will also be understood as comprising mixtures of surfactants. Suitable anionic surfactants can be both water-soluble soaps and water-soluble synthetic surface-active compounds. Suitable soaps are the alkali metal salts, alkaline earth metal salts or unsubstituted or substituted ammonium salts of higher fatty acids (C_{10} - C_{22}), e.g. the sodium or potassium salts of oleic or stearic acid, or of natural fatty acid mixtures which can be obtained, e.g. from coconut oil or tallow oil. Further suitable surfactants are also the fatty acid methyltaurin salts as well as modified and unmodified phospholipids.

More frequently, however, so-called synthetic surfactants are used, especially fatty sulfonates, fatty sulfates, sulfonated benzimidazole derivatives or alkylarylsulfonates. The fatty sulfonates or sulfates are usually in the forms of alkali metal salts, alkaline earth metal salts or unsubstituted or substituted ammonium salts and generally contain a C_8 - C_{22} alkyl radical which also includes the alkyl moiety of acyl radicals, e.g. the sodium or calcium salt of lignosulfonic acid, of dodecylsulfate, or of a mixture of fatty alcohol sulfates obtained from natural fatty acids. These compounds also comprise the salts of sulfuric acid esters and sulfonic acids of fatty alcohol/ethylene oxide adducts. The sulfonated benzimidazole derivatives preferably contain 2 sulfonic acid groups and one fatty acid radical containing about 8 to 22 carbon atoms. Examples of alkylarylsulfonates are the sodium, calcium or triethanolamine salts of dodecylbenzenesulfonic acid, dibutylnaphthalenesulfonic acid, or of a naphthalenesulfonic acid/formaldehyde condensation product. Also suitable are corresponding phosphates, e.g. salts of the phosphoric acid ester of an adduct of *p*-nonylphenol with 4 to 14 moles of ethylene oxide.

Non-ionic surfactant are preferably polyglycol ether derivatives of aliphatic or cycloaliphatic alcohols, or saturated or unsaturated fatty acids and alkylphenols, said derivatives containing 3 to 30 glycol ether groups and 8 to 20 carbon atoms in the

(aliphatic) hydrocarbon moiety and 6 to 18 carbon atoms in the alkyl moiety of the alkylphenols.

Further suitable non-ionic surfactants are the water-soluble adducts of polyethylene oxide with polypropylene glycol, ethylenediaminopolypropylene glycol and alkylpolypropylene glycol containing 1 to 10 carbon atoms in the alkyl chain, which adducts contain 20 to 250 ethylene glycol ether groups and 10 to 100 propylene glycol ether groups. These compounds usually contain 1 to 5 ethylene glycol units per propylene glycol unit. Representative examples of non-ionic surfactants are nonylphenolpolyethoxyethanols, castor oil polyglycer ethers, polypropylene/polyethylene oxide adducts, tributylphenoxypropoxyethanol, polyethylene glycol and octylphenoxypropoxyethanol. Fatty acid esters of polyoxyethylene sorbitan, such as polyoxyethylene sorbitan trioleate, are also suitable non-ionic surfactants.

Cationic surfactants are preferably quaternary ammonium salts which contain, as N-substituent, at least one C₆-C₂₂ alkyl radical and, as further substituents, lower unsubstituted or halogenated alkyl, benzyl or hydroxyl-lower alkyl radicals. The salts are preferably in the form of halides, methylsulfates or ethylsulfates, e.g., stearyltrimethylammonium chloride or benzyl-di-(2-chloroethyl)ethylammonium bromide.

The surfactants customarily employed in the art of formulation are described, e.g., in "McCutcheon's Detergents and Emulsifiers Annual", MC Publishing Corp. Ridgewood, N.J., 1979; Dr. Helmut Stache, "Tensid Taschenbuch" (Handbook of Surfactants), Carl Hanser Verlag, Munich/Vienna.

Another particularly preferred characteristic of an entomocidal composition of the present invention is the persistence of the active ingredient when applied to plants and soil. Possible causes for loss of activity include inactivation by ultra-violet light, heat, leaf exudates and pH. For example, at high pH, particularly in the presence of reductant, δ-endotoxin crystals are solubilized and thus become more accessible to proteolytic inactivation. High leaf pH might also be important, particularly where the leaf surface can be in the range of pH 8-10. Formulation of an entomocidal composition of the present invention can address these problems by either including additives to help prevent loss of the active ingredient or encapsulating the material in such a way that the active ingredient is protected from inactivation. Encapsulation

can be accomplished chemically (McGuire and Shasha, J Econ Entomol 85: 1425-1433, 1992) or biologically (Barnes and Cummings, 1986; EP-A 0 192 319). Chemical encapsulation involves a process in which the active ingredient is coated with a polymer while biological encapsulation involves the expression of the δ -endotoxin genes in a microbe. For biological encapsulation, the intact microbe containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form is used as the active ingredient in the formulation. The addition of UV protectants might effectively reduce irradiation damage. Inactivation due to heat could also be controlled by including an appropriate additive.

Preferred within the present application are formulations comprising living microorganisms as active ingredient either in form of the vegetative cell or more preferable in form of spores, if available. Suitable formulations may consist, for example, of polymer gels which are crosslinked with polyvalent cations and comprise these microorganisms. This is described, for example, by D.R. Fravel et al. in Phytopathology, Vol. 75, No. 7, 774-777, 1985 for alginate as the polymer material. It is also known from this publication that carrier materials can be co-used. These formulations are as a rule prepared by mixing solutions of naturally occurring or synthetic gel-forming polymers, for example alginates, and aqueous salt solutions of polyvalent metal ions such that individual droplets form, it being possible for the microorganisms to be suspended in one of the two or in both reaction solutions. Gel formation starts with the mixing in drop form. Subsequent drying of these gel particles is possible. This process is called ionotropic gelling. Depending on the degree of drying, compact and hard particles of polymers which are structurally crosslinked via polyvalent cations and comprise the microorganisms and a carrier present predominantly uniformly distributed are formed. The size of the particles can be up to 5 mm.

Compositions based on partly crosslinked polysaccharides which, in addition to a microorganism, for example, can also comprise finely divided silicic acid as the carrier material, crosslinking taking place, for example, via Ca^{++} ions, are described in EP-A1-0 097 571. The compositions have a water activity of not more than 0.3. W.J. Cornick et al. describe in a review article [New Directions in Biological Control: Alternatives for Suppressing Agricultural Pests and Diseases, pages 345-372, Alan R.

Liss, Inc. (1990)] various formulation systems, granules with vermiculite as the carrier and compact alginate beads prepared by the ionotropic gelling process being mentioned. Such compositions are also disclosed by D.R.Fravel in Pesticide Formulations and Application Systems: 11th Volume, ASTM STP 1112 American Society for Testing and Materials, Philadelphia, 1992, pages 173 to 179 and can be used to formulate the recombinant microorganisms according to the invention.

The entomocidal compositions of the invention usually contain from about 0.1 to about 99%, preferably about 0.1 to about 95%, and most preferably from about 3 to about 90% of the active ingredient, from about 1 to about 99.9%, preferably from about 1 to about 99%, and most preferably from about 5 to about 95% of a solid or liquid adjuvant, and from about 0 to about 25%, preferably about 0.1 to about 25%, and most preferably from about 0.1 to about 20% of a surfactant.

In a preferred embodiment of the invention the entomocidal compositions usually contain 0.1 to 99%, preferably 0.1 to 95%, of a recombinant *Bacillus spp* strain, such as *Bacillus cereus* or *Bacillus thuringiensis* strain containing at least one DNA molecule comprising a nucleotide sequence encoding the novel insect-specific proteins in recombinant form, or combination thereof with other active ingredients, 1 to 99.9% of a solid or liquid adjuvant, and 0 to 25%, preferably 0.1 to 20%, of a surfactant.

Whereas commercial products are preferably formulated as concentrates, the end user will normally employ dilute formulations of substantially lower concentration. The entomocidal compositions may also contain further ingredients, such as stabilizers, antifoams, viscosity regulators, binders, tackifiers as well as fertilizers or other active ingredients in order to obtain special effects.

In one embodiment of the invention a *Bacillus cereus* microorganism has been isolated which is capable of killing *Diabrotica virgifera virgifera*, and *Diabrotica longicornis barberi*. The novel *B. cereus* strain AB78 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, IL 61604, USA and given Accession No. NRRL B-21058.

A fraction protein has been substantially purified from the *B. cereus* strain. This purification of the protein has been verified by SDS-PAGE and biological activity. The

protein has a molecular weight of about 60 to about 100 kDa, particularly about 70 to about 90 kDa, more particularly about 80 kDa, hereinafter VIP.

Amino-terminal sequencing has revealed the N-terminal amino-acid sequence to be:

NH₂-Lys-Arg-Glu-Ile-Asp-Glu-Asp-Thr-Asp-Thr-Asx-Gly-Asp-Ser-Ile-Pro-(SEQ ID NO:8) where Asx represents either Asp or Asn. The entire amino acid sequence is given in SEQ ID NO:7. The DNA sequence which encodes the amino acid sequence of SEQ ID NO:7 is disclosed in SEQ ID NO:6.

An oligonucleotide probe for the region of the gene encoding amino acids 3-9 of the NH₂-terminus has been generated. The probe was synthesized based on the codon usage of a *Bacillus thuringiensis* (Bt) δ-endotoxin gene. The nucleotide sequence of the oligonucleotide probe used for Southern hybridizations was as follows:

5'- GAA ATT GAT CAA GAT ACN GAT -3' (SEQ ID NO:9)

where N represents any base.

In addition, the DNA probe for the Bc AB78 VIP1 gene described herein, permits the screening of any *Bacillus* strain or other organisms to determine whether the VIP1 gene (or related gene) is naturally present or whether a particular transformed organism includes the VIP1 gene.

The invention now being generally described, the same will be better understood by reference to the following detailed examples that are provided for the purpose of illustration and are not to be considered limiting of the invention unless so specified.

A standard nomenclature has been developed based on the sequence identity of the proteins encompassed by the present invention. The gene and protein names for the detailed examples which follow and their relationship to the names used in the parent application [US application serial no 314594/08] are shown below.

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<i>Gene / Protein</i>	<i>Gene / Protein</i>	<i>Description of Protein</i>
<i>Name under Standard Nomenclature</i>	<i>Name in Parent</i>	
VIP1A(a)	VIP1	VIP1 from strain AB78 as disclosed in SEQ ID NO:5.
VIP2A(a)	VIP2	VIP2 from strain AB78 as disclosed in SEQ ID NO:2.
VIP1A(b)	VIP1 homolog	VIP1 from <i>Bacillus thuringiensis</i> var. <i>tenebrionis</i> as disclosed in SEQ ID NO:21.
VIP2A(b)	VIP2 homolog	VIP2 from <i>Bacillus thuringiensis</i> var. <i>tenebrionis</i> as disclosed in SEQ ID NO:20.
VIP3A(a)	--	VIP from strain AB88 as disclosed in SEQ ID NO:28 of the present application
VIP3A(b)	--	VIP from strain AB424 as disclosed in SEQ ID NO:31 of the present application

EXPERIMENTALFormulation Examples

The active ingredient used in the following formulation examples are *Bacillus cereus* strain AB78 having Accession No. NRRL B-21058; *Bacillus thuringiensis* strains having Accession Nos. NRRL B-21060, NRRL B-21224, NRRL B-21225, NRRL B-21226, NRRL B-21227, and NRRL B-21439; and *Bacillus spp* strains having Accession Nos NRRL B-21228, NRRL B-21229, and NRRL B-21230. All the mentioned strains are natural isolates comprising the insect-specific proteins according to the invention.

Alternatively, the isolated insect-specific proteins are used as the active ingredient alone or in combination with the above-mentioned *Bacillus* strains.

A1. Wettable powders

	a)	b)	c)
<i>Bacillus thuringiensis</i> spores	25%	50%	75%
sodium lignosulfonate	5%	5%	--
sodium laurylsulfate	3%	--	5%
sodium diisobutylnaphthalenesulfonate	--	6%	10%
octylphenol polyethylene glycol ether (7-8 moles of ethylene oxide)	--	2%	--
highly dispersed silicid acid	5%	10%	10%
kaolin	62%	27%	--

The spores are thoroughly mixed with the adjuvants and the mixture is thoroughly ground in a suitable mill, affording wettable powders which can be diluted with water to give suspensions of the desired concentrations.

A2. Emulsifiable concentrate

<i>Bacillus thuringiensis</i> spores	10%
octylphenol polyethylene glycol ether (4-5 moles ethylene oxide)	3%
clacium dodecylbenzensulfonate	3%

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castor oil polyglycol ether (36 moles of ethylene oxide)	4%
cyclohexanone	30%
xylene mixture	50%

Emulsions of any required concentration can be obtained from this concentrate by dilution with water.

A3. Dusts

	a)	b)
<i>Bacillus thuringiensis</i> spores	5%	8%
talcum	95%	--
kaolin	--	92%

Ready for use dusts are obtained by mixing the active ingredient with the carriers and grinding the mixture in a suitable mill.

A4. Extruder Granulate

<i>Bacillus thuringiensis</i> spores	10%
sodium lignosulfonate	2%
carboxymethylcellulose	1%
kaolin	87%

The active ingredient or combination is mixed and ground with the adjuvants and the mixture is subsequently moistened with water. The mixture is extruded, granulated and the dried in a stream of air.

A5. Coated Granule

<i>Bacillus thuringiensis</i> spores	3%
polyethylene glycol (mol wt 200)	3%
kaolin	94%

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The active ingredient or combination is uniformly applied in a mixer to the kaolin moistened with polyethylene glycol. Non-dusty coated granulates are obtained in this manner.

A6. Suspension Concentrate

<i>Bacillus thuringiensis</i> spores	40%
ethylene glycol	10%
nonylphenol polyethylene glycol ether (15 moles of ethylene oxide)	6%
sodium lignosulfonate	10%
carboxymethylcellulose	1%
37% aqueous formaldehyde solution	0.2%
silicone oil in the form of a 75% aqueous solution	0.8%
water	32%

The active ingredient or combination is intimately mixed with the adjuvants giving a suspension concentrate from which suspensions of any desired concentration can be obtained by dilution with water.

EXAMPLE 1. AB78 ISOLATION AND CHARACTERIZATION

Bacillus cereus strain AB78 was isolated as a plate contaminant in the laboratory on T3 media (per liter: 3 g tryptone, 2 g tryptose, 1.5 g yeast extract, 0.05 M sodium phosphate (pH 6.8), and 0.005 g MnCl₂; Travers, R.S. 1983). During log phase growth, AB78 gave significant activity against western corn rootworm. Antibiotic activity against gram-positive *Bacillus* spp. was also demonstrated (Table 12).

TABLE 12

Antibiotic activity of AB78 culture supernatant

Bacteria tested	Zone of inhibition(cm)	
	AB78	Streptomycin
<i>E. coli</i>	0.0	3.0
<i>B. megaterium</i>	1.1	2.2
<i>B. mycoides</i>	1.3	2.1
<i>B. cereus</i> CB	1.0	2.0
<i>B. cereus</i> 11950	1.3	2.1
<i>B. cereus</i> 14579	1.0	2.4
<i>B. cereus</i> AB78	0.0	2.2
<i>Bt</i> var. <i>israelensis</i>	1.1	2.2
<i>Bt</i> var. <i>tenebrionis</i>	0.9	2.3

Morphological characteristics of AB78 are as follows:

Vegetative rods straight, 3.1-5.0 mm long and 0.5-2.0 mm wide. Cells with rounded ends, single in short chains. Single subterminal, cylindrical-oval, endospore formed per cell. No parasporal crystal formed. Colonies opaque, erose, lobate and flat. No pigments produced. Cells motile. Flagella present.

Growth characteristics of AB78 are as follows:

Facultative anaerobe with optimum growth temperature of 21-30°C. Will grow at 15, 20, 25, 30 and 37°C. Will not grow above 40°C. Grows in 5-7% NaCl.

Table 13 provides the biochemical profile of AB78.

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TABLE 13.
Biochemical characteristics of *B. cereus* strain AB78.

Acid from L-arabinose	-	Methylene blue reoxidized	+
Gas from L-arabinose	-	Nitrate reduced	+
Acid from D-xylose	-	NO ₃ reduced to NO ₂	+
Gas from D-xylose	-	VP	+
Acid from D-glucose	+	H ₂ O ₂ decomposed.	+
Gas from D-glucose	-	Indole	-
Acid from lactose	-	Tyrosine decomposed	+
Gas from lactose	-	Dihydroxiacetone	-
Acid from sucrose	-	Litmus milk acid	-
Gas from sucrose	-	Litmus milk coagulated	-
Acid from D-mannitol	-	Litmus milk alkaline	-
Gas from D-mannitol	-	Litmus milk peptonized	-
Propionate utilization	+	Litmus milk reduced	-
Citrate utilization	+	Casein hydrolyzed	+
Hippurate hydrolysis	w	Starch hydrolyzed	+
Methylene blue reduced	+	Gelatin liquidified	+
Lecithinase produced	w		

w= weak reaction

EXAMPLE 2. BACTERIAL CULTURE

A subculture of Bc strain AB78 was used to inoculate the following medium, known as TB broth:

Tryptone	12	g/l
Yeast Extract	24	g/l
Glycerol	4	ml/l
KH ₂ PO ₄	2.1	g/l
K ₂ HPO ₄	14.7	g/l
pH 7.4		

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The potassium phosphate was added to the autoclaved broth after cooling. Flasks were incubated at 30°C on a rotary shaker at 250 rpm for 24 h-36 h, which represents an early to mid-log growth phase.

The above procedure can be readily scaled up to large fermentors by procedures well known in the art.

During vegetative growth, usually 24-36 h. after starting the culture, which represents an early to mid-log growth phase, AB78 bacteria were centrifuged from the culture supernatant. The culture supernatant containing the active protein was used in bioassays.

EXAMPLE 3. INSECT BIOASSAYS

B. cereus strain AB78 was tested against various insects as described below.

Western, Northern and Southern corn rootworm, *Diabrotica virgifera virgifera*, *D. longcornis barberi* and *D. undecimpunctata howardi*, respectively: dilutions were made of AB78 culture supernatant grown 24-36 h., mixed with molten artificial diet (Marrone *et al.* (1985) *J. of Economic Entomology* 78:290-293) and allowed to solidify. Solidified diet was cut and placed in dishes. Neonate larvae were placed on the diet and held at 30°C. Mortality was recorded after 6 days.

E. coli clone bioassay: *E. coli* cells were grown overnight in broth containing 100 µg/ml ampicillin at 37°C. Ten ml culture was sonicated 3X for 20 sec each. 500 µl of sonicated culture was added to molten western corn rootworm diet.

Colorado potato beetle, *Leptinotarsa decemlineata*: dilutions in Triton X-100 (to give final concentration of 0.1% TX-100) were made of AB78 culture supernatant grown 24-36 h. Five cm² potato leaf pieces were dipped into these dilutions, air dried, and placed on moistened filter paper in plastic dishes. Neonate larvae were placed on the leaf pieces and held at 30°C. Mortality was recorded after 3-5 days.

Yellow mealworm, *Tenebrio molitor*. dilutions were made of AB78 culture supernatant grown 24-36 h., mixed with molten artificial diet (Bioserv #F9240) and allowed to solidify. Solidified diet was cut and placed in plastic dishes. Neonate larvae were placed on the diet and held at 30°C. Mortality was recorded after 6-8 days.

European corn borer, black cutworm, tobacco budworm, tobacco hornworm and beet armyworm; *Ostrinia nubilalis*, *Agrotis ipsilon*, *Heliothis virescens*, *Manduca sexta* and *Spodoptera exigua*, respectively: dilutions, in TX-100 (to give final concentration of 0.1% TX-100), were made of AB78 culture supernatant grown 24-36 hrs. 100 µl was pipetted onto the surface of 18 cm² of solidified artificial diet (Bioserv #F9240) and allowed to air dry. Neonate larvae were then placed onto the surface of the diet and held at 30°C. Mortality was recorded after 3-6 days.

Northern house mosquito, *Culex pipiens*: dilutions were made of AB78 culture supernatant grown 24-36 h. 100 µl was pipetted into 10 ml water in a 30 ml plastic cup. Third instar larvae were added to the water and held at room temperature. Mortality was recorded after 24-48 hours. The spectrum of entomocidal activity of AB78 is given in Table 14.

TABLE 14
Activity of AB78 culture supernatant against various insect species

Insect species tested to date	Order	Activity
Western corn rootworm (<i>Diabrotica virgifera</i> <i>virgifera</i>)	Col	+++
Northern corn rootworm (<i>Diabrotica longicornis</i> <i>barberi</i>)	Col	+++
Southern corn rootworm (<i>Diabrotica undecimpunctata</i> <i>howardi</i>)	Col	-
Colorado potato beetle (<i>Leptinotarsa decemlineata</i>)	Col	-
Yellow mealworm (<i>Tenebrio molitor</i>)	Col	-

European corn borer	
(<i>Ostrinia nubilalis</i>)	Lep
Tobacco budworm	
(<i>Heliothis virescens</i>)	Lep
Tobacco hornworm	
(<i>Manduca sexta</i>)	Lep
Beet armyworm	
(<i>Spodoptera exigua</i>)	Lep
Black cutworm	
(<i>Agrotis ipsilon</i>)	Lep
Northern house mosquito	
(<i>Culex pipiens</i>)	Dip

The newly discovered *B. cereus* strain AB78 showed a significantly different spectrum of insecticidal activity as compared to known coleopteran active δ-endotoxins from Bt. In particular, AB78 showed more selective activity against beetles than known coleopteran-active Bt strains in that it was specifically active against *Diabrotica spp.* More specifically, it was most active against *D. virgifera virgifera* and *D. longicornis barberi* but not *D. undecimpunctata howardi*.

A number of *Bacillus* strains were bioassayed for activity during vegetative growth (Table 15) against western corn rootworm. The results demonstrate that AB78 is unique in that activity against western corn rootworm is not a general phenomenon.

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TABLE 15

Activity of culture supernatants from various *Bacillus spp.* against western corn rootworm

<i>Bacillus</i> strain	Percent
	WCRW mortality
<i>B. cereus</i> AB78 (Bat.1)	100
<i>B. cereus</i> AB78 (Bat.2)	100
<i>B. cereus</i> (Carolina Bio.)	12
<i>B. cereus</i> ATCC 11950	12
<i>B. cereus</i> ATCC 14579	8
<i>B. mycoides</i> (Carolina Bio.)	30
<i>B. popilliae</i>	28
<i>B. thuringiensis</i> HD135	41
<i>B. thuringiensis</i> HD191	9
<i>B. thuringiensis</i> GC91	4
<i>B. thuringiensis isrealensis</i>	24
Water Control	4

Specific activity of AB78 against western corn rootworm is provided in Table 16.

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TABLE 16

Activity of AB78 culture supernatant against neonate western corn rootworm

Culture supernatant concentration (μ l/ml)	Percent WCRW mortality
100	100
25	87
10	80
5	40
2.5	20
1	6
0	0

The LC₅₀ was calculated to be 6.2 μ l of culture supernatant per ml of western corn rootworm diet.

The cell pellet was also bioassayed and had no activity against WCRW. Thus, the presence of activity only in the supernatant indicates that this VIP is an exotoxin.

EXAMPLE 4. ISOLATION AND PURIFICATION OF CORN ROOTWORM ACTIVE PROTEINS FROM AB78.

Culture media free of cells and debris was made to 70% saturation by the addition of solid ammonium sulfate (472 g/L). Dissolution was at room temperature followed by cooling in an ice bath and centrifugation at 10,000 X g for thirty minutes to pellet the precipitated proteins. The supernatant was discarded and the pellet was dissolved in 1/10 the original volume of 20 mM TRIS-HCl at pH 7.5. The dissolved pellet was desalting either by dialysis in 20 mM TRIS-HCl pH 7.5, or passing through a desalting column.

The desalting material was titrated to pH 3.5 using 20 mM sodium citrate pH 2.5. Following a thirty minute room temperature incubation the solution was centrifuged at

3000 X g for ten minutes. The supernatant at this stage contained the greatest amount of active protein.

Following neutralization of the pH to 7.0 the supernatant was applied to a Mono-Q, anion exchange, column equilibrated with 20 mM TRIS pH 7.5 at a flow rate of 300 mL/min. The column was developed with a stepwise and linear gradient employing 400 mM NaCl in 20 mM TRIS pH 7.5.

Bioassay of the column fractions and SDS-PAGE analysis were used to confirm the active fractions. SDS-PAGE analysis identified the biologically active protein as having components of a molecular weight in the range of about 80 kDa and 50 kDa.

EXAMPLE 5. SEQUENCE ANALYSIS OF THE CORN ROOTWORM ACTIVE PROTEIN

The 80 kDa component isolated by SDS-PAGE was transferred to PVDF membrane and was subjected to amino-terminal sequencing as performed by repetitive Edman cycles on an ABI 470 pulsed-liquid sequencer. Transfer was carried out in 10 mM CAPS buffer with 10% methanol pH 11.0 as follows:

Incubation of the gel following electrophoresis was done in transfer buffer for five minutes. ProBlott PVDF membrane was wetted with 100% MeOH briefly then equilibrated in transfer buffer. The sandwich was arranged between foam sponges and filter paper squares with the configuration of cathode-gel-membrane-anode.

Transfer was performed at 70 V constant voltage for 1 hour.

Following transfer, the membrane was rinsed with water and stained for two minutes with 0.25% Coomassie Blue R-250 in 50% MeOH.

Destaining was done with several rinses with 50% MeOH 40% water 10% acetic acid.

Following destaining the membrane was air dried prior to excision of the bands for sequence analysis. A BlottCartridge and appropriate cycles were utilized to achieve maximum efficiency and yield. Data analysis was performed using model 610 Sequence Analysis software for identifying and quantifying the PTH-amino acid derivatives for each sequential cycle.

The N-terminal sequence was determined to be:

NH₂-Lys-Arg-Glu-Ile-Asp-Glu-Asp-Thr-Asp-Thr-Asx-Gly-Asp-Ser-Ile-Pro-

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(SEQ ID NO:8) where Asx represents Asp or Asn. The complete amino acid sequence for the 80 kDa component is disclosed in SEQ ID NO:7. The DNA sequence which encodes SEQ ID NO:7 is disclosed in SEQ ID NO:6.

EXAMPLE 6. CONSTRUCTION OF DNA PROBE

An oligonucleotide probe for the region of the gene encoding amino acids 3-9 of the N-terminal sequence (Example 5) was generated. The probe was synthesized based on the codon usage of a *Bacillus thuringiensis* (Bt) δ-endotoxin gene. The nucleotide sequence

5'- GAA ATT GAT CAA GAT ACN GAT -3' (SEQ ID NO:9)

was used as a probe in Southern hybridizations. The oligonucleotide was synthesized using standard procedures and equipment.

EXAMPLE 7. ISOELECTRIC POINT DETERMINATION OF THE CORN ROOTWORM ACTIVE PROTEIN

Purified protein from step 5 of the purification process was analyzed on a 3-9 pI isoelectric focusing gel using the Phastgel electrophoresis system (Pharmacia). Standard operating procedures for the unit were followed for both the separation and silver staining development procedures. The pI was approximated at about 4.9.

EXAMPLE 8. PCR DATA ON AB78

PCR analysis (See, for example US patent application serial no. 08/008,006; and, Carozzi *et al.* (1991) *Appl. Environ. Microbiol.* 57(11):3057-3061, herein incorporated by reference.) was used to verify that the *B. cereus* strain AB78 did not contain any insecticidal crystal protein genes of *B. thuringiensis* or *B. sphaericus* (Table 17).

TABLE 17

Bacillus insecticidal crystal protein gene primers tested by PCR against AB78 DNA.

Primers Tested	Product Produced
2 sets specific for CryIIIA	Negative
CryIIIB	Negative
2 sets specific for CryIA	Negative
CryIA(a)	Negative
CryIA(b) specific	Negative
CryIB	Negative
CryIC specific	Negative
CryIE specific	Negative
2 sets specific for <i>B. sphaericus</i>	Negative
2 sets specific for CryIV	Negative
<i>Bacillus</i> control (PI-PLC)	Positive

EXAMPLE 9. COSMID CLONING OF TOTAL DNA FROM *B. CEREUS* STRAIN AB78

The VIP1A(a) gene was cloned from total DNA prepared from strain AB78 as follows:

Isolation of AB78 DNA was as follows:

1. Grow bacteria in 10 ml L-broth overnight. (Use 50 ml sterile centrifuge tube)
2. Add 25 ml of fresh L-broth and ampicillin (30 µg/ml).
3. Grow cells 2-6 h. at 30°C with shaking.
4. Spin cells in a 50 ml polypropylene orange cap tube in IEC benchtop clinical centrifuge at 3/4 speed.
5. Resuspend cell pellet in 10 ml TES (TES = 50 mM TRIS pH 8.0, 100 mM EDTA, 15 mM NaCl).
6. Add 30 mg lysozyme and incubate 2 hrs at 37°C.

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7. Add 200 µl 20% SDS and 400 µl Proteinase K stock (20 mg/ml). Incubate at 37°C.
8. Add 200 µl fresh Proteinase K. Incubate 1 hr. at 55°C. Add 5 ml TES to make 15 ml final volume.
9. Phenol extract twice (10 ml phenol, spin at room temperature at 3/4 speed in an IEC benchtop clinical centrifuge). Transfer supernatant (upper phase) to a clean tube using a wide bore pipette.
10. Extract once with 1:1 vol. phenol:chloroform/isoamyl alcohol (24:1 ratio).
11. Precipitate DNA with an equal volume of cold isopropanol; Centrifuge to pellet DNA.
12. Resuspend pellet in 5 ml TE.
13. Precipitate DNA with 0.5 ml 3M NaOAc pH 5.2 and 11 ml 95% ethanol. Place at -20°C for 2 h.
14. "Hook" DNA from tube with a plastic loop, transfer to a microfuge tube, spin, pipette off excess ethanol, dry in vacuo.
15. Resuspend in 0.5 ml TE. Incubate 90 min. at 65°C to help get DNA back into solution.
16. Determine concentration using standard procedures.

Cosmid Cloning of AB78

All procedures, unless indicated otherwise, were performed according to Stratagene Protocol, Supercos 1 Instruction Manual, Cat. No. 251301.

Generally, the steps were as follows:

- A. Sau 3A partial digestion of the AB78 DNA.
- B. Preparation of vector DNA
- C. Ligation and packaging of DNA
- D. Tittering the cosmid library

1. Start a culture of HB101 cells by placing 50 ml of an overnight culture in 5 mls of TB with 0.2% maltose. Incubate 3.5 hrs. at 37°C.
2. Spin out cells and resuspend in 0.5 ml 10 mM MgSO₄.
3. Add together:
 - 100 l cells
 - 100 l diluted packaging mixture
 - 100 l 10 mM MgSO₄

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30 l TB

4. Adsorb at room temperature for 30 minutes with no shaking.
5. Add 1 ml TB and mix gently. Incubate 30 minutes at 37°C.
6. Plate 200 l onto L-amp plates. Incubate at 37°C overnight.

At least 400 cosmid clones were selected at random and screened for activity against western corn rootworm as described in Example 3. DNA from 5 active clones and 5 non-active clones were used in Southern hybridizations. Results demonstrated that hybridization using the above described oligonucleotide probe correlated with western corn rootworm activity (Table 18).

Cosmid clones P3-12 and P5-4 have been deposited with the Agricultural Research Service Patent Culture Collection (NRRL) and given Accession Nos. NRRL B-21061 and NRRL B-21059 respectively.

TABLE 18
Activity of AB78 cosmid clones against western corn rootworm.

Clone	Mean percent mortality (N=4)
Clones which hybridize with probe	
P1-73	47
P1-83	64
P2-2	69
P3-12	85
P5-4	97
Clones which do not hybridize with probe	
P1-2	5
P3-8	4

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P3-9	12
P3-18	0
P4-6	9

EXAMPLE 10. IDENTIFICATION OF A 6 KB REGION ACTIVE AGAINST WESTERN CORN ROOTWORM.

DNA from P3-12 was partially digested with restriction enzyme Sau 3A, and ligated into the *E. coli* vector pUC19 and transformed into *E. coli*. A DNA probe specific for the 80 kDa VIP1A(a) protein was synthesized by PCR amplification of a portion of P3-12 DNA. Oligonucleotides MK113 and MK117, which hybridize to portions of VIP1A(a), were synthesized using the partial amino acid sequence of the 80 kDa protein. Plasmid subclones were identified by colony hybridization to the PCR-generated probe, and tested for activity against western corn rootworm. One such clone, PL2, hybridized to the PCR-generated fragment, and was active against western corn rootworm in the assay previously described.

A 6 kb *Cla* I restriction fragment from pL2 was cloned into the *Sma* I site of the *E. coli-Bacillus* shuttle vector pHT 3101 (Lereclus, D. et al., *FEMS Microbiology Letters* 60:211-218 (1989)) to yield pCIB6021. This construct confers anti-western corn rootworm activity upon both *Bacillus* and *E.coli* strains, in either orientation. pCIB6022 contains this same 6 kb *Cla* I fragment in pBluescript SK(+) (Stratagene), produces equivalent VIP1A(a) protein (by western blot), and is also active against western corn rootworm.

The nucleotide sequence of pCIB6022 was determined by the dideoxy termination method of Sanger et al., *Proc. Natl. Acad. Sci. USA*, 74:5463-5467 (1977), using PRISM Ready Reaction Dye Deoxy Terminator Cycle Sequencing Kits and PRISM Sequenase® Terminator Double-Stranded DNA Sequencing Kit and analyzed on an ABI 373 automatic sequencer. The sequence is given in SEQ ID NO:1. The 6 kb fragment encodes both VIP1A(a) and VIP2A(a), as indicated by the open reading frames described in SEQ ID NO:1. The sequence encoding VIP2A(a) is further disclosed in SEQ ID NO:4. The relationship between VIP1A(a) and VIP2A(a) within the 6 kb fragment found in pCIB6022 is depicted in Table 19. pCIB6022 was

deposited with the Agricultural Research Service, Patent Culture Collection, (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA, and given the Accession No. NRRL B-21222.

EXAMPLE 11. FUNCTIONAL DISSECTION OF THE VIP1A(a) DNA REGION.

To confirm that the VIP1A(a) open reading frame (ORF) is necessary for insecticidal activity a translational frameshift mutation was created in the gene. The restriction enzyme Bgl II recognizes a unique site located 857 bp into the coding region of VIP1A(a). pCIB6201 was digested with Bgl II, and the single-stranded ends filled-in with DNA polymerase (Klenow fragment) and dNTPS. The plasmid was re-ligated and transformed into *E. coli*. The resulting plasmid, pCIB6203, contains a four nucleotide insertion in the coding region of VIP1A(a). pCIB6203 does not confer WCRW insecticidal activity, confirming that VIP1A(a) is an essential component of western corn rootworm activity.

To further define the region necessary to encode VIP1A(a), subclones of the VIP1A(a) and VIP2A(a) (auxiliary protein) region were constructed and tested for their ability to complement the mutation in pCIB6203. pCIB6023 contains the 3.7kb Xba I-EcoRV fragment in pBluescript SK(+) (Stratagene). Western blot analysis indicates that pCIB6023 produces VIP1A(a) protein of equal size and quantity as clones PL2 and pCIB6022. pCIB6023 contains the entire gene encoding the 80 kD protein. pCIB6023 was deposited with the Agricultural Research Service, Patent Culture Collection, (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA, and given the Accession No. NRRL B-21223N. pCIB6206 contains the 4.3 kb Xba I-Cla I fragment from pCIB6022 in pBluescript SK(+) (Stratagene). pCIB6206 was also deposited with the Agricultural Research Service, Patent Culture Collection, (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA, and given the Accession No. NRRL B-21321.

pCIB6023, pCIB6206, and pCIB6203 do not produce detectable western corn rootworm activity when tested individually. However, a mixture of cells containing pCIB6203 (VIP1A(a)-mutated, plus VIP2A(a)) and cells containing pCIB6023 (only

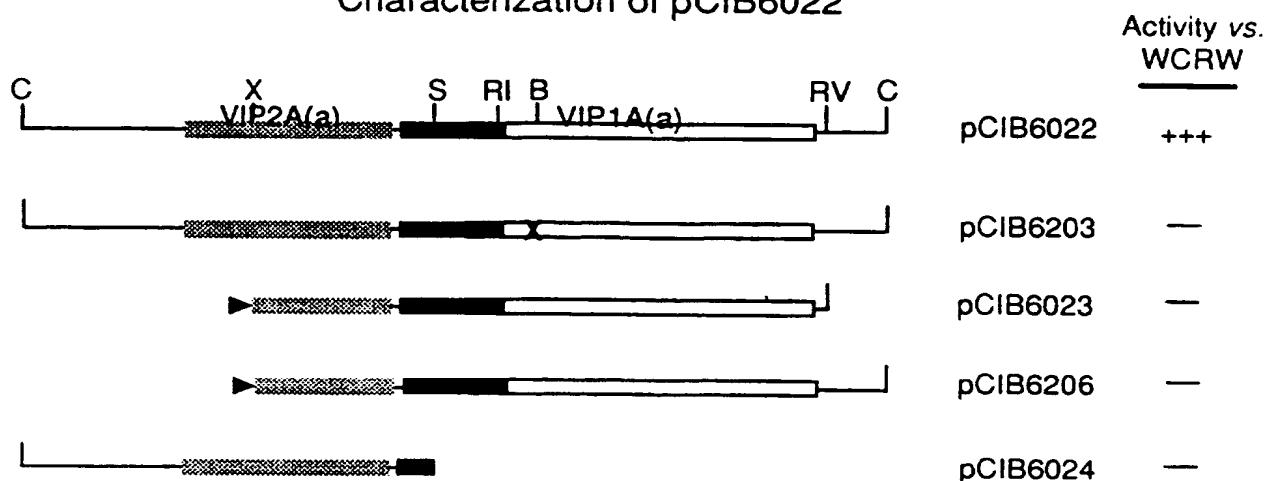
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VIP1A(a)) shows high activity against western corn rootworm. Similarly, a mixture of cells containing pCIB6206 and cells containing pCIB6203 shows high activity against western corn rootworm.

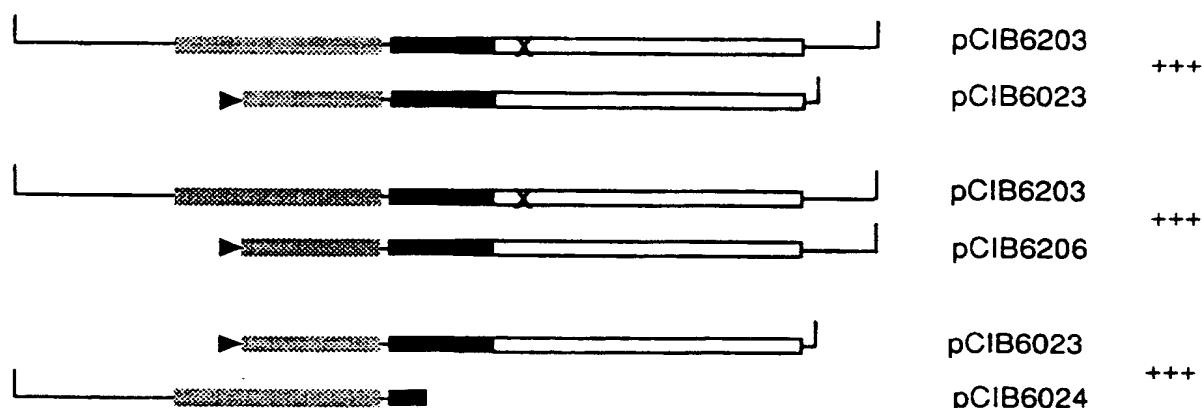
To further define the limits of VIP2A(a), we constructed pCIB6024, which contains the entirety of VIP2A(a), but lacks most of the VIP1A(a) coding region. pCIB6024 was constructed by gel purifying the 2.2 kb Cla I-Sca I restriction fragment from pCIB6022, filling in the single-stranded ends with DNA polymerase (Klenow fragment) and dNTPs, and ligating this fragment into pBluescript SK(+) vector (Stratagene) digested with the enzyme Eco RV. Cells containing pCIB6024 exhibit no activity against western corn rootworm. However, a mixture of cells containing pCIB6024 and cells containing pCIB6023 shows high activity against western corn rootworm .(See Table 19).

Thus, pCIB6023 and pCIB6206 must produce a functional VIP1A(a) gene product, while pCIB6203 and pCIB6024 must produce a functional VIP2A(a) gene product. These results suggest a requirement for a gene product(s) from the VIP2A(a) region, in combination with VIP1A(a), to confer maximal western corn rootworm activity. (See Table 19.)

Table 19
Characterization of pCIB6022



Functional Complementation of VIP



Boxed regions represent the extent of VIP1A(a) and VIP2A(a). White box represents the portion of VIP1 encoding the 80 kDa peptide observed in *Bacillus*. Dark box represents the N-terminal 'propeptide' of VIP1A(a) predicted by DNA sequence analysis. Stippled box represents the VIP2A(a) coding region. Large 'X' represents the location of the frameshift mutation introduced into VIP1A(a). Arrows represent constructs transcribed by the beta-galactosidase

EXAMPLE 12. AB78 ANTIBODY PRODUCTION

Antibody production was initiated in 2 Lewis rats to allow for both the possibility of moving to production of hybridoma cell lines and also to produce enough serum for limited screening of genomic DNA library. Another factor was the very limited amount of antigen available and the fact that it could only be produced to purity by PAGE and subsequent electrotransfer to nitrocellulose.

Due to the limited availability of antigen on nitrocellulose, the nitrocellulose was emulsified in DMSO and injected into the hind footpads of the animals to elicit B-cell production in the popliteal lymph nodes just upstream. A strong reacting serum was produced as judged by western blot analysis with the first production bleed. Several subsequent injections and bleeds produced enough serum to accomplish all of the screening required.

Hybridoma production with one of the rats was then initiated. The popliteal lymph node was excised, macerated, and the resulting cells fused with mouse myeloma P3x63Ag8.653. Subsequent cell screening was accomplished as described below. Four initial wells were selected which gave the highest emulsified antigen reaction to be moved to limited dilution cloning. An additional 10 wells were chosen for expansion and cryopreservation.

Procedure to Emulsify AB78 on nitrocellulose in DMSO for ELISA screening:

After electrotransfer of AB78 samples run on PAGE to nitrocellulose, the reversible stain Ponceau S is used to visualize all protein transferred. The band corresponding to AB78 toxin, previously identified and N-terminal sequenced, was identified and excised from nitrocellulose. Each band is approximately 1 mm x 5 mm in size to minimize the amount of nitrocellulose emulsified. A single band is placed in a microfuge tube with 250 µl of DMSO and macerated using a plastic pestle (Kontes, Vineland, NJ). To aid in emulsification, the DMSO mixture is heated for 2-3 minutes at 37 C-45 C. Some further maceration might be necessary following heating; however, all of the nitrocellulose should be emulsified. Once the AB78 sample is emulsified, it is placed on ice. In preparation for microplate coating with the emulsified antigen, the sample must be diluted in borate buffered saline as follows: 1:5, 1:10, 1:15, 1:20, 1:30, 1:50, 1:100, and 0. The coating antigen must be prepared fresh immediately prior to use.

ELISA protocol:

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1. Coat with AB78/DMSO in BBS. Incubate overnight at 4°C.
2. Wash plate 3X with 1X ELISA wash buffer.
3. Block (1% BSA & 0.05% Tween 20 in PBS) for 30 minutes at Room Temperature.
4. Wash plate 3X with 1X ELISA wash buffer.
5. Add rat serum. Incubate 1.5 hours at 37°C.
6. Wash plate 3X with 1X ELISA wash buffer.
7. Add goat anti-rat at a concentration of 2 µg/ml in ELISA diluent. Incubate 1 hr. at 37°C.
8. Wash plate 3X with 1X ELISA wash buffer.
9. Add rabbit anti-goat alkaline phosphatase at 2 µg/ml in ELISA diluent. Incubate 1 hr. at 37°C.
10. Wash 3X with 1X ELISA wash buffer.
11. Add Substrate. Incubate 30 minutes at room temperature.
12. Stop with 3N NaOH after 30 minutes.

Preparation of VIP2A(a) Antisera

A partially purified AB78 culture supernatant was separated by discontinuous SDS PAGE (Novex) following manufacturer's instructions. Separated proteins were electrophoresed to nitrocellulose (S&S #21640) as described by Towbin *et al.*, (1979). The nitrocellulose was stained with Ponceau S and the VIP2A(a) band identified. The VIP2A(a) band was excised and emulsified in DMSO immediately prior to injection. A rabbit was initially immunized with emulsified VIP2A(a) mixed approximately 1:1 with Freund's Complete adjuvant by intramuscular injection at four different sites. Subsequent immunizations occurred at four week intervals and were identical to the first, except for the use of Freund' Incomplete adjuvant. The first serum harvested following immunization reacted with VIP2A(a) protein. Western blot analysis of AB78 culture supernatant using this antisera identifies predominately full length VIP2A(a) protein.

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EXAMPLE 13. ACTIVATION OF INSECTICIDAL ACTIVITY OF NON-ACTIVE BT STRAINS WITH AB78 VIP CLONES.

Adding pCIB6203 together with a 24 h culture (early to mid-log phase) supernatant from Bt strain GC91 produces 100% mortality in *Diabrotica virgifera virgifera*. Neither pCIB6203 nor GC91 is active on *Diabrotica virgifera virgifera* by itself. Data are shown below:

Test material	Percent <i>Diabrotica</i> mortality
pCIB6203	0
GC91	16
pCIB6203 + GC91	100
Control	0

EXAMPLE 14. ISOLATION AND BIOLOGICAL ACTIVITY OF *B. CEREUS* AB81.

A second *B. cereus* strain, designated AB81, was isolated from grain bin dust samples by standard methodologies. A subculture of AB81 was grown and prepared for bioassay as described in Example 2. Biological activity was evaluated as described in Example 3. The results are as follows:

Insect species tested	Percent Mortality
<i>Ostrinia nubilalis</i>	0
<i>Agrotis ipsilon</i>	0
<i>Diabrotica virgifera virgifera</i>	55

EXAMPLE 15. ISOLATION AND BIOLOGICAL ACTIVITY OF
B. THURINGIENSIS AB6.

A *B. thuringiensis* strain, designated AB6, was isolated from grain bin dust samples by standard methods known in the art. A subculture of AB6 was grown and prepared for bioassay as described in Example 2. Half of the sample was autoclaved 15 minutes to test for the presence of β -exotoxin.

Biological activity was evaluated as described in Example 3. The results are as follows:

Insect species tested	Percent Mortality
<i>Ostrinia nubilalis</i>	0
<i>Agrotis ipsilon</i>	100
<i>Agrotis ipsilon</i> (autoclaved sample)	0
<i>Diabrotica virgifera virgifera</i>	0

The reduction of insecticidal activity of the culture supernatant to insignificant levels by autoclaving indicates that the active principle is not β -exotoxin.

Strain AB6 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA, and given Accession No. NRRL B-21060.

EXAMPLE 16. ISOLATION AND BIOLOGICAL CHARACTERIZATION OF
B. THURINGIENSIS AB88.

A Bt strain, designated AB88, was isolated from grain bin dust samples by standard methodologies. A subculture of AB88 was grown and prepared for bioassay as described in Example 2. Half of the sample was autoclaved 15 minutes to test for the presence of β -exotoxin. Biological activity was evaluated against a number of insect species as described in Example 3. The results are as follows:

Insect species tested	Order	Percent mortality of culture supernatant	
		Non-autoclaved	Autoclaved
<i>Agrotis ipsilon</i>	<i>Lepidoptera</i>	100	5
<i>Ostrinia nubilalis</i>	<i>Lepidoptera</i>	100	0
<i>Spodoptera frugiperda</i>	<i>Lepidoptera</i>	100	4
<i>Helicoverpa zea</i>	<i>Lepidoptera</i>	100	12
<i>Heliothis virescens</i>	<i>Lepidoptera</i>	100	12
<i>Leptinotarsa decemlineata</i>	<i>Coleoptera</i>	0	0
<i>Diabrotica virgifera</i>	<i>Coleoptera</i>	0	5
<i>virgifera</i>			

The reduction of insecticidal activity of the culture supernatant to insignificant levels by autoclaving indicates that the active principle is not β -exotoxin.

Delta-endotoxin crystals were purified from strain AB88 by standard methodologies. No activity from pure crystals was observed when bioassayed against *Agrotis ipsilon*.

EXAMPLE 17. PURIFICATION OF VIPS FROM STRAIN AB88:

Bacterial liquid culture was grown overnight [for 12h] at 30°C in TB media. Cells were centrifuged at 5000 x g for 20 minutes and the supernatant retained. Proteins present in the supernatant were precipitated with ammonium sulfate (70% saturation),

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centrifuged [at 5000 x g for 15 minutes] and the pellet retained. The pellet was resuspended in the original volume of 20 mM Tris pH 7.5 and dialyzed overnight against the same buffer at 4°C. AB88 dialysate was more turbid than comparable material from AB78. The dialysate was titrated to pH 4.5 using 20 mM sodium citrate (pH 2.5) and, after 30 min incubation at room temperature, the solution was centrifuged at 3000 x g for 10 min. The protein pellet was redissolved in 20 mM Bis-Tris-Propane pH 9.0.

AB88 proteins have been separated by several different methods following clarification including isoelectric focusing (Rotofor, BioRad, Hercules, CA), precipitation at pH 4.5, ion-exchange chromatography, size exclusion chromatography and ultrafiltration.

Proteins were separated on a Poros HQ/N anion exchange column (PerSeptive Biosystems, Cambridge, MA) using a linear gradient from 0 to 500 mM NaCl in 20 mM Bis-Tris-Propane pH 9.0 at a flow rate of 4 ml/min. The insecticidal protein eluted at 250 mM NaCl.

European corn borer (ECB)-active protein remained in the pellet obtained by pH 4.5 precipitation of dialysate. When preparative IEF was done on the dialysate using pH 3-10 ampholytes, ECB insecticidal activity was found in all fractions with pH of 7 or greater. SDS-PAGE analysis of these fractions showed protein bands of MW ~60 kDa and ~80 kDa. The 60 kDa and 80 kDa bands were separated by anion exchange HPLC on a Poros-Q column (PerSeptive Biosystems, Cambridge, MA). N-terminal sequence was obtained from two fractions containing proteins of slightly differing MW, but both of approximately 60 kDa in size. The sequences obtained were similar to each other and to some δ-endotoxins.

anion exchange fraction 23 (smaller): xEPFVSAxxQxxx (SEQ ID NO:10)

anion exchange fraction 28 (larger): xEYENVEPFVSAx (SEQ ID NO:11)

When the ECB-active pH 4.5 pellet was further separated by anion exchange on a Poros-Q column, activity was found only in fractions containing a major band of ~60 kDa.

Black cutworm-active protein also remained in the pellet when AB88 dialysate was brought down to pH 4.5. In preparative IEF using pH 3-10 ampholytes, activity was not found in the ECB-active IEF fractions; instead, it was highest in a fraction of pH 4.5-5.0. Its major components have molecular weights of ~35 and ~80 kDa.

The pH 4.5 pellet was separated by anion exchange HPLC to yield fractions containing only the 35 kDa material and fractions containing both 35 kDa and 80 kDa bands.

EXAMPLE 18. CHARACTERIZATION OF AB88 VIP.

Fractions containing the various lepidopteran active vegetative proteins were generated as described in Example 17. Fractions with insecticidal activity were separated in 8 to 16% SDS-polyacrylamide gels and transferred to PVDF membranes [LeGendre et al, (1989) in: A Practical Guide to Protein and Peptide Purification for Microsequencing, ed Matsudaria PT (Academic Press Inc, New York)]. Biological analysis of fractions demonstrated that different VIPs were responsible for the different lepidopteran species activity.

The *Agrotis ipsilon* activity is due to an 80 kDa and/or a 35 kDa protein, either delivered singly or in combination. These proteins are not related to any δ-endotoxins from Bt as evidenced by the lack of sequence homology of known Bt δ-endotoxin sequences. The vip3A(a) insecticidal protein from strain AB88 is present mostly (at least 75% of the total) in supernatants of AB88 cultures.

Also, these proteins are not found in the AB88 δ-endotoxin crystal. N-terminal sequences of the major δ-endotoxin proteins were compared with the N-terminal sequences of the 80 kDa and 35 kDa VIP and revealed no sequence homology. The N-terminal sequence of the vip3A(a) insecticidal protein posses a number of positively charged residues (from Asn2 to Asn7) followed by a hydrophobic core region (from Thr8 to Ile34). Unlike most of the known secretion proteins, the vip3A(a) insecticidal protein from strain AB88 is not N-terminally processed during export.

A summary of the results follows:

<i>Agrotis</i> VIP N-terminal sequences	N-terminal sequence of major δ-endotoxin proteins
	130 kDa
	MDNNPNINE (SEQ ID NO:14)
80 kDa	80 kDa
MNKNNTKLPTRALP (SEQ ID NO:12)	MDNNPNINE (SEQ ID NO:15)
	60 kDa
	MNVLNSGRTTI (SEQ ID NO:16)
35 kDa	
ALSENTGKDGGYIVP (SEQ ID NO:13)	

The *Ostrinia nubilalis* activity is due to a 60 kDa VIP and the *Spodoptera frugiperda* activity is due to a VIP of unknown size.

Bacillus thuringiensis strain AB88 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA and given the Accession No. NRRL B-21225.

EXAMPLE 18A. ISOLATION AND BIOLOGICAL ACTIVITY OF *B. THURINGIENSIS* AB424

A *B. thuringiensis* strain, designated AB424, was isolated from a moss covered pine cone sample by standard methods known in the art. A subculture of AB424 was grown and prepared for bioassay as described in Example 2.

Biological activity was evaluated as described in Example 3. The results are as follows:

Insect species tested	Percent mortality
<i>Ostrinia nubilalis</i>	100
<i>Agrotis ipsilon</i>	100
<i>Diabrotica virgifera</i>	0
<i>virgifera</i>	

Strain AB424 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA, and given Accession No. NRRL B-21439.

EXAMPLE 18B. CLONING OF THE VIP3A(a) and VIP3A(b) GENES WHICH ENCODE PROTEINS ACTIVE AGAINST BLACK CUTWORM.

Total DNA from isolates AB88 and AB424 was isolated [Ausubel et al (1988), in: Current Protocols in Molecular Biology (John Wiley & Sons, NY)] and digested with the restriction enzymes *Xba*I [library of 4.0 to 5.0 Kb size-fractionated *Xba*I fragments of *B thuringiensis* AB88 DNA] and *Eco*RI [library of 4.5 to 6.0 Kb size-fractionated *Eco*RI fragments *B thuringiensis* AB424 DNA] respectively, ligated into pBluescript vector previously linearized with the same enzymes and dephosphorylated, and transformed into *E. coli* DH5 α strain. Recombinant clones were blotted onto nitrocellulose filters which were subsequently probed with a 32 P labeled 33-bases long oligonucleotide corresponding to the 11-N terminal amino acids of the 80 kDa protein active against *Agrotis ipsilon* (black cutworm). Hybridization was carried out at 42°C in 2 x SSC/0.1% SDS (1 x SSC = 0.15 M NaCl/0.015 M sodium citrate, pH 7.4) for 5 min and twice at 50°C in 1 x SSC/0.1 SDS for 10 min. Four out of 400 recombinant clones were positive. Insect bioassays of the positive recombinants exhibited toxicity to black cutworm larvae comparable to that of AB88 or AB424 supernatants.

Plasmid pCIB7104 contains a 4.5 Kb *Xba*I fragment of AB88 DNA. Subclones were constructed to define the coding region of the insecticidal protein.

E. coli pCIB7105 was constructed by cloning the 3.5 Kb *Xba*I-*Acc*I fragment of pCIB7104 into pBluescript.

Plasmid pCIB7106 contained a 5.0 Kb *Eco*RI fragment of AB424 DNA. This fragment was further digested with *Hinc*II to render a 2.8 kb *Eco*RI-*Hinc*II insert (pCIB7107), which still encoded a functional insecticidal protein.

The nucleotide sequence of pCIB7104, a positive recombinant clone from AB88, and of pCIB7107, a positive recombinant clone from AB424, was determined by the dideoxy termination method of Sanger *et al.*, Proc. Natl. Acad. Sci. USA, 74: 5463-5467 (1977), using PRISM Ready Reaction Dye Deoxy Terminator Cycle Sequencing Kits and PRISM Sequenase® Terminator Double-Stranded DNA Sequencing Kit and analysed on an ABI 373 automatic sequencer.

The clone pCIB7104 contains the VIP3A(a) gene whose coding region is disclosed in SEQ ID NO:28 and the encoded protein sequence is disclosed in SEQ ID NO:29. A synthetic version of the coding region designed to be highly expressed in maize is given in SEQ ID NO:30. Any number of synthetic genes can be designed based on the amino acid sequence given in SEQ ID NO:29.

The clone pCIB7107 contains the VIP3A(b) gene whose coding region is disclosed in SEQ ID NO:31 and the encoded protein is disclosed in SEQ ID NO:32. Both pCIB7104 and pCIB7107 have been deposited with the Agricultural Research Service Patent Culture Collection (NRRL) and given Accession Nos. NRRL B-21422 and B-21423, respectively.

The VIP3A(a) gene contains an open reading frame (ORF) that extends from nucleotide 732 to 3105. This ORF encodes a peptide of 791 amino acids corresponding to a molecular mass of 88,500 daltons. A Shine-Dalgarno (SD) sequence is located 6 bases before the first methionine and its sequence identifies a strong SD for *Bacillus*.

The VIP3A(b) gene is 98% identical to VIP3A(a).

When blot of total DNA isolated from AB88 *B. thuringiensis* cells were probed with a 33-base fragment that spans the N-terminal region of the VIP3A-insecticidal protein, single bands could be observed in different restriction digests. This result was

confirmed by using larger probes spanning the coding region of the gene. A search of the GenBank data base revealed no homology to known proteins.

EXAMPLE 18C. EXPRESSION OF THE VIP3A INSECTICIDAL PROTEINS

The time course for expression of the VIP3A(a) insecticidal protein was analyzed by western blot. Samples from *Bacillus thuringiensis* AB88 cultures were taken throughout 1st growth curve and sporulation. The VIP3A(a) insecticidal protein can be detected in the supernatants of AB88 cultures during logarithmic phase, as early as 15 h after initiating the culture. It reached its maximum level during early stages of stationary phase and remained at high levels during and after sporulation. Similar results were obtained when supernatants of AB424 *Bacillus cereus* cultures were used. The levels of VIP3A(a) insecticidal protein reflected the expression of the VIP3A(a) gene as determined by Northern blot. The initiation of the sporulation was determined by direct microscopic observations and by analyzing the presence of δ-endotoxins in cell pellets. Cry-I type proteins could be detected late in the stationary phase , during and after sporulation.

EXAMPLE 18D. IDENTIFICATION OF NOVEL VIP3-LIKE GENES BY HYBRIDIZATION

To identify *Bacillus* containing genes related to the VIP3A(a) from isolate AB88, a collection of *Bacillus* isolates was screened by hybridization. Cultures of 463 *Bacillus* strains were grown in microtiter wells until sporulation. A 96-pin colony stampel was used to transfer the cultures to 150 mm plates containing L-agar. Inoculated plates were kept at 30°C for 10 hours, then at 4°C overnight. Colonies were blotted onto nylon filters and probed with a 1.2Kb *Hind*III VIP3A(a) derived fragment. Hybridization was performed overnight at 62°C using hybridization conditions of Maniatis *et al.* Molecular Cloning: A Laboratory Manual (1982). Filters were washed with 2xSSC/0.1% SDS at 62°C and exposed to X-ray film.

Of the 463 *Bacillus* strains screened, 60 contain VIP3-like genes that could be detected by hybridization. Further characterization of some of them (AB6 and AB426)

showed that their supernatants contain a BCW insecticidal protein similar to the Vip3 protein that are active against black cutworm.

**EXAMPLE 18E. CHARACTERIZATION OF A *B. thuringiensis* STRAIN M2194
CONTAINING A CRYPTIC VIP3-LIKE GENE**

A *B. thuringiensis* strain, designated M2194, was shown to contain VIP3-like gene(s) by colony hybridization as described in Example 18C. The M2194 VIP3 like gene is considered cryptic since no expression can be detected throughout the bacterial growth phases either by immunoblot analysis using polyclonal antibodies raised against the VIP3A(a) protein isolated from AB88 or by bioassay as described in Example 3.

Antiserum against purified VIP3A(a) insecticidal protein was produced in rabbits. Nitrocellulose-bound protein (50 µg) was dissolved in DMSO and emulsified with Freund's complete adjuvant (Difco). Two rabbits were given subcutaneous injections each month for three month. They were bled 10 days after the second and third injection and the serum was recovered from the blood sample [Harlow et al (1988) in : Antibodies: A Laboratory Manual (Cold Spring Harbor Lab Press, Plainview, NY)].

The M2194 VIP3-like gene was cloned into pKS by following the protocol described in Example 9, which created pCIB7108. *E. coli* containing pCIB7108 which comprises the M2194 VIP3 gene were active against black cutworm demonstrating that the gene encodes a functional protein with insecticidal activity. The plasmid pCIB7108 has been deposited with the Agricultural Research Service Patent Culture Collection (NRRL) and given Accession No. NRRL B-21438.

EXAMPLE 18F. INSECTICIDAL ACTIVITY OF VIP3A PROTEINS

The activity spectrum of VIP3A insecticidal proteins was qualitatively determined in insect bioassays in which recombinant *E. coli* carrying the VIP3A genes were fed to larvae. In these assays, cells carrying the VIP3A(a) and VIP3A(b) genes were insecticidal to *Agrotis ipsilon*, *Spodoptera frugiperda*, *Spodoptera exigua*, *Heliothis virescens* and *Helicoverpa zea*. Under the same experimental conditions, bacterial extracts containing VIP3A proteins did not show any activity against *Ostrinia nubilalis*.

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Effect of VIP⁺A insecticidal proteins on *Agrotis ipsilon* larvae

<u>Treatment</u>	<u>(%) Mortality</u>
TB medium	5
AB88 Supernatant	100
Ab424 Supernatant	100
Buffer	7
<i>E coli</i> pKS	10
<i>E coli</i> pCIB7104 (AB88)	100
<i>E coli</i> pCIB7105 (AB88)	100
<i>E coli</i> pCIB7106 (AB424)	100
<i>E coli</i> pCIB7107 (AB424)	100

Effect of VIP3A insecticidal proteins on lepidopteran insect larvae

<u>Treatment</u>	<u>Insect</u>	<u>(%) Mortality</u>
<i>E coli</i> pKS	BCW	10
	FAW	5
	BAW	10
	TBW	8
	CEW	10
	ECB	5
<i>E coli</i> pCIB7105		
<i>E coli</i> pCIB7107	BCW	100
	FAW	100
	BAW	100
	TBW	100
	CEW	50
	ECB	10

BCW = Black Cut Worm; FAW = Fall Army Worm; BAW = Beet Army Worm; TBW = Tobacco Bud Worm; CEW = Corn Ear Worm; ECB = European Corn Borer

**EXAMPLE 19. ISOLATION AND BIOLOGICAL ACTIVITY OF OTHER
BACILLUS SP.**

Other *Bacillus* species have been isolated which produce proteins with insecticidal activity during vegetative growth. These strains were isolated from environmental samples by standard methodologies. Isolates were prepared for bioassay and assayed as described in Examples 2 and 3 respectively. Isolates which produced insecticidal proteins during vegetative growth with activity against *Agrotis epsilon* in the bioassay are tabulated below. No correlation was observed between the presence of a δ-endotoxin crystal and vegetative insecticidal protein production.

<i>Bacillus</i> isolate	Presence of δ-endotoxin crystal	Percent mortality
AB6	+	100
AB53	-	80
AB88	+	100
AB195	-	60
AB211	-	70
AB217	-	83
AB272	-	80
AB279	-	70
AB289	+	100
AB292	+	80
AB294	-	100
AB300	-	80
AB359	-	100

Isolates AB289, AB294 and AB359 have been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria IL 61604, USA and given the Accession Numbers NRRL B-21227, NRRL B-21229, and NRRL B-21226 respectively.

Bacillus isolates which produce insecticidal proteins during vegetative growth with activity against *Diabrotica virgifera virgifera* are tabulated below.

<i>Bacillus</i> isolate	Presence of δ -endotoxin crystal	Percent mortality
AB52	-	50
AB59	-	71
AB68	+	60
AB78	-	100
AB122	-	57
AB218	-	64
AB256	-	64

Isolates AB59 and AB256 have been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria Illinois 61604, USA, and given the Accession Numbers NRRL B-21228 and NRRL B-21230, respectively.

EXAMPLE 20. IDENTIFICATION OF NOVEL VIP1/VIP2 LIKE GENES BY HYBRIDIZATION

To identify strains containing genes related to those found in the VIP1A(a)/VIP2A(a) region of AB78, a collection of *Bacillus* strains was screened by hybridization. Independent cultures of 463 *Bacillus* strains were grown in wells of 96 well microtiter dishes (five plates total) until the cultures sporulated. Of the strains tested, 288 were categorized as *Bacillus thuringiensis*, and 175 were categorized as other *Bacillus* species based on the presence or absence of δ -endotoxin crystals. For each microtiter dish, a 96-pin colony stamper was used to transfer approximately 10 μ l of spore culture to two 150 mm plates containing L agar. Inoculated plates were grown 4-8 hours at 30 °C, then chilled to 4 °C. Colonies were transferred to nylon filters, and the cells lysed by standard methods known in the art. The filters were hybridized to a DNA probe generated from DNA fragments containing both VIP1A(a) and VIP2A(a) DNA sequences. Hybridization was performed overnight at 65 °C using the hybridization conditions of Church and Gilbert (Church, G.M., and W. Gilbert,

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PNAS, 81:1991-1995 (1984)). Filters were washed with 2x SSC containing 0.1% SDS at 65 °C and exposed to X-Ray film.

Of the 463 *Bacillus* strains screened, 55 strains were identified that hybridized to the VIP1A(a)/VIP2A(a) probe. DNA was isolated from 22 of these strains, and analyzed using a Southern blot with VIP1A(a)/VIP2A(a) DNA as probes. These strains were grouped into 8 classes based on their Southern blot pattern. Each class differed in Southern blot pattern from AB78. One class had a pattern identical to that of the VIP1A(a)/VIP2A(a) homologs from *Bacillus thuringiensis* var *tenebrionis* (see below). Each of the 22 strains was tested for activity against western corn rootworm (WCRW). Three strains, AB433, AB434, and AB435 were found to be active on WCRW. Western blot analysis using VIP2A(a) antisera revealed that strains AB6, AB433, AB434, AB435, AB444, and AB445 produce a protein(s) of equivalent size to VIP2A(a).

Notable among the strains identified was *Bacillus thuringiensis* strain AB6, (NRRL B-21060) which produced a VIP active against black cutworm (*Agrotis ipsilon*) as described in Example 15. Western blot analysis with polyclonal antisera to VIP2A(a) and polyclonal antisera to VIP1A(a) suggests that AB6 produces proteins similar to VIP2A(a) and VIP1A(a). Thus, AB6 may contain VIPs similar to VIP1A(a) and VIP2A(a), but with a different spectrum of insecticidal activity.

**EXAMPLE 21. CLONING OF A VIP1A(a)/VIP2A(a) HOMOLOG FROM
BACILLUS THURINGIENSIS VAR. TENEBRIONIS.**

Several previously characterized *Bacillus* strains were tested for presence of DNA similar to VIP1A(a)/VIP2A(a) by Southern blot analysis. DNA from *Bacillus* strains AB78, AB88, GC91, HD-1 and ATCC 10876 was analyzed for presence of VIP1A(a)/VIP2A(a) like sequences. DNA from Bt strains GC91 and HD-1, and the Bc strain ATCC 10876 did not hybridize to VIP2A(a)/VIP1A(a) DNA, indicating they lack DNA sequences similar to VIP1A(a)/VIP2A(a) genes. Similarly, DNA from the insecticidal strain AB88 (Example 16) did not hybridize to VIP1A(a)/VIP2A(a) DNA region, suggesting that the VIP activity produced by this strain does not result from VIP1A(a)/VIP2A(a) homologs. In contrast, *Bacillus thuringiensis* var. *tenebrionis* (Btt)

contained sequences that hybridized to the VIP1A(a)/VIP2A(a) region. Further analysis confirmed that Btt contains VIP1A(a)/VIP2A(a) like sequences.

To characterize the Btt homologs of VIP2A(a) and VIP1A(a), the genes encoding these proteins were cloned. Southern blot analysis identified a 9.5 kb Eco RI restriction fragment likely to contain the coding regions for the homologs. Genomic DNA was digested with Eco RI, and DNA fragments of approximately 9.5 kb in length were gel-purified. This DNA was ligated into pBluescript SK(+) digested with Eco RI, and transformed into *E. coli* to generate a plasmid library. Approximately 10,000 colonies were screened by colony hybridization for the presence of VIP2A(a) homologous sequences. Twenty eight positive colonies were identified. All twenty eight clones are identical, and contain VIP1A(a)/VIP2A(a) homologs. Clone pCIB7100 has been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria Illinois 61604, USA, and given the Accession Number B-21322. Several subclones were constructed from pCIB7100. A 3.8 kb Xba I fragment from pCIB7100 was cloned into pBluescript SK(+) to yield pCIB7101. A 1.8 kb Hind III fragment and a 1.4 kb Hind III fragment from pCIB7100 were cloned into pBluescript SK(+) to yield pCIB7102 and pCIB7103, respectively. Subclones pCIB7101, pCIB7102 and pCIB7103 have been deposited in the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria Illinois 61604, USA, and given the Accession Numbers B-21323, B-21324 and B-21325 respectively.

The DNA sequence of the region of pCIB7100 containing the VIP2A(a)/VIP1A(a) homologs was determined by the dideoxy chain termination method (Sanger *et al.*, 1977, Proc. Natl. Acad. Sci. USA 74:5463-5467). Reactions were performed using PRISM Ready Reaction Dye Deoxy Terminator Cycle Sequencing Kits and PRISM Sequenase® Terminator Double-Stranded DNA Sequencing Kits, and analyzed on an ABI model 373 automated sequencer. Custom oligonucleotides were used as primers to determine the DNA sequence in certain regions. The DNA sequence of this region is shown in SEQ ID NO:19.

The 4 kb region shown in SEQ ID NO:19 contains two open readings frames (ORFs), which encode proteins with a high degree of similarity to VIP1A(a) and VIP2A(a) proteins from strain AB78. The amino acid sequence of the VIP2A(a)

homolog, designated as VIP2A(b) using the standardized nomenclature, is found at SEQ ID NO:20 and the amino acid sequence of the VIP1A(a) homolog, designated as VIP1A(b) using the standardized nomenclature, is disclosed at SEQ ID NO:21. The VIP2A(b) protein exhibits 91% amino acid identity to VIP2A(a) from AB78. An alignment of the amino acid sequences of the two VIP2 proteins is provided in Table 20. The VIP1A(b) protein exhibits 77 % amino acid identity to VIP1A(a) from AB78. An alignment of these two VIP1 proteins is provided in Table 21. The alignment shown in Table 21 discloses the similarity between VIP1A(b) and VIP1A(a) from AB78. This alignment reveals that the amino terminal regions of the two VIP1 proteins share higher amino acid identity in the amino-terminal region than in the carboxy terminal region. In fact, the amino terminal two thirds (up to aa 618 of the VIP1A(b) sequence shown in Table 21) of the two proteins exhibit 91% identity, while the carboxy-terminal third (from aa 619-833 of VIP1A(b)) exhibit only 35% identity.

Western blot analysis indicated that *Bacillus thuringiensis* var. *tenebrionis* (Btt) produces both VIP1A(a) like and VIP2A(a) like proteins. However, these proteins do not appear to have activity against western corn rootworm. Bioassay for activity against western corn rootworm was performed using either a 24 h culture supernatant from Btt or *E. coli* clone pCIB7100 (which contains the entire region of the VIP1A(a)/VIP2A(a) homologs). No activity against western corn rootworm was detected in either case.

Given the similarity between the VIP2 proteins from Btt and AB78, the ability of VIP2A(b) from Btt to substitute for VIP2A(a) from AB78 was tested. Cells containing pCIB6206 (which produces AB78 VIP1A(a) but not VIP2A(a) protein) were mixed with Btt culture supernatant, and tested for activity against western corn rootworm. While neither Btt culture supernatant nor cells containing pCIB6206 had activity on WCRW, the mixture of Btt and pCIB6206 gave high activity against WCRW. Furthermore, additional bioassay showed that the Btt clone pCIB7100, which contains the Btt VIP1A(b)/VIP2A(b) genes in *E. coli*, also confers activity against WCRW when mixed with pCIB6206. Thus, the VIP2A(b) protein produced by Btt is functionally equivalent to the VIP2A(a) protein produced by AB78.

Thus, the ability to identify new strains with insecticidal activity by using VIP DNA as hybridization probes has been demonstrated. Furthermore, *Bacillus* strains that contain VIP1A(a)/VIP2A(a) like sequences, produce VIP1A(a)/VIP2A(a) like protein.

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yet demonstrate toxicity toward different insect pests. Similar methods can identify many more members of the VIP1/VIP2 family. Furthermore, use of similar methods can identify homologs of other varieties of VIPs (for example, the VIPs from AB88).

TABLE 20

Alignment of VIP2 Amino Acid Sequences from *Bacillus thuringiensis* var. *tenebrionis* (VIP2A(b)) vs. AB78 (VIP2A(a))

Btt	1 MORMEGKLFFVSKTLQVVTRTVLLSTVYSITLLNNVVIKADQLNINSQSK 50 SEQ ID NO:20
	. : : : : . :
AB78	1 MKRMEGKLFMVSKKLQVVTKTVLLSTVFSISILLNEVVIKAEQLNINSQSK 50 SEQ ID NO:2
	51 YTNLQNLKIPDNAEDFKEDKGKAKEWGKEKGEEWRPPATEKGEMNNFLDN 100
	:.. : : : . .
	51 YTNLQNLKITDKVEDFKEDKEKAKEWGKEKEKEWKLTAKEKGKMNFLDN 100
	101 KNDIKTNYKEITFSMAGSCDEIKDLEEIDKIFDKANLSSIIITYKNVEP 150
	. : . : . .
	101 KNDIXTNYKEITFSMAGSFEDIEKDLKEIDKMFDTNLNSNSIITYKNVEP 150
	151 ATIGFNKSLTEGNTINSDAMAQFKEQFLGKDMKFD SYLDTHLTAQQVSSK 200
	. : : : : . :
	151 TTIGFNKSLTEGNTINSDAMAQFKEQFLDRDIKFDSYLDTHLTAQQVSSK 200
	201 KRVILKVTVPSKGSTPTKAGVILNNNEYKMLIDNGYVLHVDKVKVVK 250
	. : : : : : .
	201 ERVILKVTVPSKGSTPTKAGVILNNSEYKMLIDNGYMVHVDKVKVVK 250
	251 KGMECLQVEGTLKKSLDFKNDINAEEAHSGMKIYEDWAKNLASQREALD 300
	: : : : : . : .
	251 KGVECLQIEGTLKKSLDFKNDINAEEAHSGMKNYEAWAKDLTDSQREALD 300

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301 GYARQDYKEINNYLRNQGGSGNEKLDACLKNISDALGKKPIPENITVYRW 350
 |||
 301 GYARQDYKEINNYLRNQGGSGNEKLDACLKNISDALGKKPIPENITVYRW 350

351 CGMPEFGYQISDPLPSLKDFEEQFLNTIKEDKGYMSTSLSSERLAAFGSR 400
 |||
 351 CGMPEFGYQISDPLPSLKDFEEQFLNTIKEDKGYMSTSLSSERLAAFGSR 400

401 KIILRLQVPKGSTGAYLSAIGGFASEKEILDKDSKYHIDKATEVIIKGV 450
 |||
 401 KIILRLQVPKGSTGAYLSAIGGFASEKEILDKDSKYHIDKVTTEVIIKGV 450

451 KRYVVDAATLLTN 462
 |||
 451 KRYVVDAATLLTN 462

TABLE 21

**Alignment of VIP1 Amino Acid Sequences from *Bacillus thuringiensis* var.
tenebrionis (VIP1A(b)) vs. AB78 (VIP1A(a))**

Btt	1 MKNMKKKLASVVTGMLLAPMFINGNVNAVNDASKINQISTTQENQQKEMD 50 SEQ ID NO:21
Ab78	1 MKNMKKKLASVVTCTLLAPMFINGNVNAVYADSKTNQISTTQKNQQKEMD 50 SEQ ID NO:5
	51 RKGLLGYYFKGKDFNNLTMFAPTRDNTLMYDQQTANALLDKKQQEYQSIR 100
	51 RKGLLGYYFKGKDFSNLTMFAPTRDSTLIYDQQTANKLLDKKKQQEYQSIR 100
	101 WIGLIQRKETGDFTFNLSEDEQAIIEIDGKIIISNKGKEKQVVHLEKEKLV 150
	101 WIGLIQSKETGDFTFNLSEDEQAIIEINGKIIISNKGKEKQVVHLEKGKLV 150
	151 PIKIEYQSDTKFNIDSCKTFKELKLFKIDSQNQSQVQ...LRNPEFNKKE 197

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151 PIKIEYQSDTKFNIDSKTFKELKLFKIDSQNQPQQVQQDELRNPEFNKE 200

198 SQEFLAKASKTNLFQKMKRDIDEDTDGDSSIPDLWEENGYTIQNKVAV 247
|||||:|||.||||.|||||:|||||||||||||||||||||||||::|||

201 SQEFLAKPSKINLFTQKMKREIDEDTDGDSSIPDLWEENGYTIQNRIAV 250

248 KWDDSLASKGYTKFVNPLDSHTVGDPYTDYEKAARDLDLSNAKETFNPL 297
|||||||||||||||||:|||||||||||||||||||||||||||||||||

251 KWDDSLASKGYTKFVNPLESHTVGDPYTDYEKAARDLDLSNAKETFNPL 300

298 VAAFPSVNVSMEKVILSPNENLSNSVESHSSTNWSYTNTEGASIEAGGGP 347
|||||||||||||||||:|||||||||||||||||||||||||:||| ||

301 VAAFPSVNVSMEKVILSPNENLSNSVESHSSTNWSYTNTEGASVEAGIGP 350

348 LGLSFGSVTYQHSETVAQEWGTSTGNTSQFNTASAGYLNANVRYNNVGT 397
|:|||||,|||||||||||||||||||||||||||||||||||||

351 KGISFGSVNYQHSETVAQEWGTSTGNTSQFNTASAGYLNANVRYNNVGT 400

398 GAIYDVKPTTSFVLNNNTIATITAKSNSTALRISPQDYPEIGENAIIT 447
|||||||||:|||||||||,|||:|||:|||, |:|:|||

401 GAIYDVKPTTSFVLNNDTIATITAKSNSTALNISPGEYPKGONGIAIT 450

448 SMDDFNSHPITLNKQQVNQLINNKPIMLETDQTDGVYKIRDTHGNIVTGG 497
|||||||||,|||:|||:|||:|||:|||:|||:|||:|||

451 SMDDFNSHPITLNKQQVNQLINNKPMMLTNQTDGVYKIKDTHGNIVTGG 500

498 EWNGVTQQIKAKTASITVDDGKQVAEKRAAKDYGHPEDKTPPLTLKDTL 547
||||,|||||||||..|||||:;||||,||||,|

501 EWNGVIQQIKAKTASIIVDDGERVAEKRAAKDYENPEDKTPSLTLKDAL 550

548 KLSYPDEIKEETNGLLYYDDKPIYESSVMYLDENTAKEVKKQINDTTGKF 597
|||||,.:|||,.:|||||||||||||,||:|||||

551 KLSYPDEIKEIEGLLYYKNKPIYESSVMYLDENTAKEVTKQLNDTTGKF 600

- 91 -

598 KDVNHLYDVKLTPKMNFTIKMASLYDGAENNHS LGTWYLTYNVAGGNTG 647

.... 601 KDVSHLYDVKLTPKMNVTIKLSI LYDNEAESNDNSIGKWTNTNIVSGGNNC 650

648 KROYRSAHSCAHVALSSEAKKKI NONANYYT SMYMKADSTTERTIEFACE 687

10.1007/s00162-017-0462-0

651 KKOYSSNNPDLNTLNTDAOEKLKNRWDYVTSIYMKSEKNTOCETIDCE 700

698 KSAITSKKVI TNNONYORVU LI VANSERNDMD KIVI BONOSTI LUSTVU - 245

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748 TREVISA IN RASIL SPRINT EXTRASTYLE VERSO IL MATTINO

THE UNIVERSITY OF MARYLAND

751 ITDVASIKRENI TDSEI KOIVGRYVUJ FDSII TATVOSVABDU

789 NIKLONUM KETENİYUK SURVEYOR-TECHNIQUE

THESE ARE THE NAMES OF THE SAILORS WHO DIED IN THE WAR OF 1812.

801. ПРИВОДЫ ИЗМЕРЯЮЩИЕ СОСТАВЫ

831 KVA 822

851 EOC 853

EXAMPLE 22. FUSION OF VIP PROTEINS TO MAKE A SINGLE POLYPEPTIDE

VIP proteins may occur in nature as single polypeptides, or as two or more interacting polypeptides. When an active VIP is comprised of two or more interacting protein chains, these protein chains can be produced as a single polypeptide chain from a gene resulting from the fusion of the two (or more) VIP coding regions. The genes encoding the two chains are fused by merging the coding regions of the genes to produce a single open reading frame encoding both VIP polypeptides. The composite polypeptides can be fused to produce the smaller polypeptide as the NH₂ terminus of the fusion protein, or they can be fused to produce the larger of the

polypeptides as the NH₂ terminus of the fusion protein. A linker region can optionally be used between the two polypeptide domains. Such linkers are known in the art. This linker can optionally be designed to contain protease cleavage sites such that once the single fused polypeptide is ingested by the target insect it is cleaved in the linker region to liberate the two polypeptide components of the active VIP molecule.

VIP1A(a) and VIP2A(a) from *B. cereus* strain AB78 are fused to make a single polypeptide by fusing their coding regions. The resulting DNA comprises a sequence given in SEQ ID NO:22 with the encoded protein given in SEQ ID NO:23. In like manner, other fusion proteins may be produced.

The fusion of the genes encoding VIP1A(a) and VIP2A(a) is accomplished using standard techniques of molecular biology. The nucleotides deleted between the VIP1A(a) and VIP2A(a) coding regions are deleted using known mutagenesis techniques or, alternatively, the coding regions are fused using PCR techniques.

The fused VIP polypeptides can be expressed in other organisms using a synthetic gene, or partially synthetic gene, optimized for expression in the alternative host. For instance, to express the fused VIP polypeptide from above in maize, one makes a synthetic gene using the maize preferred codons for each amino acid, see for example EP-A 0618976, herein incorporated by reference. Synthetic DNA sequences created according to these methods are disclosed in SEQ ID NO:17 (maize optimized version of the 100 kDa VIP1A(a) coding sequence), SEQ ID NO:18 (maize optimized version of the 80 kDa VIP1A(a) coding sequence) and SEQ ID NO:24 (maize optimized version of the VIP2A(a) coding sequence).

Synthetic VIP1 and VIP2 genes optimized for expression in maize can be fused using PCR techniques, or the synthetic genes can be designed to be fused at a common restriction site. Alternatively, the synthetic fusion gene can be designed to encode a single polypeptide comprised of both VIP1 and VIP2 domains.

Addition of a peptide linker between the VIP1 and VIP2 domains of the fusion protein can be accomplished by PCR mutagenesis, use of a synthetic DNA linker encoding the linker peptide, or other methods known in the art.

The fused VIP polypeptides can be comprised of one or more binding domains. If more than one binding domain is used in the fusion, multiple target pests are controlled using such a fusion. The other binding domains can be obtained by using all or part of other VIPs; *Bacillus thuringiensis* endotoxins, or parts thereof; or other

proteins capable of binding to the target pest or appropriate binding domains derived from such binding proteins.

One example of a fusion construction comprising a maize optimized DNA sequence encoding a single polypeptide chain fusion having VIP2A(a) at the N-terminal end and VIP1A(a) at the C-terminal end is provided by pCIB5531. A DNA sequence encoding a linker with the peptide sequence PSTPPTPSPSTPPTPS (SEQ ID NO:47) has been inserted between the two coding regions. The sequence encoding this linker and relevant cloning sites is 5'-CCC GGG CCT TCT ACT CCC CCA ACT CCC TCT CCT AGC ACG CCT CCG ACA CCT AGC GAT ATC GGA TC C-3' (SEQ ID NO:48). Oligonucleotides were synthesized to represent both the upper and lower strands and cloned into a pUC vector following hybridization and phosphorylation using standard procedures. ^{sequences needed} A(a) was removed using PCR and replaced by the ^{sequences needed} translation fusion was made by ligating the gene from pCIB5522 (see Example 24), a fragment of the VIP2A(a) gene (identical to synthetic linker having ends that would ligate ^{sequences needed} BamHI site at the 5' end and with ^{sequences needed} PstI-end described below (See SEQ ID NO:35). The fusion was obtained by a four way ligation that resulted in a plasmid containing the VIP2A(a) gene without a translation stop codon, with a linker and the VIP1A(a) coding region without the *Bacillus* secretion signal. The DNA sequence for this construction is disclosed in SEQ ID NO:49, which encodes the fusion protein disclosed in SEQ ID NO:50. A single polypeptide fusion where VIP1A(a) is at the N-terminal end and VIP2A(a) is at the C-terminal end can be made in a similar fashion. Furthermore, either one or both genes can be linked in a translation fusion with or without a linker at either the 5' or the 3' end to other molecules like toxin encoding genes or reporter genes.

EXAMPLE 23. TARGETING OF VIP2 TO PLANT ORGANELLES

Various mechanisms for targeting gene products are known to exist in plants and the sequences controlling the functioning of these mechanisms have been characterized in some detail. For example, the targeting of gene products to the

chloroplast is controlled by a signal sequence found at the amino-terminal end of various proteins. This signal is cleaved during chloroplast import, yielding the mature protein (e.g. Comai *et al.* J. Biol. Chem. 263: 15104-15109 (1988)). These signal sequences can be fused to heterologous gene products such as VIP2 to effect the import of those products into the chloroplast (van den Broeck *et al.* Nature 313: 358-363 (1985)). DNA encoding for appropriate signal sequences can be isolated from the 5' end of the cDNAs encoding the RUBISCO protein, the CAB protein, the EPSP synthase enzyme, the GS2 protein and many other proteins which are known to be chloroplast localized.

Other gene products are localized to other organelles such as the mitochondrion and the peroxisome (e.g. Unger *et al.* Plant Molec. Biol. 13: 411-418 (1989)). The cDNAs encoding these products can also be manipulated to effect the targeting of heterologous gene products such as VIP2 to these organelles. Examples of such sequences are the nuclear-encoded ATPases and specific aspartate amino transferase isoforms for mitochondria. Similarly, targeting to cellular protein bodies has been described by Rogers *et al.* (Proc. Natl. Acad. Sci. USA 82: 6512-6516 (1985)).

By the fusion of the appropriate targeting sequences described above to coding sequences of interest such as VIP2 it is possible to direct the transgene product to any organelle or cell compartment. For chloroplast targeting, for example, the chloroplast signal sequence from the RUBISCO gene, the CAB gene, the EPSP synthase gene, or the GS2 gene is fused in frame to the amino-terminal ATG of the transgene. The signal sequence selected should include the known cleavage site and the fusion constructed should take into account any amino acids after the cleavage site which are required for cleavage. In some cases this requirement may be fulfilled by the addition of a small number of amino acids between the cleavage site and the start codon ATG, or alternatively replacement of some amino acids within the coding sequence. Fusions constructed for chloroplast import can be tested for efficacy of chloroplast uptake by *in vitro* translation of *in vitro* transcribed constructions followed by *in vitro* chloroplast uptake using techniques described by (Bartlett *et al.* In: Edelmann *et al.* (Eds.) Methods in Chloroplast Molecular Biology, Elsevier. pp 1081-1091 (1982); Wasmann *et al.* Mol. Gen. Genet. 205: 446-453 (1986)). These

construction techniques are well known in the art and are equally applicable to mitochondria and peroxisomes.

The above described mechanisms for cellular targeting can be utilized not only in conjunction with their cognate promoters, but also in conjunction with heterologous promoters so as to effect a specific cell targeting goal under the transcriptional regulation of a promoter which has an expression pattern different to that of the promoter from which the targeting signal derives.

A DNA sequence encoding a secretion signal is present in the native *Bacillus* VIP2 gene. This signal is not present in the mature protein which has the N-terminal sequence of LKITDKVEDF (amino acid residues 57 to 66 of SEQ ID NO:2). It is possible to engineer VIP2 to be secreted out of the plant cell or to be targeted to subcellular organelles such as the endoplasmic reticulum, vacuole, mitochondria or plastids including chloroplasts. Hybrid proteins made by fusion of a secretion signal peptide to a marker gene have been successfully targeted into the secretion pathway. (Itirriaga G. et al., The Plant Cell, 1: 381-390 (1989) , Denecke et al., The Plant Cell, 2:51-59 (1990). Amino-terminal sequences have been identified that are responsible for targeting to the ER, the apoplast, and extracellular secretion from aleurone cells (Koehler & Ho, Plant Cell 2: 769-783 (1990)).

The presence of additional signals are required for the protein to be retained in the endoplasmic reticulum or the vacuole. The peptide sequence KDEL/HDEL at the carboxy-terminal of a protein is required for its retention in the endoplasmic reticulum (reviewed by Pelham, Annual Review Cell Biol., 5:1-23 (1989). The signals for retention of proteins in the vacuole have also been characterized. Vacuolar targeting signals may be present either at the amino-terminal portion, (Holwerda et al., The Plant Cell, 4:307-318 (1992), Nakamura et al., Plant Physiol., 101:1-5 (1993)), carboxy-terminal portion, or in the internal sequence of the targeted protein. (Tague et al., The Plant Cell, 4:307-318 (1992), Saalbach et al., The Plant Cell, 3:695-708 (1991)). Additionally, amino-terminal sequences in conjunction with carboxy-terminal sequences are responsible for vacuolar targeting of gene products (Shinshi et al. Plant Molec. Biol. 14: 357-368 (1990)). Similarly, proteins may be targeted to the mitochondria or plastids using specific carboxy terminal signal peptide fusions (Heijne et al., Eur. J. Biochem., 180:535-545 (1989), Archer and Keegstra, Plant Molecular Biology, 23:1105-1115 (1993)).

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In order to target VIP2, either for secretion or to the various subcellular organelles, a maize optimized DNA sequence encoding a known signal peptide(s) may be designed to be at the 5' or the 3' end of the gene as required. To secrete VIP2 out of the cell, a DNA sequence encoding the eukaryotic secretion signal peptide MGWSWIFLFLSGAAGVHCL (SEQ ID NO:25) from PCT application No. IB95/00497 or any other described in the literature (Itirriaga *et al.*, The Plant Cell, 1:381-390 (1989), Denecke, *et al.*, The Plant Cell, 2:51-59 (1990)) may be added to the 5' end of either the complete VIP2 gene sequence or to the sequence truncated to encode the mature protein or the gene truncated to nucleotide 286 or encoding a protein to start at amino acid residue 94 (methionine). To target VIP2 to be retained in the endoplasmic reticulum, a DNA sequence encoding the ER signal peptide KDEL /HDEL, in addition to the secretion signal, can be added to the 3' end of the gene. For vacuolar targeting a DNA sequence encoding the signal peptide SSSSFADSNSPIRVTDRAAST (SEQ ID NO:3; Holwerda *et al.*, The Plant Cell, 4:307-318 (1992)) can be designed to be adjacent to the secretion signal or a sequence encoding a carboxyl signal peptide as described by Dombrowski *et al.*, The Plant Cell, 5:587-596 (1993) or a functional variation may be inserted at the 3' end of the gene. Similarly, VIP2 can be designed to be targeted to either the mitochondria or the plastids, including the chloroplasts, by inserting sequences in the VIP2 sequence described that would encode the required targeting signals. The bacterial secretion signal present in VIP2 may be retained or removed from the final construction.

One example of a construction which incorporates a eukaryotic secretion signal fused to a coding sequence for a VIP is provided by pCIB5528. Oligonucleotides corresponding to both the upper and lower strand of sequences encoding the secretion signal peptide of SEQ ID NO:25 was synthesized and has the sequence 5'-GGATCCACC ATG GGC TGG AGC TGG ATC TTC CTG TTC CTG AGC GGC GCC GCG GGC GTG CAC TGC CTGCAG-3' (SEQ ID NO:41). When hybridized, the 5' end of the secretion signal resembled "sticky-ends" corresponding to restriction sites BamHI and PstI. The oligonucleotide was hybridized and phosphorylated and ligated into pCIB5527 (construction described in Example 23A) which had been digested with BamHI/ PstI using standard procedures. The resulting maize optimized coding sequence is disclosed in SEQ ID NO:42 which encodes the protein disclosed

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in SEQ ID NO:43. This encoded protein comprises the eukaryotic secretion signal in place of the *Bacillus* secretion signal.

One example of a construction which incorporates a vacuolar targetting signal fused to a coding sequence for a VIP is provided by pCIB5533. Oligonucleotides corresponding to both the upper and lower strand of sequences encoding the vacuolar targetting peptide of SEQ ID NO:3 was synthesized and has the sequence 5'-CCG CGG GCG TGC ACT GCC TCA GCA GCA GCA GCT TCG CCG ACA GCA ACC CCA TCC GCG TGA CCG ACC GCG CCG CCA GCA CCC TGC AG-3' (SEQ ID NO:44). When hybridized, the 5' end of the vacuolar targetting signal resembled "sticky-ends" corresponding to restriction sites SacII and PstI. The oligonucleotide was hybridized and phosphorylated and ligated into pCIB5528 (construction described above) which had been digested with SacII / PstI using standard procedures. The resulting maize optimized coding sequence is disclosed in SEQ ID NO:45 which encodes the protein disclosed in SEQ ID NO:46. This encoded protein comprises the vacuolar targetting peptide in addition to the eukaryotic secretion signal.

The VIP1 gene can also be designed to be secreted or targeted to subcellular organelles by similar procedures.

**EXAMPLE 23A. REMOVAL OF BACILLUS SECRETION SIGNAL FROM
VIP1A(a) AND VIP2A(a)**

VIP1A(a) and VIP2A(a) are secreted during the growth of strain AB78. The nature of peptide sequences that act as secretion signals has been described in the literature (Simonen and Palva, Microbiological reviews, pg. 109-137 (1993)). Following the information in the above publication, the putative secretion signal was identified in both genes. In VIP1A(a) this signal is composed of amino acids 1-33 (See SEQ ID NO:5). Processing of the secretion signal probably occurs after the serine at amino acid 33. The secretion signal in VIP2A(a) was identified as amino acids 1-49 (See SEQ ID NO:2). N-terminal peptide analysis of the secreted mature VIP2A(a) protein revealed the N-terminal sequence LKITDKVEDFKEDK. This sequence is found beginning at amino acid 57 in SEQ ID NO:2. The genes encoding these proteins have been modified by removal of the *Bacillus* secretion signals.

A maize optimized VIP1A(a) coding region was constructed which had the sequences encoding the first 33 amino acids, i.e., the secretion signal, removed from its 5' end. This modification was obtained by PCR using an forward primer that

contained the sequence 5'-GGA TCC ACC ATG AAG ACC AAC CAG ATC AGC-3' (SEQ ID NO:33), which hybridizes with the maize optimized gene (SEQ ID NO:26) at nucleotide position 100, and added a BamHI restriction site and a eukaryotic translation start site consensus including a start codon. The reverse primer that contained the sequence 5'-AAG CTT CAG CTC CTT G-3' (SEQ ID NO:34) hybridizes on the complementary strand at nucleotide position 507. A 527 bp amplification product was obtained containing the restriction sites BamHI at the 5' end and HindIII site at the 3' end. The amplification product was cloned into a T- vector (described in Example 24, below) and sequenced to ensure the correct DNA sequence. The BamHI / HindIII fragment was then obtained by restriction digest and used to replace the BamHI/HindIII fragment of the maize optimized VIP1A(a) gene cloned in the root-preferred promoter cassette. The construct obtained was designated pCIB5526. The maize optimized coding region for VIP1A(a) with the *Bacillus* secretion signal removed is disclosed as SEQ ID NO:35 and the encoded protein is disclosed as SEQ ID NO:36.

The gene encoding the processed form of VIP2A(a), i.e., a coding region with the secretion signal removed, was constructed by a procedure similar to that described for that used to construct the processed form of VIP1A(a), above. The modification was obtained by PCR using the forward primer 5'-GGA TCC ACC ATG CTG CAG AAC CTG AAG ATC AC -3' (SEQ ID NO:37). This primer hybridizes at nucleotide position 150 of the maize optimized VIP2A(a) gene (SEQ ID NO:27). A silent mutation has been inserted at nucleotide position 15 of this primer to obtain a PstI restriction site. The reverse primer has the sequence 5'-AAG CTT CCA CTC CTT CTC-3' (SEQ ID NO:38). A 259 bp product was obtained with HindIII restriction site at the 3' end. The amplification product was cloned into a T- vector, sequenced and ligated to a BamHI /HindIII digested root-preferred promoter cassette containing the maize optimized VIP2A(a). The construct obtained was designated pCIB5527. The maize optimized coding region for VIP2A(a) with the *Bacillus* secretion signal removed is disclosed as SEQ ID NO:39 and the encoded protein is disclosed as SEQ ID NO:40.

**EXAMPLE 24. CONSTRUCTION AND CLONING OF THE VIP1A(a) AND VIP2A(a)
MAIZE OPTIMIZED GENES**

Design: The maize optimized genes were designed by reverse translation of the native VIP1A(a) and VIP2A(a) protein sequences using codons that are used most often in maize (Murray *et al.*, Nucleic Acid Research, 17:477-498 (1989)). To facilitate cloning, the DNA sequence was further modified to incorporate unique restriction sites at intervals of every 200-360 nucleotides. VIP1A(a) was designed to be cloned in 11 such fragments and VIP2A(a) was cloned in 5 fragments. Following cloning of the individual fragments, adjacent fragments were joined using the restriction sites common to both fragments, to obtain the complete gene. To clone each fragment, oligonucleotides (50-85 nucleotides) were designed to represent both the upper and the lower strand of the DNA. The upper oligo of the first oligo pair was designed to have a 15 bp single stranded region at the 3' end which was homologous to a similar single stranded region of the lower strand of the next oligo pair to direct the orientation and sequence of the various oligo pairs within a given fragment. The oligos are also designed such that when all the oligos representing a fragment are hybridized, the ends have single stranded regions corresponding to the particular restriction site to be formed. The structure of each oligomer was examined for stable secondary structures such as hairpin loops using the OLIGO program from NBI Inc. Whenever necessary, nucleotides were changed to decrease the stability of the secondary structure without changing the amino acid sequence of the protein. A plant ribosomal binding site consensus sequence, TAAACAATG (Joshi *et al.*, Nucleic Acid Res., 15:6643-6653 (1987)) or eukaryotic ribosomal binding site consensus sequence CCACCATG (Kozak, Nucleic Acid Research, 12:857-872 (1984)) was inserted at the translational start codon of the gene.

Cloning: Oligos were synthesized by IDT Inc., and were supplied as lyophilized powders. They were resuspended at a concentration of 200 µM. To 30 µl of each oligo formamide was added a final concentration of 25-50% and the sample was boiled for two minutes before separation on a premade 10% polyacrylamide / urea gel obtained from Novex. After electrophoresis, the oligo was detected by UV shadowing by placing the gel on a TLC plate containing a fluorescent indicator and exposing it to UV light. The region containing DNA of the correct size was excised and extracted

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from the polyacryamide by an overnight incubation of the minced gel fragment in a buffer containing 0.4 M LiCl, 0.1 mM EDTA. The DNA was separated from the gel residue by centrifugation through a Millipore UFMC filter. The extracted DNA was ethanol precipitated by the addition of 2 volumes of absolute alcohol. After centrifugation, the precipitate was resuspended in dH₂O at a concentration of 2.5 μM. Fragments were cloned either by hybridization of the oligos and ligation with the appropriate vector or by amplification of the hybridized fragment using a equimolar mixture of all the oligos for a particular fragment as a template and end-specific PCR primers.

Cloning by hybridization and ligation: Homologous double stranded oligo pairs were obtained by mixing 5 μl of the upper and of the lower oligo for each oligo pair with buffer containing 1X polynucleotide kinase (PNK) buffer (70 mM Tris-HCl (pH 7.6), 10 mM MgCl₂, 5 mM dithiothreitol (DTT)), 50 mM KCl, and 5 % formamide in a final volume of 50 μl. The oligos were boiled for 10 minutes and slow cooled to 37° C or room temperature. 10 μl was removed for analysis on a 4% agarose in a TAE buffer system (Metaphore®; FMC). Each hybridized oligo pair was kinased by the addition of ATP at a final concentration of 1 mM, BSA at a final concentration of 100 μg per ml and 200 units of polynucleotide kinase and 1 μl of 10X PNK buffer in a volume of 10 μl. Following hybridization and phosphorylation, the reaction was incubated at 37° C for 2 hours to overnight. 10 μl of each of the oligo pairs for a particular fragment, were mixed in a final volume of 50 μl. The oligo pairs were hybridized by heating at 80° C for 10 minutes and slow cooling to 37° C. 2 μl of oligos was mixed with about 100 ng of an appropriate vector and ligated using a buffer containing 50 mM Tris-HCl (pH 7.8), 10 mM MgCl₂, 10 mM DTT, 1 mM ATP. The reaction was incubated at room temp. for 2 hours to overnight and transformed into DH5α strain of *E.coli*, plated on L- plates containing ampicillin at a concentration of 100 μg/ml using standard procedures. Positive clones were further characterized and confirmed by PCR miniscreen described in detail in EP-A 0618976 using the universal primers "Reverse" and M13 "-20 " as primers. Positive clones were identified by digestion of DNA with appropriate enzymes followed by sequencing. Recombinants that had the expected DNA sequence were then selected for further work.

PCR Amplification and cloning into T- vector:

PCR amplification was carried out by using a mixture of all the oligomers that represented the upper and the lower strand of a particular fragment (final concentration 5 mM each) as template, specific end primers for the particular fragment (final concentration 2 μ M) 200 μ M of each dATP, dTTP, dCTP and dGTP, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl₂, 0.01% gelatin and 5 units of Taq polymerase in a final reaction volume of 50 μ l. The amplification reaction was carried out in a Perkin Elmer thermocycler 9600 by incubation at 95° C for 1 min (1 cycle), followed by 20 cycles of 95 °C for 45 sec., 50 °C for 45 sec., 72 °C for 30 sec. Finally the reaction was incubated for 5 min at 72°C before analyzing the product. 10 μ l of the reaction was analyzed on a 2.5% Nusieve (FMC) agarose gel in a TAE buffer system. The correct size fragment was gel purified and used for cloning into a PCR cloning vector or T-vector. T-vector construction was as described by Marchuk *et al.*, Nucleic Acid Research, 19:1154 (1991). pBluescriptsK+ (Stratagene®, Ca.) was used as the parent vector. Transformation and identification of the correct clone was carried out as described above.

Fragments 1, 3, 4, 5, 6, 8, and 9 of VIP1A(a) and fragments 2 and 4 of VIP2A(a) were obtained by cloning of PCR amplification products; whereas, fragments 2, 7, 10 and 11 of VIP1A(a) and fragments 1, 3, and 5 of VIP2A(a) were obtained by hybridization/ ligation.

Once fragments with the desired sequence were obtained, the complete gene was assembled by cloning together adjacent fragments. The complete gene was resequenced and tested for activity against WCRW before moving it into plant expression vectors containing the root preferred promoter (disclosed in U.S. patent application serial no. 08/017,209, herein incorporated by reference) and the rice actin promoter.

One such plant expression vector is pCIB5521. The maize optimized VIP1A(a) coding region (SEQ ID NO:26) was cloned in a plant expression vector containing the root preferred promoter at the 5' of the gene with the PEP Carboxylase intron #9 followed by the 35S terminator at the 3' end. The plasmid also contains sequences for ampicillin resistance from the plasmid pUC19. Another plant expression vector is pCIB5522, which contains the maize optimized VIP2A(a) coding region (SEQ ID

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NO:27) fused to the root preferred promoter at the 5' of the gene with the PEP Carboxylase intron #9 followed by the 35S terminator at the 3' end.

EXAMPLE 25. NAD AFFINITY CHROMATOGRAPHY

A purification strategy was used based on the affinity of VIP2 for the substrate NAD. The supernatant from the pH 3.5 sodium citrate buffer treatment described in Example 4 was dialyzed in 20 mM TRIS pH 7.5 overnight. The neutralized supernatant was added to an equal volume of washed NAD agarose and incubated with gentle rocking at 4° C overnight. The resin and protein solution were added to a 10 ml disposable polypropylene column and the protein solution allowed to flow out. The column was washed with 5 column volumes of 20 mM TRIS pH 7.5 then washed with 2-5 column volumes of 20 mM TRIS pH 7.5, 100 mM NaCl, followed by 2-5 column volumes of 20 mM TRIS 7.5. The VIP proteins were eluted in 20 mM TRIS pH 7.5 supplemented with 5 mM NAD. Approximately 3 column volumes of the effluent were collected and concentrated in a Centricon -10. Yield is typically about 7-15 µg of protein per ml of resin.

When the purified proteins were analyzed by SDS-PAGE followed by silver staining, two polypeptides were visible, one with Mr of approximately 80,000 and one with Mr of approximately 45,000. N-terminal sequencing revealed that the Mr 80,000 protein corresponded to a proteolytically processed form of VIP1A(A) and the Mr 45,000 form corresponded to a proteolytically processed form of VIP2A(a). The co-purification of VIP1A(a) with VIP2A(a) indicates that the two proteins probably form a complex and have protein-protein interacting regions. VIP1A(a) and VIP2A(a) proteins purified in this manner were biologically active against western corn rootworm.

EXAMPLE 26. EXPRESSION OF MAIZE OPTIMIZED VIP1A(a) AND VIP2A(a)

E. coli strains containing different plasmids comprising VIP genes were assayed for expression of VIPs. *E. coli* strains harboring the individual plasmids were grown overnight in L-broth and expressed protein was extracted from the culture as described in Example 3, above. Protein expression was assayed by Western Blot analysis using antibodies developed using standard methods known in the art, similar

to those described in Example 12, above. Also, insecticidal activity of the expressed proteins were tested against Western corn rootworm according to the method in Example 3, above. The results of the *E. coli* expression assays are described below.

Expression of VIPs in *E. coli*

Extract of <i>E. coli</i> Strain Harboring Indicated Plasmid	Assay No. 1	Assay No. 2	Protein Detected
% Mortality			
Control	0	0	no
pCIB5521 (maize optimized VIP1A(a))	47	27	yes
pCIB5522 (maize optimized VIP2A(a))	7	7	yes
pCIB6024 (native VIP2A(a))	13	13	yes
pCIB6206 (native VIP1A(a))	27	40	yes
Extracts pCIB5521 + pCIB5522 combined	87	47	
Extracts pCIB5521 + pCIB6024 combined	93	100	
Extracts pCIB5522 + pCIB6206 combined	100	100	
Extracts pCIB6024 + pCIB6206 combined	100	100	

The DNA from these plasmids was used to transiently express the VIPs in a maize protoplast expression system. Protoplasts were isolated from maize 2717 Line 6 suspension cultures by digestion of the cell walls using Cellulase RS and Macerase R10 in appropriate buffer. Protoplasts were recovered by sieving and centrifugation. Protoplasts were transformed by a standard direct gene transfer method using approximately 75 g plasmid DNA and PEG-40. Treated protoplasts were incubated overnight in the dark at room temperature. Analysis of VIP expression was

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accomplished on protoplast explants by Western blot analysis and insecticidal activity against Western corn rootworm as described above for the expression in *E. coli*. The results of the maize protoplast expression assays are described below.

Expression of VIPs in Plant Protoplasts

<i>Extract Tested</i>	<i>Assay No. 1</i>	<i>Assay No. 2</i>	<i>Protein Detected</i>
	% Mortality		
No DNA control	27	10	no
pCIB5521 (p) (maize optimized VIP1A(a))	20 (0)	30	yes
pCIB5522 (p) (maize optmized VIP2A(a))	20 (0)	20	yes
Extracts pCIB5521 (p) + pCIB5522 (p) combined	87 (82)	90	
Extracts pCIB5521 (p) + pCIB5522 (e) combined	100	-	
Extracts pCIB5522 (p) + pCIB5521 (e) combined	53 (36)	-	
Extracts pCIB5521 (p) + pCIB6024 (e) combined	100	-	
Extracts pCIB5522 (p) + pCIB6206 (e) combined	100	-	
pCIB6024(e) (native VIP2A(a))	0	-	yes
pCIB6206(e) (native VIP1A(a))	20	-	yes
pCIB5521 + pCIB 5522 (plasmids delivered by cotransformation)	100	100	yes

(p) = extract of protoplast culture transformed with indicated plasmid

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(e) = extract of *E. coli* strain harboring indicated plasmid

The expression data obtained with both *E. coli* and maize protoplasts show that the maize optimized VIP1A(a) and VIP2A(a) genes make the same protein as the native VIP1A(a) and VIP2A(a) genes, respectively, and that the proteins encoded by the maize optimized genes are functionally equivalent to the proteins encoded by the native genes.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

The following deposits have been made at Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA:

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Strain designation		Deposition Number	Deposition Date
1.	<i>E. coli</i> PL2	NRRL B-21221	March 09, 1994
2.	<i>E. coli</i> PL2	NRRL B-21221N	September 02, 1994
3.	<i>E. coli</i> pCIB6022	NRRL B-21222	March 09, 1994
4.	<i>E. coli</i> pCIB6023	NRRL B-21223	March 09, 1994
5.	<i>E. coli</i> pCIB6023	NRRL B-21223N	September 02, 1994
6.	<i>Bacillus thuringiensis</i> HD73-78VIP	NRRL B-21224	March 09, 1994
7.	<i>Bacillus thuringiensis</i> AB88	NRRL B-21225	March 09, 1994
8.	<i>Bacillus thuringiensis</i> AB359	NRRL B-21226	March 09, 1994
9.	<i>Bacillus thuringiensis</i> AB289	NRRL B-21227	March 09, 1994
10.	<i>Bacillus</i> sp. AB59	NRRL B-21228	March 09, 1994
11.	<i>Bacillus</i> sp. AB294	NRRL B-21229	March 09, 1994
12.	<i>Bacillus</i> sp. AB256	NRRL B-21230	March 09, 1994
13.	<i>E. coli</i> P5-4	NRRL B-21059	March 18, 1993
14.	<i>E. coli</i> P3-12	NRRL B-21061	March 18, 1993
15.	<i>Bacillus cereus</i> AB78	NRRL B-21058	March 18, 1993
16.	<i>Bacillus thuringiensis</i> AB6	NRRL B-21060	March 18, 1993
17.	<i>E. coli</i> pCIB6202	NRRL B-21321	September 02, 1994
18.	<i>E. coli</i> pCIB7100	NRRL B-21322	September 02, 1994
19.	<i>E. coli</i> pCIB7101	NRRL B-21323	September 02, 1994
20.	<i>E. coli</i> pCIB7102	NRRL B-21324	September 02, 1994
21.	<i>E. coli</i> pCIB7103	NRRL B-21325	September 02, 1994
22.	<i>E. coli</i> pCIB7104	NRRL B-21422	March 24, 1995
23.	<i>E. coli</i> pCIB7107	NRRL B-21423	March 24, 1995
24.	<i>E. coli</i> pCIB7108	NRRL B-21438	May 05, 1995
25.	<i>Bacillus thuringiensis</i> AB424	NRRL B-21439	May 05, 1995

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Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (A) NAME: CIBA-GEIGY AG
- (B) STREET: Klybeckstr. 141
- (C) CITY: Basel
- (E) COUNTRY: Switzerland
- (F) POSTAL CODE (ZIP): 4002
- (G) TELEPHONE: +41 61 69 11 11
- (H) TELEFAX: + 41 61 696 79 76
- (I) TELEX: 962 991

(ii) TITLE OF INVENTION: Novel Pesticidal Proteins and Strains

(iii) NUMBER OF SEQUENCES: 52

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30B

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6049 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Bacillus cereus
- (B) STRAIN: AB78
- (C) INDIVIDUAL ISOLATE: NRRL B-21058

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1082..2467
- (D) OTHER INFORMATION: /product= "VIP2A(a)"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 2475..5126

(D) OTHER INFORMATION: /note= "Coding sequence for the 100 kd VIP1A(a) protein. This coding sequence is repeated in SEQ ID NO:4 and translated separately."

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATCGATACAA TGTTGTTTA CTTAGACCGG TAGTCTCTGT AATTTGTTA ATGCTATATT	60
CTTACTTTG ATACATTTA ATAGCCATT CAACCTTATC AGTATGTTT TGTGGCTTC	120
CTCCTTTT TCCACGAGCT CTAGCTGCGT TTAATCCTGT TTTGGTACGT TCGCTAATAA	180
TATCTCTTTC TAATTCTGCA ATACTTGCCA TCATTGAAA GAAGAATTTC CCCATAGCAT	240
TAGAGGTATC AATGTTGTCA TGAATAGAAA TAAAATCTAC ACCTAGCTCT TTGAATTTT	300
CACTTAACTC AATTAGGTGT TTTGTAGAGC GAGAAATTG ATCAAGTTG TAAACAACTA	360
TCTTATCGCC TTTACGTAAT ACTTTAGCA ACTCTTCGAG TTGAGGGCGC TCTTTTTTA	420
TTCCTGTTAT TTTCTCCTGA TATAGCCTTT CTACACCATA TTGTTGCAA GCATCTATT	480
GCATATCGAG ATTTTGTCT TCTGTGCTGA CACGAGCATA ACCAAAAATC AAATTGGTTT	540
CACTCCCTAT CTAAATATAT CTATTAAAAT AGCACCAAAA ACCTTATTAA ATTAAAATAA	600
GGAACTTTGT TTTGGATAT GGATTTGGT ACTCAATATG GATGAGTTT TAACGCTTTT	660
GTTAAAAAAC AAACAAGTGC CATAAACGGT CGTTTTGGG ATGACATAAT AAATAATCTG	720
TTTGATTAAC CTAACCTTGT ATCCTTACAG CCCAGTTTA TTTGTACTTC AACTGACTGA	780
ATATGAAAAC AACATGAAGG TTTCATAAAA TTTATATATT TTCCATAACG GATGCTCTAT	840
CTTTAGGTTA TAGITAAATT ATAAGAAAAA AACAAACGGA GGGAGTGAAA AAAAGCATCT	900
TCTCTATAAT TTTACAGGCT CTTTAATAAG AAGGGGGAG ATTAGATAAT AAATATGAAT	960
ATCTATCTAT AATTGTTGC TTCTACAATA ACTTATCTAA CTTTCATATA CAACAACAAA	1020
ACAGACTAAA TCCAGATTGT ATATTCAATT TCAGTTGTTC CTTTATAAAA TAATTTCATA	1080
A ATG AAA AGA ATG GAG GGA AAG TTG TTT ATG GTG TCA AAA AAA TTA Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys Leu	1126
1 5 10 15	
CAA GTA GTT ACT AAA ACT GTA TTG CTT AGT ACA GTT TTC TCT ATA TCT Gln Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile Ser	1174
20 25 30	
TTA TTA AAT AAT GAA GTG ATA AAA GCT GAA CAA TTA AAT ATA AAT TCT Leu Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn Ser	1222
35 40 45	
CAA AGT AAA TAT ACT AAC TTG CAA AAT CTA AAA ATC ACT GAC AAG GTA Gln Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Thr Asp Lys Val	1270
50 55 60	
GAG GAT TTT AAA GAA GAT AAG GAA AAA GCG AAA GAA TGG GGG AAA GAA	1318

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Glu Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu		
65	70	75
AAA GAA AAA GAG TGG AAA CTA ACT GCT ACT GAA AAA GGA AAA ATG AAT		1366
Lys Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn		
80	85	90
95		
AAT TTT TTA GAT AAT AAA AAT GAT ATA AAG ACA AAT TAT AAA GAA ATT		1414
Asn Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile		
100	105	110
ACT TTT TCT ATG GCA GGC TCA TTT GAA GAT GAA ATA AAA GAT TTA AAA		1462
Thr Phe Ser Met Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys		
115	120	125
GAA ATT GAT AAG ATG TTT GAT AAA ACC AAT CTA TCA AAT TCT ATT ATC		1510
Glu Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile		
130	135	140
ACC TAT AAA AAT GTG GAA CCG ACA ACA ATT GGA TTT AAT AAA TCT TTA		1558
Thr Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu		
145	150	155
ACA GAA GGT AAT ACG ATT AAT TCT GAT GCA ATG GCA CAG TTT AAA GAA		1606
Thr Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu		
160	165	170
175		
CAA TTT TTA GAT AGG GAT ATT AAG TTT GAT AGT TAT CTA GAT ACG CAT		1654
Gln Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His		
180	185	190
TTA ACT GCT CAA CAA GTT TCC AGT AAA GAA AGA GTT ATT TTG AAG GTT		1702
Leu Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val		
195	200	205
ACG GTT CCG AGT GGG AAA GGT TCT ACT ACT CCA ACA AAA GCA GGT GTC		1750
Thr Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val		
210	215	220
ATT TTA AAT AAT AGT GAA TAC AAA ATG CTC ATT GAT AAT GGG TAT ATG		1798
Ile Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met		
225	230	235
GTC CAT GTA GAT AAG GTA TCA AAA GTG GTG AAA AAA GGG GTG GAG TGC		1846
Val His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys		
240	245	250
255		
TTA CAA ATT GAA GGG ACT TTA AAA AAG AGT CTT GAC TTT AAA AAT GAT		1894
Leu Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp		
260	265	270
ATA AAT GCT GAA GCG CAT AGC TGG GGT ATG AAG AAT TAT GAA GAG TGG		1942
Ile Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp		
275	280	285

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GCT AAA GAT TTA ACC GAT TCG CAA AGG GAA GCT TTA GAT GGG TAT GCT Ala Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala 290 295 300	1990
AGG CAA GAT TAT AAA GAA ATC AAT AAT TAT TTA AGA AAT CAA GGC GGA Arg Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly 305 310 315	2038
AGT GGA AAT GAA AAA CTA GAT GCT CAA ATA AAA AAT ATT TCT GAT GCT Ser Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala 320 325 330 335	2086
TTA GGG AAG AAA CCA ATA CCG GAA AAT ATT ACT GTG TAT AGA TGG TGT Leu Gly Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys 340 345 350	2134
GGC ATG CCG GAA TTT GGT TAT CAA ATT AGT GAT CCG TTA CCT TCT TTA Gly Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu 355 360 365	2182
AAA GAT TTT GAA GAA CAA TTT TTA AAT ACA ATC AAA GAA GAC AAA GGA Lys Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly 370 375 380	2230
TAT ATG AGT ACA AGC TTA TCG AGT GAA CGT CTT GCA GCT TTT GGA TCT Tyr Met Ser Thr Ser Leu Ser Glu Arg Leu Ala Ala Phe Gly Ser 385 390 395	2278
AGA AAA ATT ATA TTA CGA TTA CAA GTT CCG AAA GGA AGT ACG GGT GCG Arg Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala 400 405 410 415	2326
TAT TTA AGT GCC ATT GGT GGA TTT GCA AGT GAA AAA GAG ATC CTA CTT Tyr Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu 420 425 430	2374
GAT AAA GAT AGT AAA TAT CAT ATT GAT AAA GTA ACA GAG GTA ATT ATT Asp Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile 435 440 445	2422
AAA GGT GTT AAG CGA TAT GTA GTG GAT GCA ACA TTA TTA ACA AAT Lys Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn 450 455 460	2467
TAAGGAGATG AAAATATGA AGAAAAAGTT AGCAAGTGTGTT GTAACGTGTA CGTTATTAGC	2527
TCCTATGTT TTGAATGGAA ATGTGAATGC TGTTTACGCA GACAGCAAAA CAAATCAAAT	2587
TTCTACAACA CAGAAAAATC AACAGAAAGA GATGGACCGA AAAGGATTAC TTGGGTATTA	2647
TTTCAAAGGA AAAGATTTA GTAATCTTAC TATGTTGCA CCGACACGTG ATAGTACTCT	2707
TATTTATGAT CAACAAACAG CAAATAACT ATTAGATAAA AAACAACAAG AATATCAGTC	2767
TATTCGTTGG ATTGGTTGA TTCAGAGTAA AGAAACGGGA GATTCACAT TTAACCTATC	2827

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TGAGGGATGAA CAGGCAATTAA TAGAAATCAA TGGAAAATT ATTTCTAATA AAGGGAAAGA	2887
AAAGCAAGTT GTCCATTAG AAAAAGGAAA ATTAGTTCGA ATCAAAATAG AGTATCAATC	2947
AGATACAAAAA TTAAATATTG ACAGTAAAAC ATTTAAAGAA CTTAAATTAT TTAAATAGA	3007
TAGTCAAAAC CAACCCCAGC AAGTCCAGCA AGATGAACTG AGAAATCCTG AATTTAACAA	3067
GAAAGAATCA CAGGAATTCT TAGCGAAACC ATCGAAAATA AATCTTTCA CTAAAAAAT	3127
GAAAAGGGAA ATTGATGAAG ACACGGATAC GGATGGGAC TCTATTCTG ACCTTTGGGA	3187
AGAAAATGGG TATACGATTC ACAATAGAAT CGCTGTAAAG TGGGACGATT CTCTAGCAAG	3247
TAAAGGTAT ACGAAATTG TTTCAAATCC ACTAGAAAGT CACACAGTTG GTGATCCTTA	3307
TACAGATTAT GAAAAGGCAG CAAGAGATCT AGATTGTCA AATGCAAAGG AAACGTTAA	3367
CCCATTGGTA GCTGCTTTTC CAAGTGTGAA TGTTAGTATG GAAAAGGTGA TATTATCACC	3427
AAATGAAAAT TTATCCAATA GTGTAGAGTC TCATTCAATCC ACGAATTGGT CTTATACAAA	3487
TACAGAAGGT GCTTCTGTG AAGCGGGGAT TGGACCAAAA GGTATTCGT TCGGAGTTAG	3547
CGTAAACTAT CAACACTCTG AAACAGTTGC ACAAGAATGG GGAACATCTA CAGGAAATAC	3607
TTCGCAATTTC AATACGGCTT CAGCGGGATA TTAAATGCA AATGTTCGAT ATAACAATGT	3667
AGGAACGTGGT GCCATCTACG ATGTAAAACC TACAACAAGT TTTGTATTAA ATAACGATAC	3727
TATCGCAACT ATTACGGCGA AATCTAATTTC TACAGCCTTA AATATATCTC CTGGAGAAAG	3787
TTACCCGAAA AAAGGACAAA ATGGAATCGC AATAACATCA ATGGATGATT TTAATTCCCA	3847
TCCGATTACA TTAAATAAAA ACAAGTAGA TAATCTGCTA AATAATAAAC CTATGATGTT	3907
GGAAACAAAC CAAACAGATG GTGTTTATAA GATAAAAGAT ACACATGGAA ATATAGTAAC	3967
TGGCGGAGAA TGGAATGGTG TCATACAACA AATCAAGGCT AAAACAGCGT CTATTATTGT	4027
GGATGATGGG GAACGTGTAG CAGAAAAACG TGTAGCGGCA AAAGATTATG AAAATCCAGA	4087
AGATAAAACA CCGTCTTAA CTTTAAAAGA TGCCCTGAAG CTTTCATATC CAGATGAAAT	4147
AAAAGAAATA GAGGGATTAT TATATTATAA AAACAAACCG ATATACGAAT CGAGCGTTAT	4207
GACTTACTTA GATGAAAATA CAGCAAAAGA AGTGCACAAA CAATTAAATG ATACCACTGG	4267
GAAATTAAA GATGTAAGTC ATTTATATGA TGAAAATG ACTCCAAAAA TGAATGTTAC	4327
AATCAAATTG TCTATACTTT ATGATAATGC TGAGTCTAAT GATAACTCAA TTGGTAAATG	4387
GACAAACACA AATATTGTTT CAGGTGGAAA TAACGGAAAA AAACAATATT CTTCTAATAA	4447

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TCCGGATGCT AATTGACAT TAAATACAGA TGCTCAAGAA AAATTAATA AAAATCGTGA	4507
CTATTATATA AGTTTATATA TGAAAGTCAGA AAAAAACACA CAATGTGAGA TTACTATAGA	4567
TGGGGAGATT TATCCGATCA CTACAAAAAC AGTGAATGTG AATAAAGACA ATTACAAAAG	4627
ATTAGATATT ATAGCTCATA ATATAAAAAG TAATCCAATT TCTTCACTTC ATATTAAAAC	4687
GAATGATGAA ATAACTTAT TTTGGGATGA TATTCTATA ACAGATGTAG CATCAATAAA	4747
ACCGGAAAAT TTAACAGATT CAGAAATTAA ACAGATTTAT AGTAGGTATG GTATTAAGTT	4807
AGAAGATGGA ATCCTTATTG ATAAAAAAGG TGGGATTCAT TATGGTGAAT TTATTAATGA	4867
AGCTAGTTT AATATTGAAC CATTGCAAAA TTATGTGACC AAATATGAAG TTACTTATAG	4927
TAGTGAGTTA GGACCAAACG TGAGTGACAC ACTTGAAAGT GATAAAATT ACAAGGATGG	4987
GACAATTAAA TTTGATTTA CCAAATATAG TAAAAATGAA CAAGGATTAT TTTATGACAG	5047
TGGATTAAT TGGGACTTTA AAATTAATGC TATTACTTAT GATGGTAAAG AGATGAATGT	5107
TTTCATAGA TATAATAAT AGTTATTATA TCTATGAAGC TGGTGCTAAA GATAGTGTAA	5167
AAGTTAATAT ACTGTAGGAT TGTAATAAAA GTAATGGAAT TGATATCGTA CTTGGAGTG	5227
GGGGATACTT TGTAATAGT TCTATCAGAA ACATTAGACT AAGAAAAGTT ACTACCCCCA	5287
CTTGAAAATG AAGATTCAAC TGATTACAAA CAACCTGTTA AATATTATAA GGTTTAACCA	5347
AAATATTAAA CTCTTATGT TAATACTGTA ATATAAAGAG TTTAATTGTA TTCAAATGAA	5407
GCTTCCCAC AAAATTAGAC TGATTATCTA ATGAAATAAT CAGTCTAATT TTGTAGAACCA	5467
GGTCTGGTAT TATTGTACGT GGTCACTAAA AGATATCTAA TATTATTGGG CAAGGCCTTC	5527
CATGATTGAA TCCTCGAATG TCTGCCCTT TTCATTATT TAAGAAGGAT TGTGGAGAAA	5587
TTATGGTTA GATAATGAAG AAAGACTTC A TTCTAATT TTGATGTTAA ATAAATCAA	5647
ATTTGGCGAT TCACATTGTT TAATCCACTG ATAAAACATA CTGGAGTGT CTTAAAAAAT	5707
CAGCTTTTTT CTTTATAAAA TTTTGCTTAG CGTACGAAAT TCGTGTGTTG TTGGTGGGAC	5767
CCCATGCCA TCAACTTAAG AGTAAATTAG TAATGAACCT TCGTTCATCT GGATTAAAAT	5827
AACCTCAAAT TAGGACATGT TTTTAAAAAT AAGCAGACCA AATAAGCCTA GAATAGGTAT	5887
CATTTTTAAA AATTATGCTG CTTTCTTTG TTTTCCAAAT CCATTATACT CATAAGCAAC	5947
ACCCATAATG TCAAAGACTG TTTTGTCTC ATATCGATAA GCTTGATATC GAATTCTGC	6007
AGCCCGGGGG ATCCACTAGT TCTAGAGCGG CCGCCACCGC GG	6049

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 462 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys Leu Gln
1 5 10 15

Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile Ser Leu
20 25 30

Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn Ser Gln
35 40 45

Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu
50 55 60

Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys
65 70 75 80

Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn
85 90 95

Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr
100 105 110

Phe Ser Met Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu
115 120 125

Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr
130 135 140

Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr
145 150 155 160

Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln
165 170 175

Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu
180 185 190

Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr
195 200 205

Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile
210 215 220

Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val
225 230 235 240

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His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu
245 250 255

Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile
260 265 270

Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala
275 280 285

Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg
290 295 300

Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser
305 310 315 320

Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu
325 330 335

Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly
340 345 350

Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys
355 360 365

Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr
370 375 380

Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg
385 390 395 400

Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr
405 410 415

Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp
420 425 430

Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys
435 440 445

Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn
450 455 460

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

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(A) NAME/KEY: Peptide
 (B) LOCATION: 1..20
 (D) OTHER INFORMATION: /note= "Signal peptide for vacuolar targetting"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Ser	Ser	Ser	Ser	Phe	Ala	Asp	Ser	Asn	Pro	Ile	Arg	Val	Thr	Asp	Arg
1					5					10				15	
Ala	Ala	Ser	Thr												
			20												

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2655 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Bacillus cereus
 (B) STRAIN: AB78
 (C) INDIVIDUAL ISOLATE: NRRL B-21058

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 1..2652
 (D) OTHER INFORMATION: /product= "100 kDa protein VIP1A(a)"
 /note= "This sequence is identical to the portion of SEQ ID NO:1
 between and including nucleotide 2475 to 5126."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

ATG	AAA	AAT	ATG	AAG	AAG	TTA	GCA	AGT	GTT	GTA	ACG	TGT	ACG	TTA	48
Met			Met	Lys	Lys	Lys	Lys			Leu	Ala	Ser	Val	Val	
465									470					475	
TTA	GCT	CCT	ATG	TTT	TTG	AAT	GGA	AAT	GTG	AAT	GCT	GTG	TAC	GCA	96
Leu	Ala	Pro	Met	Phe	Leu	Asn	Gly	Asn	Val	Asn	Ala	Val	Tyr	Ala	
480							485						490		
AGC	AAA	ACA	AAT	CAA	ATT	TCT	ACA	ACA	CAG	AAA	AAT	CAA	CAG	AAA	144
Ser															
495								500						510	

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ATG GAC CGA AAA GGA TTA CTT GGG TAT TAT TTC AAA GGA AAA GAT TTT Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys Asp Phe 515 520 525	192
AGT AAT CTT ACT ATG TTT GCA CCG ACA CGT GAT AGT ACT CTT ATT TAT Ser Asn Leu Thr Met Phe Ala Pro Thr Arg Asp Ser Thr Leu Ile Tyr 530 535 540	240
GAT CAA CAA ACA GCA AAT AAA CTA TTA GAT AAA AAA CAA CAA GAA TAT Asp Gln Gln Thr Ala Asn Lys Leu Leu Asp Lys Lys Gln Gln Glu Tyr 545 550 555	288
CAG TCT ATT CGT TGG ATT GGT TTG ATT CAG AGT AAA GAA ACG GGA GAT Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln Ser Lys Glu Thr Gly Asp 560 565 570	336
TTC ACA TTT AAC TTA TCT GAG GAT GAA CAG GCA ATT ATA GAA ATC AAT Phe Thr Phe Asn Leu Ser Glu Asp Glu Gln Ala Ile Ile Glu Ile Asn 575 580 585 590	384
GGG AAA ATT ATT TCT AAT AAA GGG AAA GAA AAG CAA GTT GTC CAT TTA Gly Lys Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val His Leu 595 600 605	432
GAA AAA GGA AAA TTA GTT CCA ATC AAA ATA GAG TAT CAA TCA GAT ACA Glu Lys Gly Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser Asp Thr 610 615 620	480
AAA TTT AAT ATT GAC AGT AAA ACA TTT AAA GAA CTT AAA TTA TTT AAA Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu Phe Lys 625 630 635	528
ATA GAT AGT CAA AAC CAA CCC CAG CAA GTC CAG CAA GAT GAA CTG AGA Ile Asp Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu Leu Arg 640 645 650	576
AAT CCT GAA TTT AAC AAG AAA GAA TCA CAG GAA TTC TTA GCG AAA CCA Asn Pro Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Pro 655 660 665 670	624
TCG AAA ATA AAT CTT TTC ACT CAA AAA ATG AAA AGG GAA ATT GAT GAA Ser Lys Ile Asn Leu Phe Thr Gln Lys Met Lys Arg Glu Ile Asp Glu 675 680 685	672
GAC ACG GAT ACG GAT GGG GAC TCT ATT CCT GAC CTT TGG GAA GAA AAT Asp Thr Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn 690 695 700	720
GGG TAT ACG ATT CAA AAT AGA ATC GCT GTA AAG TGG GAC GAT TCT CTA Gly Tyr Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp Ser Leu 705 710 715	768
GCA AGT AAA GGG TAT ACG AAA TTT GTT TCA AAT CCA CTA GAA AGT CAC Ala Ser Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu Ser His 720 725 730	816

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ACA GTT GGT GAT CCT TAT ACA GAT TAT GAA AAG GCA GCA AGA GAT CTA Thr Val Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu 735 740 745 750	864
GAT TTG TCA AAT GCA AAG GAA ACG TTT AAC CCA TTG GTA GCT GCT TTT Asp Leu Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe 755 760 765	912
CCA AGT GTG AAT GTT AGT ATG GAA AAG GTG ATA TTA TCA CCA AAT GAA Pro Ser Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu 770 775 780	960
AAT TTA TCC AAT AGT GTA GAG TCT CAT TCA TCC ACG AAT TGG TCT TAT Asn Leu Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr 785 790 795	1008
ACA AAT ACA GAA GGT GCT TCT GTT GAA GCG GGG ATT GGA CCA AAA GGT Thr Asn Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro Lys Gly 800 805 810	1056
ATT TCG TTC GGA GTT AGC GTA AAC TAT CAA CAC TCT GAA ACA GTT GCA Ile Ser Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr Val Ala 815 820 825 830	1104
CAA GAA TGG GGA ACA TCT ACA GGA AAT ACT TCG CAA TTC AAT ACG GCT Gln Glu Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala 835 840 845	1152
TCA GCG GGA TAT TTA AAT GCA AAT GTT CGA TAT AAC AAT GTA GGA ACT Ser Ala Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr 850 855 860	1200
GGT GCC ATC TAC GAT GTA AAA CCT ACA ACA AGT TTT GTA TTA AAT AAC Gly Ala Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn 865 870 875	1248
GAT ACT ATC GCA ACT ATT ACG GCG AAA TCT AAT TCT ACA GCC TTA AAT Asp Thr Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Asn 880 885 890	1296
ATA TCT CCT GGA GAA AGT TAC CCG AAA AAA GGA CAA AAT GGA ATC GCA Ile Ser Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly Ile Ala 895 900 905 910	1344
ATA ACA TCA ATG GAT GAT TTT AAT TCC CAT CCG ATT ACA TTA AAT AAA Ile Thr Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys 915 920 925	1392
AAA CAA GTA GAT AAT CTG CTA AAT AAT AAA CCT ATG ATG TTG GAA ACA Lys Gln Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu Glu Thr 930 935 940	1440
AAC CAA ACA GAT GGT GTT TAT AAG ATA AAA GAT ACA CAT GGA AAT ATA Asn Gln Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly Asn Ile	1488

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945	950	955	
GTA ACT GGC GGA GAA TGG AAT GGT GTC ATA CAA CAA ATC AAG GCT AAA Val Thr Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys Ala Lys 960 965 970			1536
ACA GCG TCT ATT ATT GTG GAT GAT GGG GAA CGT GTA GCA GAA AAA CGT Thr Ala Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu Lys Arg 975 980 985 990			1584
GTA GCG GCA AAA GAT TAT GAA AAT CCA GAA GAT AAA ACA CCG TCT TTA Val Ala Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro Ser Leu 995 1000 1005			1632
ACT TTA AAA GAT GCC CTG AAG CTT TCA TAT CCA GAT GAA ATA AAA GAA Thr Leu Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu 1010 1015 1020			1680
ATA GAG GGA TTA TTA TAT TAT AAA AAC AAA CCG ATA TAC GAA TCG AGC Ile Glu Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu Ser Ser 1025 1030 1035			1728
GTT ATG ACT TAC TTA GAT GAA AAT ACA GCA AAA GAA GTG ACC AAA CAA Val Met Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr Lys Gln 1040 1045 1050			1776
TTA AAT GAT ACC ACT GGG AAA TTT AAA GAT GTA AGT CAT TTA TAT GAT Leu Asn Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu Tyr Asp 1055 1060 1065 1070			1824
GTA AAA CTG ACT CCA AAA ATG AAT GTT ACA ATC AAA TTG TCT ATA CTT Val Lys Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser Ile Leu 1075 1080 1085			1872
TAT GAT AAT GCT GAG TCT AAT GAT AAC TCA ATT GGT AAA TGG ACA AAC Tyr Asp Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp Thr Asn 1090 1095 1100			1920
ACA AAT ATT GTT TCA GGT GGA AAT AAC GGA AAA AAA CAA TAT TCT TCT Thr Asn Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr Ser Ser 1105 1110 1115			1968
AAT AAT CCG GAT GCT AAT TTG ACA TTA AAT ACA GAT GCT CAA GAA AAA Asn Asn Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln Glu Lys 1120 1125 1130			2016
TTA AAT AAA AAT CGT GAC TAT TAT ATA AGT TTA TAT ATG AAG TCA GAA Leu Asn Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys Ser Glu 1135 1140 1145 1150			2064
AAA AAC ACA CAA TGT GAG ATT ACT ATA GAT GGG GAG ATT TAT CCG ATC Lys Asn Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr Pro Ile 1155 1160 1165			2112
ACT ACA AAA ACA GTG AAT GTG AAT AAA GAC AAT TAC AAA AGA TTA GAT			2160

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Thr Thr Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg Leu Asp		1170	1175	1180	
ATT ATA GCT CAT AAT ATA AAA AGT AAT CCA ATT TCT TCA CTT CAT ATT					2208
Ile Ile Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu His Ile					
1185	1190	1195			
AAA ACG AAT GAT GAA ATA ACT TTA TTT TGG GAT GAT ATT TCT ATA ACA					2256
Lys Thr Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser Ile Thr					
1200	1205	1210			
GAT GTA GCA TCA ATA AAA CCG GAA AAT TTA ACA GAT TCA GAA ATT AAA					2304
Asp Val Ala Ser Ile Lys Pro Glu Asn Leu Thr Asp Ser Glu Ile Lys					
1215	1220	1225	1230		
CAG ATT TAT AGT AGG TAT GGT ATT AAG TTA GAA GAT GGA ATC CTT ATT					2352
Gln Ile Tyr Ser Arg Tyr Gly Ile Lys Leu Glu Asp Gly Ile Leu Ile					
1235	1240	1245			
GAT AAA AAA GGT GGG ATT CAT TAT GGT GAA TTT ATT AAT GAA GCT AGT					2400
Asp Lys Lys Gly Gly Ile His Tyr Gly Glu Phe Ile Asn Glu Ala Ser					
1250	1255	1260			
TTT AAT ATT GAA CCA TTG CAA AAT TAT GTG ACC AAA TAT GAA GTT ACT					2448
Phe Asn Ile Glu Pro Leu Gln Asn Tyr Val Thr Lys Tyr Glu Val Thr					
1265	1270	1275			
TAT AGT AGT GAG TTA GGA CCA AAC GTG AGT GAC ACA CTT GAA AGT GAT					2496
Tyr Ser Ser Glu Leu Gly Pro Asn Val Ser Asp Thr Leu Glu Ser Asp					
1280	1285	1290			
AAA ATT TAC AAG GAT GGG ACA ATT AAA TTT GAT TTT ACC AAA TAT AGT					2544
Lys Ile Tyr Lys Asp Gly Thr Ile Lys Phe Asp Phe Thr Lys Tyr Ser					
1295	1300	1305	1310		
AAA AAT GAA CAA GGA TTA TTT TAT GAC AGT GGA TTA AAT TGG GAC TTT					2592
Lys Asn Glu Gln Gly Leu Phe Tyr Asp Ser Gly Leu Asn Trp Asp Phe					
1315	1320	1325			
AAA ATT AAT GCT ATT ACT TAT GAT GGT AAA GAG ATG AAT GTT TTT CAT					2640
Lys Ile Asn Ala Ile Thr Tyr Asp Gly Lys Glu Met Asn Val Phe His					
1330	1335	1340			
AGA TAT AAT AAA TAG					2655
Arg Tyr Asn Lys					
1345					

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 884 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Lys Asn Met Lys Lys Leu Ala Ser Val Val Thr Cys Thr Leu
1 5 10 15

Leu Ala Pro Met Phe Leu Asn Gly Asn Val Asn Ala Val Tyr Ala Asp
20 25 30

Ser Lys Thr Asn Gln Ile Ser Thr Thr Gln Lys Asn Gln Gln Lys Glu
35 40 45

Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys Asp Phe
50 55 60

Ser Asn Leu Thr Met Phe Ala Pro Thr Arg Asp Ser Thr Leu Ile Tyr
65 70 75 80

Asp Gln Gln Thr Ala Asn Lys Leu Leu Asp Lys Lys Gln Gln Glu Tyr
85 90 95

Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln Ser Lys Glu Thr Gly Asp
100 105 110

Phe Thr Phe Asn Leu Ser Glu Asp Glu Gln Ala Ile Ile Glu Ile Asn
115 120 125

Gly Lys Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val His Leu
130 135 140

Glu Lys Gly Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser Asp Thr
145 150 155 160

Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu Phe Lys
165 170 175

Ile Asp Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu Leu Arg
180 185 190

Asn Pro Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Pro
195 200 205

Ser Lys Ile Asn Leu Phe Thr Gln Lys Met Lys Arg Glu Ile Asp Glu
210 215 220

Asp Thr Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn
225 230 235 240

Gly Tyr Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp Ser Leu
245 250 255

Ala Ser Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu Ser His
260 265 270

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Thr Val Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu
275 280 285

Asp Leu Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe
290 295 300

Pro Ser Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu
305 310 315 320

Asn Leu Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr
325 330 335

Thr Asn Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro Lys Gly
340 345 350

Ile Ser Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr Val Ala
355 360 365

Gln Glu Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala
370 375 380

Ser Ala Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr
385 390 395 400

Gly Ala Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn
405 410 415

Asp Thr Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Asn
420 425 430

Ile Ser Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly Ile Ala
435 440 445

Ile Thr Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys
450 455 460

Lys Gln Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu Glu Thr
465 470 475 480

Asn Gln Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly Asn Ile
485 490 495

Val Thr Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys Ala Lys
500 505 510

Thr Ala Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu Lys Arg
515 520 525

Val Ala Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro Ser Leu
530 535 540

Thr Leu Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu
545 550 555 560

Ile Glu Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu Ser Ser

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565	570	575
Val Met Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr Lys Gln 580	585	590
Leu Asn Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu Tyr Asp 595	600	605
Val Lys Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser Ile Leu 610	615	620
Tyr Asp Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp Thr Asn 625	630	635
Thr Asn Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr Ser Ser 645	650	655
Asn Asn Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln Glu Lys 660	665	670
Leu Asn Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys Ser Glu 675	680	685
Lys Asn Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr Pro Ile 690	695	700
Thr Thr Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg Leu Asp 705	710	720
Ile Ile Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu His Ile 725	730	735
Lys Thr Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser Ile Thr 740	745	750
Asp Val Ala Ser Ile Lys Pro Glu Asn Leu Thr Asp Ser Glu Ile Lys 755	760	765
Gln Ile Tyr Ser Arg Tyr Gly Ile Lys Leu Glu Asp Gly Ile Leu Ile 770	775	780
Asp Lys Lys Gly Gly Ile His Tyr Gly Glu Phe Ile Asn Glu Ala Ser 785	790	800
Phe Asn Ile Glu Pro Leu Gln Asn Tyr Val Thr Lys Tyr Glu Val Thr 805	810	815
Tyr Ser Ser Glu Leu Gly Pro Asn Val Ser Asp Thr Leu Glu Ser Asp 820	825	830
Lys Ile Tyr Lys Asp Gly Thr Ile Lys Phe Asp Phe Thr Lys Tyr Ser 835	840	845
Lys Asn Glu Gln Gly Leu Phe Tyr Asp Ser Gly Leu Asn Trp Asp Phe 850	855	860

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Lys Ile Asn Ala Ile Thr Tyr Asp Gly Lys Glu Met Asn Val Phe His
 865 870 875 880

Arg Tyr Asn Lys

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2004 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Bacillus cereus*
- (B) STRAIN: AB78
- (C) INDIVIDUAL ISOLATE: NRRL B-21058

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..2001

(D) OTHER INFORMATION: /product= "80 kDa protein VIP1A(a)"

/note= "This sequence is identical to that found in SEQ ID NO:1 between and including nucleotide positions 3126 and 5126"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATG AAA AGG GAA ATT GAT GAA GAC ACG GAT ACG GAT GGG GAC TCT ATT	48
Met Lys Arg Glu Ile Asp Glu Asp Thr Asp Thr Asp Gly Asp Ser Ile	
885 890 895 900	

CCT GAC CTT TGG GAA GAA AAT GGG TAT ACG ATT CAA AAT AGA ATC GCT	96
Pro Asp Leu Trp Glu Glu Asn Gly Tyr Thr Ile Gln Asn Arg Ile Ala	
905 910 915	

GTA AAG TGG GAC GAT TCT CTA GCA AGT AAA GGG TAT ACG AAA TTT GTT	144
Val Lys Trp Asp Asp Ser Leu Ala Ser Lys Gly Tyr Thr Lys Phe Val	
920 925 930	

TCA AAT CCA CTA GAA AGT CAC ACA GTT GGT GAT CCT TAT ACA GAT TAT	192
Ser Asn Pro Leu Glu Ser His Thr Val Gly Asp Pro Tyr Thr Asp Tyr	
935 940 945	

GAA AAG GCA GCA AGA GAT CTA GAT TTG TCA AAT GCA AAG GAA ACG TTT	240
Glu Lys Ala Ala Arg Asp Leu Asp Leu Ser Asn Ala Lys Glu Thr Phe	
950 955 960	

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AAC CCA TTG GTA GCT GCT TTT CCA AGT GTG AAT GTT AGT ATG GAA AAG Asn Pro Leu Val Ala Ala Phe Pro Ser Val Asn Val Ser Met Glu Lys 965 970 975 980	288
GTG ATA TTA TCA CCA AAT GAA AAT TTA TCC AAT AGT GTA GAG TCT CAT Val Ile Leu Ser Pro Asn Glu Asn Leu Ser Asn Ser Val Glu Ser His 985 990 995	336
TCA TCC ACG AAT TGG TCT TAT ACA AAT ACA GAA GGT GCT TCT GTT GAA Ser Ser Thr Asn Trp Ser Tyr Thr Asn Thr Glu Gly Ala Ser Val Glu 1000 1005 1010	384
GCG GGG ATT GGA CCA AAA GGT ATT TCG TTC GGA GTT AGC GTA AAC TAT Ala Gly Ile Gly Pro Lys Gly Ile Ser Phe Gly Val Ser Val Asn Tyr 1015 1020 1025	432
CAA CAC TCT GAA ACA GTT GCA CAA GAA TGG GGA ACA TCT ACA GGA AAT Gln His Ser Glu Thr Val Ala Gln Glu Trp Gly Thr Ser Thr Gly Asn 1030 1035 1040	480
ACT TCG CAA TTC AAT ACG GCT TCA GCG GGA TAT TTA AAT GCA AAT GTT Thr Ser Gln Phe Asn Thr Ala Ser Ala Gly Tyr Leu Asn Ala Asn Val 1045 1050 1055 1060	528
CGA TAT AAC AAT GTA GGA ACT GGT GCC ATC TAC GAT GTA AAA CCT ACA Arg Tyr Asn Asn Val Gly Thr Gly Ala Ile Tyr Asp Val Lys Pro Thr 1065 1070 1075	576
ACA AGT TTT GTA TTA AAT AAC GAT ACT ATC GCA ACT ATT ACG GCG AAA Thr Ser Phe Val Leu Asn Asn Asp Thr Ile Ala Thr Ile Thr Ala Lys 1080 1085 1090	624
TCT AAT TCT ACA GCC TTA AAT ATA TCT CCT GGA GAA AGT TAC CCG AAA Ser Asn Ser Thr Ala Leu Asn Ile Ser Pro Gly Glu Ser Tyr Pro Lys 1095 1100 1105	672
AAA GGA CAA AAT GGA ATC GCA ATA ACA TCA ATG GAT GAT TTT AAT TCC Lys Gly Gln Asn Gly Ile Ala Ile Thr Ser Met Asp Asp Phe Asn Ser 1110 1115 1120	720
CAT CCG ATT ACA TTA AAT AAA AAA CAA GTA GAT AAT CTG CTA AAT AAT His Pro Ile Thr Leu Asn Lys Lys Gln Val Asp Asn Leu Leu Asn Asn 1125 1130 1135 1140	768
AAA CCT ATG ATG TTG GAA ACA AAC CAA ACA GAT GGT GTT TAT AAG ATA Lys Pro Met Met Leu Glu Thr Asn Gln Thr Asp Gly Val Tyr Lys Ile 1145 1150 1155	816
AAA GAT ACA CAT GGA AAT ATA GTA ACT GGC GGA GAA TGG AAT GGT GTC Lys Asp Thr His Gly Asn Ile Val Thr Gly Gly Glu Trp Asn Gly Val 1160 1165 1170	864
ATA CAA CAA ATC AAG GCT AAA ACA GCG TCT ATT ATT GTG GAT GAT GGG Ile Gln Gln Ile Lys Ala Lys Thr Ala Ser Ile Ile Val Asp Asp Gly	912

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1175	1180	1185	
GAA CGT GTA GCA GAA AAA CGT GTA GCG GCA AAA GAT TAT GAA AAT CCA Glu Arg Val Ala Glu Lys Arg Val Ala Ala Lys Asp Tyr Glu Asn Pro 1190	1195	1200	960
GAA GAT AAA ACA CCG TCT TTA ACT TTA AAA GAT GCC CTG AAG CTT TCA Glu Asp Lys Thr Pro Ser Leu Thr Leu Lys Asp Ala Leu Lys Leu Ser 1205	1210	1215	1008
TAT CCA GAT GAA ATA AAA GAA ATA GAG GGA TTA TTA TAT TAT AAA AAC Tyr Pro Asp Glu Ile Lys Glu Ile Glu Gly Leu Leu Tyr Tyr Lys Asn 1225	1230	1235	1056
AAA CCG ATA TAC GAA TCG AGC GTT ATG ACT TAC TTA GAT GAA AAT ACA Lys Pro Ile Tyr Glu Ser Ser Val Met Thr Tyr Leu Asp Glu Asn Thr 1240	1245	1250	1104
GCA AAA GAA GTG ACC AAA CAA TTA AAT GAT ACC ACT GGG AAA TTT AAA Ala Lys Glu Val Thr Lys Gln Leu Asn Asp Thr Thr Gly Lys Phe Lys 1255	1260	1265	1152
GAT GTA AGT CAT TTA TAT GAT GTA AAA CTG ACT CCA AAA ATG AAT GTT Asp Val Ser His Leu Tyr Asp Val Lys Leu Thr Pro Lys Met Asn Val 1270	1275	1280	1200
ACA ATC AAA TTG TCT ATA CTT TAT GAT AAT GCT GAG TCT AAT GAT AAC Thr Ile Lys Leu Ser Ile Leu Tyr Asp Asn Ala Glu Ser Asn Asp Asn 1285	1290	1295	1248
TCA ATT GGT AAA TGG ACA AAC ACA AAT ATT GTT TCA GGT GGA AAT AAC Ser Ile Gly Lys Trp Thr Asn Thr Asn Ile Val Ser Gly Gly Asn Asn 1305	1310	1315	1296
GGA AAA AAA CAA TAT TCT TCT AAT AAT CCG GAT GCT AAT TTG ACA TTA Gly Lys Lys Gln Tyr Ser Ser Asn Asn Pro Asp Ala Asn Leu Thr Leu 1320	1325	1330	1344
AAT ACA GAT GCT CAA GAA AAA TTA AAT AAA AAT CGT GAC TAT TAT ATA Asn Thr Asp Ala Gln Glu Lys Leu Asn Lys Asn Arg Asp Tyr Tyr Ile 1335	1340	1345	1392
AGT TTA TAT ATG AAG TCA GAA AAA AAC ACA CAA TGT GAG ATT ACT ATA Ser Leu Tyr Met Lys Ser Glu Lys Asn Thr Gln Cys Glu Ile Thr Ile 1350	1355	1360	1440
GAT GGG GAG ATT TAT CCG ATC ACT ACA AAA ACA GTG AAT GTG AAT AAA Asp Gly Glu Ile Tyr Pro Ile Thr Thr Lys Thr Val Asn Val Asn Lys 1365	1370	1375	1488
GAC AAT TAC AAA AGA TTA GAT ATT ATA GCT CAT AAT ATA AAA AGT AAT Asp Asn Tyr Lys Arg Leu Asp Ile Ile Ala His Asn Ile Lys Ser Asn 1385	1390	1395	1536
CCA ATT TCT TCA CTT CAT ATT AAA ACG AAT GAT GAA ATA ACT TTA TTT			1584

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Pro Ile Ser Ser Leu His Ile Lys Thr Asn Asp Glu Ile Thr Leu Phe 1400 1405 1410	
TGG GAT GAT ATT TCT ATA ACA GAT GTA GCA TCA ATA AAA CCG GAA AAT Trp Asp Asp Ile Ser Ile Thr Asp Val Ala Ser Ile Lys Pro Glu Asn 1415 1420 1425	1632
TTA ACA GAT TCA GAA ATT AAA CAG ATT TAT AGT AGG TAT GGT ATT AAG Leu Thr Asp Ser Glu Ile Lys Gln Ile Tyr Ser Arg Tyr Gly Ile Lys 1430 1435 1440	1680
TTA GAA GAT GGA ATC CTT ATT GAT AAA AAA GGT GGG ATT CAT TAT GGT Leu Glu Asp Gly Ile Leu Ile Asp Lys Lys Gly Gly Ile His Tyr Gly 1445 1450 1455 1460	1728
GAA TTT ATT AAT GAA GCT AGT TTT AAT ATT GAA CCA TTG CCA AAT TAT Glu Phe Ile Asn Glu Ala Ser Phe Asn Ile Glu Pro Leu Pro Asn Tyr 1465 1470 1475	1776
GTG ACC AAA TAT GAA GTT ACT TAT AGT AGT GAG TTA GGA CCA AAC GTG Val Thr Lys Tyr Glu Val Thr Tyr Ser Ser Glu Leu Gly Pro Asn Val 1480 1485 1490	1824
AGT GAC ACA CTT GAA AGT GAT AAA ATT TAC AAG GAT GGG ACA ATT AAA Ser Asp Thr Leu Glu Ser Asp Lys Ile Tyr Lys Asp Gly Thr Ile Lys 1495 1500 1505	1872
TTT GAT TTT ACC AAA TAT AGT AAA AAT GAA CAA GGA TTA TTT TAT GAC Phe Asp Phe Thr Lys Tyr Ser Lys Asn Glu Gln Gly Leu Phe Tyr Asp 1510 1515 1520	1920
AGT GGA TTA AAT TGG GAC TTT AAA ATT AAT GCT ATT ACT TAT GAT GGT Ser Gly Leu Asn Trp Asp Phe Lys Ile Asn Ala Ile Thr Tyr Asp Gly 1525 1530 1535 1540	1968
AAA GAG ATG AAT GTT TTT CAT AGA TAT AAT AAA TAG Lys Glu Met Asn Val Phe His Arg Tyr Asn Lys 1545 1550	2004

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 667 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Lys Arg Glu Ile Asp Glu Asp Thr Asp Thr Asp Gly Asp Ser Ile 1 5 10 15
--

Pro Asp Leu Trp Glu Glu Asn Gly Tyr Thr Ile Gln Asn Arg Ile Ala

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20	25	30
Val Lys Trp Asp Asp Ser Leu Ala Ser Lys Gly Tyr Thr Lys Phe Val		
35	40	45
Ser Asn Pro Leu Glu Ser His Thr Val Gly Asp Pro Tyr Thr Asp Tyr		
50	55	60
Glu Lys Ala Ala Arg Asp Leu Asp Leu Ser Asn Ala Lys Glu Thr Phe		
65	70	75
Asn Pro Leu Val Ala Ala Phe Pro Ser Val Asn Val Ser Met Glu Lys		
85	90	95
Val Ile Leu Ser Pro Asn Glu Asn Leu Ser Asn Ser Val Glu Ser His		
100	105	110
Ser Ser Thr Asn Trp Ser Tyr Thr Asn Thr Glu Gly Ala Ser Val Glu		
115	120	125
Ala Gly Ile Gly Pro Lys Gly Ile Ser Phe Gly Val Ser Val Asn Tyr		
130	135	140
Gln His Ser Glu Thr Val Ala Gln Glu Trp Gly Thr Ser Thr Gly Asn		
145	150	155
Thr Ser Gln Phe Asn Thr Ala Ser Ala Gly Tyr Leu Asn Ala Asn Val		
165	170	175
Arg Tyr Asn Asn Val Gly Thr Gly Ala Ile Tyr Asp Val Lys Pro Thr		
180	185	190
Thr Ser Phe Val Leu Asn Asn Asp Thr Ile Ala Thr Ile Thr Ala Lys		
195	200	205
Ser Asn Ser Thr Ala Leu Asn Ile Ser Pro Gly Glu Ser Tyr Pro Lys		
210	215	220
Lys Gly Gln Asn Gly Ile Ala Ile Thr Ser Met Asp Asp Phe Asn Ser		
225	230	235
His Pro Ile Thr Leu Asn Lys Lys Gln Val Asp Asn Leu Leu Asn Asn		
245	250	255
Lys Pro Met Met Leu Glu Thr Asn Gln Thr Asp Gly Val Tyr Lys Ile		
260	265	270
Lys Asp Thr His Gly Asn Ile Val Thr Gly Gly Glu Trp Asn Gly Val		
275	280	285
Ile Gln Gln Ile Lys Ala Lys Thr Ala Ser Ile Ile Val Asp Asp Gly		
290	295	300
Glu Arg Val Ala Glu Lys Arg Val Ala Ala Lys Asp Tyr Glu Asn Pro		
305	310	315
		320

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Glu Asp Lys Thr Pro Ser Leu Thr Leu Lys Asp Ala Leu Lys Leu Ser
325 330 335

Tyr Pro Asp Glu Ile Lys Glu Ile Gly Leu Leu Tyr Tyr Lys Asn
340 345 350

Lys Pro Ile Tyr Glu Ser Ser Val Met Thr Tyr Leu Asp Glu Asn Thr
355 360 365

Ala Lys Glu Val Thr Lys Gln Leu Asn Asp Thr Thr Gly Lys Phe Lys
370 375 380

Asp Val Ser His Leu Tyr Asp Val Lys Leu Thr Pro Lys Met Asn Val
385 390 395 400

Thr Ile Lys Leu Ser Ile Leu Tyr Asp Asn Ala Glu Ser Asn Asp Asn
405 410 415

Ser Ile Gly Lys Trp Thr Asn Thr Asn Ile Val Ser Gly Gly Asn Asn
420 425 430

Gly Lys Lys Gln Tyr Ser Ser Asn Asn Pro Asp Ala Asn Leu Thr Leu
435 440 445

Asn Thr Asp Ala Gln Glu Lys Leu Asn Lys Asn Arg Asp Tyr Tyr Ile
450 455 460

Ser Leu Tyr Met Lys Ser Glu Lys Asn Thr Gln Cys Glu Ile Thr Ile
465 470 475 480

Asp Gly Glu Ile Tyr Pro Ile Thr Thr Lys Thr Val Asn Val Asn Lys
485 490 495

Asp Asn Tyr Lys Arg Leu Asp Ile Ile Ala His Asn Ile Lys Ser Asn
500 505 510

Pro Ile Ser Ser Leu His Ile Lys Thr Asn Asp Glu Ile Thr Leu Phe
515 520 525

Trp Asp Asp Ile Ser Ile Thr Asp Val Ala Ser Ile Lys Pro Glu Asn
530 535 540

Leu Thr Asp Ser Glu Ile Lys Gln Ile Tyr Ser Arg Tyr Gly Ile Lys
545 550 555 560

Leu Glu Asp Gly Ile Leu Ile Asp Lys Lys Gly Gly Ile His Tyr Gly
565 570 575

Glu Phe Ile Asn Glu Ala Ser Phe Asn Ile Glu Pro Leu Pro Asn Tyr
580 585 590

Val Thr Lys Tyr Glu Val Thr Tyr Ser Ser Glu Leu Gly Pro Asn Val
595 600 605

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Ser Asp Thr Leu Glu Ser Asp Lys Ile Tyr Lys Asp Gly Thr Ile Lys
610 615 620

Phe Asp Phe Thr Lys Tyr Ser Lys Asn Glu Gln Gly Leu Phe Tyr Asp
625 630 635 640

Ser Gly Leu Asn Trp Asp Phe Lys Ile Asn Ala Ile Thr Tyr Asp Gly
645 650 655

Lys Glu Met Asn Val Phe His Arg Tyr Asn Lys
660 665

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Bacillus cereus*
- (B) STRAIN: AB78
- (C) INDIVIDUAL ISOLATE: NRRL B-21058

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..16
- (D) OTHER INFORMATION: /note= "N-terminal sequence of protein purified from strain AB78"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Lys Arg Glu Ile Asp Glu Asp Thr Asp Thr Asx Gly Asp Ser Ile Pro
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

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(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..21
- (D) OTHER INFORMATION: /note= "Oligonucleotide probe based on amino acids 3 to 9 of SEQ ID NO:8, using codon usage of *Bacillus thuringiensis*"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GAAATTGATC AAGATA^{CNGA} T

21

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Bacillus thuringiensis*
- (B) STRAIN: AB88

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..14
- (D) OTHER INFORMATION: /note= "N-terminal amino acid sequence of protein known as anion exchange fraction 23 (smaller)"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Xaa Glu Pro Phe Val Ser Ala Xaa Xaa Xaa Gln Xaa Xaa Xaa
1 5 10

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: N-terminal

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(vi) ORIGINAL SOURCE:
(A) ORGANISM: *Bacillus thuringiensis*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Xaa Glu Tyr Glu Asn Val Glu Pro Phe Val Ser Ala Xaa
1 5 10

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 14 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: N-terminal

(vi) ORIGINAL SOURCE:
(A) ORGANISM: *Bacillus thuringiensis*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Asn Lys Asn Asn Thr Lys Leu Pro Thr Arg Ala Leu Pro
1 5 10

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:
(A) ORGANISM: *Bacillus thuringiensis*
(B) STRAIN: AB88

(ix) FEATURE:

(A) NAME/KEY: Peptide
(B) LOCATION: 1..15
(D) OTHER INFORMATION: /note= "N-terminal amino acid sequence of 35 kDa VIP active against *Agrotis ipsilon*"

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ala Leu Ser Glu Asn Thr Gly Lys Asp Gly Gly Tyr Ile Val Pro
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: N-terminal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Bacillus thuringiensis*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Asp Asn Asn Pro Asn Ile Asn Glu
1 5

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..9
- (D) OTHER INFORMATION: /note= "N-terminal sequence of 80 kDa delta-endotoxin"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Asp Asn Asn Pro Asn Ile Asn Glu
1 5

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 11 amino acids

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- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: N-terminal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Bacillus thuringiensis*

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..11

(D) OTHER INFORMATION: /note= "N-terminal sequence from 60 kDa delta-endotoxin"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Asn Val Leu Asn Ser Gly Arg Thr Thr Ile
1 5 10

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2655 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..2652

(D) OTHER INFORMATION: /note= "Maize optimized DNA sequence for 100 kd VIP1A(a) protein from AB78"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATGAAGAAC	TGAAGAAGAA	GCTGGCCAGC	GTGGTGACCT	GCACCCTGCT	GGCCCCCATG	60
TTCCTGAAC	GCAACGTGAA	CGCCGTGTAC	GCCGACAGCA	AGACCAACCA	GATCAGCACC	120
ACCCAGAAGA	ACCAGCAGAA	GGAGATGGAC	CGCAAGGGCC	TGCTGGGCTA	CTACTTCAAG	180

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GGCAAGGACT TCAGCAACCT GACCATGTT GC_{CCCCCACGC} GTGACAGCAC CCTGATCTAC 240
GACCAGCAGA CCGCCAACAA GCTGCTGGAC AAGAACGAG AGGAGTACCA GAGCATCCGC 300
TGGATCGGCC TGATCCAGAG CAAGGAGACC GGCGACTTCA CCTTCAACCT GAGCGAGGAC 360
GAGCAGGCCA TCATCGAGAT CAACGGCAAG ATCATCAGCA ACAAGGGCAA GGAGAACGAG 420
GTGGTGCACC TGGAGAAGGG CAAGCTGGTG CCCATCAAGA TCGAGTACCA GAGCGACACC 480
AAGTTCAACA TCGACAGCAA GACCTCAAG GAGCTGAAGC TTTTCAAGAT CGACAGCCAG 540
AACCAGCCCC AGCAGGTGCA GCAGGACGAG CTGCGCAACC CCGAGTTCAA CAAGAAGGAG 600
AGCCAGGAGT TCCTGGCAA GCCCAGCAAG ATCAACCTGT TCACCCAGCA GATGAAGCGC 660
GAGATCGACG AGGACACCGA CACCGACGGC GACAGCATCC CCGACCTGTG GGAGGAGAAC 720
GGCTACACCA TCCAGAACCG CATCGCCGTG AAGTGGGACG ACAGCCTGGC TAGCAAGGGC 780
TACACCAAGT TCGTGAGCAA CCCCCTGGAG AGCCACACCG TGGGGACCC CTACACCGAC 840
TACGAGAAGG CCGCCCGCGA CCTGGACCTG AGCAACGCCA AGGAGACCTT CAACCCCCTG 900
GTGGCCGCCT TCCCCAGCGT GAACGTGAGC ATGGAGAAGG TGATCCTGAG CCCAACGAG 960
AACCTGAGCA ACAGCGTGGA GAGCCACTCG AGCACCAACT GGAGCTACAC CAACACCGAG 1020
GGCGCCAGCG TGGAGGCCGG CATCGGTCCC AAGGGCATCA GCTTCGGCGT GAGCGTGAAC 1080
TACCAGCACA GCGAGACCGT GGCCCAGGAG TGGGGCACCA GCACCGGCAA CACCAGCCAG 1140
TTCAACACCG CCAGGCCCGG CTACCTGAAC GCCAACGTGC GCTACAACAA CGTGGGCACC 1200
GGCGCCATCT ACGACGTGAA GCCCACCACC AGCTTCGTGC TGAACAAACGA CACCATCGCC 1260
ACCATCACCG CCAAGTCGAA TTCCACCGCC CTGAACATCA GCCCCGGCGA GAGCTACCC 1320
AAGAAGGCC AGAACGGCAT CGCCATCACC AGCATGGACG ACTTCAACAG CCACCCCCATC 1380
ACCCCTGAACA AGAACGAGGT GGACAACCTG CTGAACAAACA AGCCCATGAT GCTGGAGACC 1440
AACCAGACCG ACGCGTCTA CAAGATCAAG GACACCCACG GCAACATCGT GACCGGGCGC 1500
GAGTGGAACG GCGTGATCCA GCAGATCAAG GCCAAGACCG CCAGCATCAT CGTCGACGAC 1560
GGCGAGCGCG TGGCCGAGAA GCGCGTGGCC GCCAAGGACT ACGAGAACCC CGAGGACAAG 1620
ACCCCCAGCC TGACCCCTGAA GGACGCCCTG AAGCTGAGCT ACCCCGACGA GATCAAGGAG 1680
ATCGAGGGCC TGCTGTACTA CAAGAACAAAG CCCATCTACG AGAGCAGCGT GATGACCTAT 1740
CTAGACGAGA ACACCGCCAA GGAGGTGACC AAGCAGCTGA ACGACACCCAC CGGCAAGTTC 1800
AAGGACGTGA GCCACCTGTA CGACGTGAAG CTGACCCCCA AGATGAACGT GACCACATCAAG 1860

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CTGAGCATCC TGTACGACAA CGCCGAGAGC AACGACAACA GCATGGCAA GTGGACCAAC	1920
ACCAACATCG TGAGCGGCGG CAACAACGGC AAGAAGCAGT ACAGCAGCAA CAACCCCGAC	1980
GCCAACCTGA CCCTGAACAC CGACGCCAG GAGAAGCTGA ACAAGAACCG CGACTACTAC	2040
ATCAGCCTGT ACATGAAGAG CGAGAAGAAC ACCCAGTGCG AGATCACCAT CGACGGCGAG	2100
ATATAACCCA TCACCACCAA GACCGTGAAC GTGAACAAGG ACAACTACAA GCCCTGGAC	2160
ATCATCGCCC ACAACATCAA GAGCAACCCC ATCAGCAGCC TGCACATCAA GACCAACGAC	2220
GAGATCACCC TGTTCTGGGA CGACATATCG ATTACCGACG TCGCCAGCAT CAAGCCCGAG	2280
AACCTGACCG ACAGCGAGAT CAAGCAGATA TACAGTCGCT ACGGCATCAA GCTGGAGGAC	2340
GGCATCCTGA TCGACAAGAA GGGCGGCATC CACTACGGCG AGTTCATCAA CGAGGCCAGC	2400
TTCAACATCG AGCCCTGCA GAACTACGTG ACCAAGTACG AGGTGACCTA CAGCAGCGAG	2460
CTGGGCCCA ACGTGAGCGA CACCCCTGGAG AGCGACAAGA TTTACAAGGA CGGCACCATC	2520
AAGTTCGACT TCACCAAGTA CAGCAAGAAC GAGCAGGGCC TGTTCTACGA CAGCGGCCTG	2580
AACTGGGACT TCAAGATCAA CGCCATCACC TACGACGGCA AGGAGATGAA CGTGTCCAC	2640
CGCTACAACA AGTAG	2655

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2004 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..2004
- (D) OTHER INFORMATION: /note= "Maize optimized DNA sequence for VIP1A(a) 80 kd protein from AB78"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ATGAAGCGCG AGATCGACGA GGACACCGAC ACCGACGGCG ACAGCATCCC CGACCTGTGG

60

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GAGGAGAACG GCTACACCAT CCAGAACCGC ATGCCGTGA AGTGGGACGA CAGCCTGGCT	120
AGCAAGGGCT ACACCAAGTT CGTGAGCAAC CCCCTGGAGA GCCACACCGT GGGCGACCCC	180
TACACCGACT ACGAGAAGGC CGCCCGCGAC CTGGACCTGA GCAACGCCAA GGAGACCTTC	240
AACCCCCCTGG TGGCCGCCTT CCCCAGCGTG AACGTGAGCA TGGAGAAGGT GATCCTGAGC	300
CCCAACGAGA ACCTGAGCAA CAGCGTGGAG AGCCACTCGA GCACCAACTG GAGCTACACC	360
AACACCGAGG GCGCCAGCGT GGAGGCCGGC ATCGGTCCCA AGGGCATCAG CTTCGCGTG	420
AGCGTGAACT ACCAGCACAG CGAGACCGTG GCCCAGGAGT GGGGCACCAG CACCGGCAAC	480
ACCAGCCAGT TCAACACCGC CAGCGCCGGC TACCTGAACG CCAACGTGCG CTACAACAAAC	540
GTGGGCACCG GCGCCATCTA CGACGTGAAG CCCACCACCA GCTTCGTGCT GAACAACGAC	600
ACCATCGCCA CCATCACCGC CAAGTCGAAT TCCACCGCCC TGAACATCAG CCCCGGCGAG	660
AGCTACCCCA AGAAGGGCCA GAACGGCATC GCCATCACCA GCATGGACGA CTTCAACAGC	720
CACCCCATCA CCCTGAACAA GAAGCAGGTG GACAACCTGC TGAACAACAA GCCCATGATG	780
CTGGAGACCA ACCAGACCGA CGCGTCTAC AAGATCAAGG ACACCCACGG CAACATCGTG	840
ACCGGCGGCG AGTGGAACCGG CGTGATCCAG CAGATCAAGG CCAAGACCGC CAGCATCATC	900
GTCGACGACG GCGAGCGCGT GGCGAGAAAG CGCGTGGCCG CCAAGGACTA CGAGAACCCC	960
GAGGACAAGA CCCCCAGCCT GACCCCTGAAG GACGCCCTGA AGCTGAGCTA CCCCCACGAG	1020
ATCAAGGAGA TCGAGGGCCT GCTGTACTAC AAGAACAAAGC CCATCTACGA GAGCAGCGTG	1080
ATGACCTATC TAGACGAGAA CACCGCCAAG GAGGTGACCA AGCAGCTGAA CGACACCACC	1140
GGCAAGTTCA AGGACGTGAG CCACCTGTAC GACGTGAAGC TGACCCCCAA GATGAACGTG	1200
ACCATCAAGC TGAGCATCCT GTACGACAAC GCCGAGAGCA ACGACAACAG CATCGGCAAG	1260
TGGACCAACA CCAACATCGT GAGCGCGGC AACAAACGGCA AGAACAGTA CAGCAGAAC	1320
AACCCCGACG CCAACCTGAC CCTGAACACC GACGCCAGG AGAAGCTGAA CAAGAACCGC	1380
GACTACTACA TCAGCCTGTA CATGAAGAGC GAGAAGAACCA CCCAGTGCAG GATCACCATC	1440
GACGGCGAGA TATAACCCAT CACCACCAAG ACCGTGAACG TGAACAAGGA CAACTACAAG	1500
CGCCTGGACA TCATCGCCA CAACATCAAG AGCAACCCC TAAGCAGCCT GCACATCAAG	1560
ACCAACGACG AGATCACCCCT GTTCTGGAC GACATATCGA TTACCGACGT CGCCAGCATC	1620
AAGCCCGAGA ACCTGACCGA CAGCGAGATC AAGCAGATAT ACAGTCGCTA CGGCATCAAG	1680
CTGGAGGACG GCATCCTGAT CGACAAGAACGGCGCATCC ACTACGGCGA GTTCATCAAC	1740

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GAGGCCAGCT TCAACATCGA GCCCCTGCAG AACTACGTGA CCAAGTACGA GGTGACCTAC	1800
AGCAGCGAGC TGGGCCCAA CGTGAGCGAC ACCCTGGAGA GCGACAAGAT TTACAAGGAC	1860
GGCACCATCA AGTCGACTT CACCAAGTAC AGCAAGAACG AGCAGGGCCT GTTCTACGAC	1920
AGCGGCCTGA ACTGGGACTT CAAGATCAAC GCCATCACCT ACGACGGCAA GGAGATGAAC	1980
GTGTTCCACC GCTACAAACAA GTAG	2004

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4074 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1386
- (D) OTHER INFORMATION: /product= "VIP2A(b) from Btt"

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1394..3895
- (D) OTHER INFORMATION: /product= "VIP1A(b) from Btt"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..4074
- (D) OTHER INFORMATION: /note= "Cloned DNA sequence from Btt which contains the genes for both VIP1A(b) and VIP2A(b)"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ATG CAA AGA ATG GAG GGA AAG TTG TTT GTG GTG TCA AAA ACA TTA CAA Met Gln Arg Met Glu Gly Lys Leu Phe Val Val Ser Lys Thr Leu Gln	48
670 675 680	
GTA GTT ACT AGA ACT GTA TTG CTT AGT ACA GTT TAC TCT ATA ACT TTA Val Val Thr Arg Thr Val Leu Leu Ser Thr Val Tyr Ser Ile Thr Leu	96
685 690 695	
TTA AAT AAT GTA GTG ATA AAA GCT GAC CAA TTA AAT ATA AAT TCT CAA Leu Asn Asn Val Val Ile Lys Ala Asp Gln Leu Asn Ile Asn Ser Gln	144
700 705 710 715	
AGT AAA TAT ACT AAC TTG CAA AAT CTA AAA ATC CCT GAT AAT GCA GAG Ser Lys Tyr Thr Asn Leu Gln Asn Leu Ile Pro Asp Asn Ala Glu	192

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720	725	730	
GAT TTT AAA GAA GAT AAG GGG AAA GCG AAA GAA TGG GGG AAA GAG AAA Asp Phe Lys Glu Asp Lys Gly Lys Ala Lys Glu Trp Gly Lys Glu Lys 735	740	745	240
GGG GAA GAG TGG AGG CCT CCT GCT ACT GAG AAA GGA GAA ATG AAT AAT Gly Glu Glu Trp Arg Pro Pro Ala Thr Glu Lys Gly Glu Met Asn Asn 750	755	760	288
TTT TTA GAT AAT AAA AAT GAT ATA AAG ACC AAT TAT AAA GAA ATT ACT Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr 765	770	775	336
TTT TCT ATG GCA GGT TCA TGT GAA GAT GAA ATA AAA GAT TTA GAA GAA Phe Ser Met Ala Gly Ser Cys Glu Asp Glu Ile Lys Asp Leu Glu Glu 780	785	790	384
ATT GAT AAG ATC TTT GAT AAA GCC AAT CTC TCG AGT TCT ATT ATC ACC Ile Asp Ile Phe Asp Lys Ala Asn Leu Ser Ser Ser Ile Ile Thr 800	805	810	432
TAT AAA AAT GTG GAA CCA GCA ACA ATT GGA TTT AAT AAA TCT TTA ACA Tyr Lys Asn Val Glu Pro Ala Thr Ile Gly Phe Asn Lys Ser Leu Thr 815	820	825	480
GAA GGT AAT ACG ATT AAT TCT GAT GCA ATG GCA CAG TTT AAA GAA CAA Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln 830	835	840	528
TTT TTA GGT AAG GAT ATG AAG TTT GAT AGT TAT CTA GAT ACT CAT TTA Phe Leu Gly Lys Asp Met Lys Phe Asp Ser Tyr Leu Asp Thr His Leu 845	850	855	576
ACT GCT CAA CAA GTT TCC AGT AAA AAA AGA GTT ATT TTG AAG GTT ACG Thr Ala Gln Gln Val Ser Ser Lys Lys Arg Val Ile Leu Lys Val Thr 860	865	870	624
GTT CCG AGT GGG AAA GGT TCT ACT ACT CCA ACA AAA GCA GGT GTC ATT Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile 880	885	890	672
TTA AAC AAT AAT GAA TAC AAA ATG CTC ATT GAT AAT GGG TAT GTG CTC Leu Asn Asn Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Val Leu 895	900	905	720
CAT GTA GAT AAG GTA TCA AAA GTA GTA AAA AAA GGG ATG GAG TGC TTA His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Met Glu Cys Leu 910	915	920	768
CAA GTT GAA GGG ACT TTA AAA AAG AGT CTC GAC TTT AAA AAT GAT ATA Gln Val Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile 925	930	935	816
AAT GCT GAA GCG CAT AGC TGG GGG ATG AAA ATT TAT GAA GAC TGG GCT			864

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Asn Ala Glu Ala His Ser Trp Gly Met Lys Ile Tyr Glu Asp Trp Ala			
940	945	950	955
AAA AAT TTA ACC GCT TCG CAA AGG GAA GCT TTA GAT GGG TAT GCT AGG			912
Lys Asn Leu Thr Ala Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg			
960	965	970	
CAA GAT TAT AAA GAA ATC AAT TAT TTG CGC AAT CAA GGC GGG AGT			960
Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser			
975	980	985	
GGA AAT GAA AAG CTG GAT GCC CAA TTA AAA AAT ATT TCT GAT GCT TTA			1008
Gly Asn Glu Lys Leu Asp Ala Gln Leu Lys Asn Ile Ser Asp Ala Leu			
990	995	1000	
GGG AAG AAA CCC ATA CCA GAA AAT ATT ACC GTG TAT AGA TGG TGT GGC			1056
Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly			
1005	1010	1015	
ATG CCG GAA TTT GGT TAT CAA ATT AGT GAT CCG TTA CCT TCT TTA AAA			1104
Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys			
1020	1025	1030	1035
GAT TTT GAA GAA CAA TTT TTA AAT ACA ATT AAA GAA GAC AAA GGG TAT			1152
Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr			
1040	1045	1050	
ATG AGT ACA AGC TTA TCG AGT GAA CGT CTT GCA GCT TTT GGA TCT AGA			1200
Met Ser Thr Ser Leu Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg			
1055	1060	1065	
AAA ATT ATA TTA CGC TTA CAA GTT CCG AAA GGA AGT ACG GGG GCG TAT			1248
Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr			
1070	1075	1080	
TTA AGT GCC ATT GGT GGA TTT GCA AGT GAA AAA GAG ATC CTA CTT GAT			1296
Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp			
1085	1090	1095	
AAA GAT AGT AAA TAT CAT ATT GAT AAA GCA ACA GAG GTA ATC ATT AAA			1344
Lys Asp Ser Lys Tyr His Ile Asp Lys Ala Thr Glu Val Ile Ile Lys			
1100	1105	1110	1115
GGT GTT AAG CGA TAT GTA GTG GAT GCA ACA TTA TTA ACA AAT			1386
Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn			
1120	1125		
TAAGGAG ATG AAA AAT ATG AAG AAA AAG TTA GCA AGT GTT GTA ACC TGT			1435
Met Lys Asn Met Lys Lys Leu Ala Ser Val Val Thr Cys			
1	5	10	
ATG TTA TTA GCT CCT ATG TTT TTG AAT GGA AAT GTG AAT GCT GTT AAC			1483
Met Leu Leu Ala Pro Met Phe Leu Asn Gly Asn Val Asn Ala Val Asn			
15	20	25	30

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GCG GAT AGT AAA ATA AAT CAG ATT TCT ACA ACG CAG GAA AAC CAA CAG Ala Asp Ser Lys Ile Asn Gln Ile Ser Thr Thr Gln Glu Asn Gln Gln 35 40 45	1531
AAA GAG ATG GAC CGA AAG GGA TTA TTG GGA TAT TAT TTC AAA GGA AAA Lys Glu Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys 50 55 60	1579
GAT TTT AAT AAT CTT ACT ATG TTT GCA CCG ACA CGT GAT AAT ACC CTT Asp Phe Asn Asn Leu Thr Met Phe Ala Pro Thr Arg Asp Asn Thr Leu 65 70 75	1627
ATG TAT GAC CAA CAA ACA GCG AAT GCA TTA TTA GAT AAA AAA CAA CAA Met Tyr Asp Gln Gln Thr Ala Asn Ala Leu Leu Asp Lys Lys Gln Gln 80 85 90	1675
GAA TAT CAG TCC ATT CGT TGG ATT GGT TTG ATT CAG CGT AAA GAA ACG Glu Tyr Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln Arg Lys Glu Thr 95 100 105 110	1723
GGC GAT TTC ACA TTT AAC TTA TCA AAG GAT GAA CAG GCA ATT ATA GAA Gly Asp Phe Thr Phe Asn Leu Ser Lys Asp Glu Gln Ala Ile Ile Glu 115 120 125	1771
ATC GAT GGG AAA ATC ATT TCT AAT AAA GGG AAA GAA AAG CAA GTT GTC Ile Asp Gly Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val 130 135 140	1819
CAT TTA GAA AAA GAA AAA TTA GTT CCA ATC AAA ATA GAG TAT CAA TCA His Leu Glu Lys Glu Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser 145 150 155	1867
GAT ACG AAA TTT AAT ATT GAT AGT AAA ACA TTT AAA GAA CTT AAA TTA Asp Thr Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu 160 165 170	1915
TTT AAA ATA GAT AGT CAA AAC CAA TCT CAA CAA GTT CAA CTG AGA AAC Phe Lys Ile Asp Ser Gln Asn Gln Ser Gln Gln Val Gln Leu Arg Asn 175 180 185 190	1963
CCT GAA TTT AAC AAA AAA GAA TCA CAG GAA TTT TTA GCA AAA GCA TCA Pro Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Ala Ser 195 200 205	2011
AAA ACA AAC CTT TTT AAG CAA AAA ATG AAA AGA GAT ATT GAT GAA GAT Lys Thr Asn Leu Phe Lys Gln Lys Met Lys Arg Asp Ile Asp Glu Asp 210 215 220	2059
ACG GAT ACA GAT GGA GAC TCC ATT CCT GAT CTT TGG GAA GAA AAT GGG Thr Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn Gly 225 230 235	2107
TAC ACG ATT CAA AAT AAA GTT GCT GTC AAA TGG GAT GAT TCG CTA GCA Tyr Thr Ile Gln Asn Lys Val Ala Val Lys Trp Asp Asp Ser Leu Ala 240 245 250	2155

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AGT AAG GGA TAT ACA AAA TTT GTT TCG AAT CCA TTA GAC AGC CAC ACA Ser Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Asp Ser His Thr 255 260 265 270	2203
GTT GGC GAT CCC TAT ACT GAT TAT GAA AAG GCC GCA AGG GAT TTA GAT Val Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu Asp 275 280 285	2251
TTA TCA AAT GCA AAG GAA ACG TTC AAC CCA TTG GTA GCT GCT TTT CCA Leu Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe Pro 290 295 300	2299
AGT GTG AAT GTT AGT ATG GAA AAG GTG ATA TTA TCA CCA AAT GAA AAT Ser Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu Asn 305 310 315	2347
TTA TCC AAT AGT GTA GAG TCT CAT TCA TCC ACG AAT TGG TCT TAT ACG Leu Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr Thr 320 325 330	2395
AAT ACA GAA GGA GCT TCC ATT GAA GCT GGT GGC GGT CCA TTA GGC CTT Asn Thr Glu Gly Ala Ser Ile Glu Ala Gly Gly Pro Leu Gly Leu 335 340 345 350	2443
TCT TTT GGC GTG AGT GTT ACT TAT CAA CAC TCT GAA ACA GTT GCA CAA Ser Phe Gly Val Ser Val Thr Tyr Gln His Ser Glu Thr Val Ala Gln 355 360 365	2491
GAA TGG GGA ACA TCT ACA GGA AAT ACT TCA CAA TTC AAT ACG GCT TCA Glu Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala Ser 370 375 380	2539
GCG GGA TAT TTA AAT GCA AAT GTT CGG TAT AAC AAT GTA GGG ACT GGT Ala Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr Gly 385 390 395	2587
GCC ATC TAT GAT GTA AAA CCT ACA ACA AGT TTT GTA TTA AAT AAC AAT Ala Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn Asn 400 405 410	2635
ACC ATC GCA ACG ATT ACA GCA AAA TCA AAT TCA ACA GCT TTA CGT ATA Thr Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Arg Ile 415 420 425 430	2683
TCT CCG GGG GAT AGT TAT CCA GAA ATA GGA GAA AAC GCT ATT GCG ATT Ser Pro Gly Asp Ser Tyr Pro Glu Ile Gly Glu Asn Ala Ile Ala Ile 435 440 445	2731
ACA TCT ATG GAT GAT TTT AAT TCT CAT CCA ATT ACA TTA AAT AAA CAA Thr Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys Gln 450 455 460	2779
CAG GTA AAT CAA TTG ATA AAT AAT AAG CCA ATT ATG CTA GAG ACA GAC Gln Val Asn Gln Leu Ile Asn Asn Lys Pro Ile Met Leu Glu Thr Asp	2827

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465	470	475	
CAA ACA GAT GGT GTT TAT AAA ATA AGA GAT ACA CAT GGA AAT ATT GTA Gln Thr Asp Gly Val Tyr Lys Ile Arg Asp Thr His Gly Asn Ile Val 480	485	490	2875
ACT GGT GGA GAA TGG AAT GGT GTA ACA CAA CAA ATT AAA GCA AAA ACA Thr Gly Gly Glu Trp Asn Gly Val Thr Gln Gln Ile Lys Ala Lys Thr 495	500	505	2923
GCG TCT ATT ATT GTG GAT GAC GGG AAA CAG GTA GCA GAA AAA CGT GTG Ala Ser Ile Ile Val Asp Asp Gly Lys Gln Val Ala Glu Lys Arg Val 515	520	525	2971
GCG GCA AAA GAT TAT GGT CAT CCA GAA GAT AAA ACA CCA CCT TTA ACT Ala Ala Lys Asp Tyr Gly His Pro Glu Asp Lys Thr Pro Pro Leu Thr 530	535	540	3019
TTA AAA GAT ACC CTG AAG CTT TCA TAC CCA GAT GAA ATA AAA GAA ACT Leu Lys Asp Thr Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu Thr 545	550	555	3067
AAT GGA TTG TTG TAC TAT GAT GAC AAA CCA ATC TAT GAA TCG AGT GTC Asn Gly Leu Leu Tyr Tyr Asp Asp Lys Pro Ile Tyr Glu Ser Ser Val 560	565	570	3115
ATG ACT TAT CTG GAT GAA AAT ACG GCA AAA GAA GTC AAA AAA CAA ATA Met Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Lys Lys Gln Ile 575	580	585	3163
AAT GAT ACA ACC GGA AAA TTT AAG GAT GTA AAT CAC TTA TAT GAT GTA Asn Asp Thr Thr Gly Lys Phe Lys Asp Val Asn His Leu Tyr Asp Val 595	600	605	3211
AAA CTG ACT CCA AAA ATG AAT TTT ACG ATT AAA ATG GCT TCC TTG TAT Lys Leu Thr Pro Lys Met Asn Phe Thr Ile Lys Met Ala Ser Leu Tyr 610	615	620	3259
GAT GGG GCT GAA AAT AAT CAT AAC TCT TTA GGA ACC TGG TAT TTA ACA Asp Gly Ala Glu Asn Asn His Asn Ser Leu Gly Thr Trp Tyr Leu Thr 625	630	635	3307
TAT AAT GTT GCT GGT GGA AAT ACT GGG AAG AGA CAA TAT CGT TCA GCT Tyr Asn Val Ala Gly Gly Asn Thr Gly Lys Arg Gln Tyr Arg Ser Ala 640	645	650	3355
CAT TCT TGT GCA CAT GTA GCT CTA TCT TCA GAA GCG AAA AAG AAA CTA His Ser Cys Ala His Val Ala Leu Ser Ser Glu Ala Lys Lys Lys Leu 655	660	665	3403
AAT CAA AAT GCG AAT TAC TAT CTT AGC ATG TAT ATG AAG GCT GAT TCT Asn Gln Asn Ala Asn Tyr Tyr Leu Ser Met Tyr Met Lys Ala Asp Ser 675	680	685	3451
ACT ACG GAA CCT ACA ATA GAA GTA GCT GGG GAA AAA TCT GCA ATA ACA			3499

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(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 462 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Gln Arg Met Glu Gly Lys Leu Phe Val Val Ser Lys Thr Leu Gln
1 5 10 15

Val Val Thr Arg Thr Val Leu Leu Ser Thr Val Tyr Ser Ile Thr Leu
20 25 30

Leu Asn Asn Val Val Ile Lys Ala Asp Gln Leu Asn Ile Asn Ser Gln
35 40 45

Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Pro Asp Asn Ala Glu
50 55 60

Asp Phe Lys Glu Asp Lys Gly Lys Ala Lys Glu Trp Gly Lys Glu Lys
65 70 75 80

Gly Glu Glu Trp Arg Pro Pro Ala Thr Glu Lys Gly Glu Met Asn Asn
85 90 95

Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr
100 105 110

Phe Ser Met Ala Gly Ser Cys Glu Asp Glu Ile Lys Asp Leu Glu Glu
115 120 125

Ile Asp Lys Ile Phe Asp Lys Ala Asn Leu Ser Ser Ser Ile Ile Thr
130 135 140

Tyr Lys Asn Val Glu Pro Ala Thr Ile Gly Phe Asn Lys Ser Leu Thr
145 150 155 160

Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln
165 170 175

Phe Leu Gly Lys Asp Met Lys Phe Asp Ser Tyr Leu Asp Thr His Leu
180 185 190

Thr Ala Gln Gln Val Ser Ser Lys Lys Arg Val Ile Leu Lys Val Thr
195 200 205

Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile
210 215 220

Leu Asn Asn Asn Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Val Leu
225 230 235 240

His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Met Glu Cys Leu
245 250 255

Gln Val Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile
260 265 270

Asn Ala Glu Ala His Ser Trp Gly Met Lys Ile Tyr Glu Asp Trp Ala
275 280 285

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Lys Asn Leu Thr Ala Ser Gln Arg Glu Alà Leu Asp Gly Tyr Ala Arg
290 295 300

Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser
305 310 315 320

Gly Asn Glu Lys Leu Asp Ala Gln Leu Lys Asn Ile Ser Asp Ala Leu
325 330 335

Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly
340 345 350

Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys
355 360 365

Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr
370 375 380

Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg
385 390 395 400

Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr
405 410 415

Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp
420 425 430

Lys Asp Ser Lys Tyr His Ile Asp Lys Ala Thr Glu Val Ile Ile Lys
435 440 445

Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn
450 455 460

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 834 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Met Lys Asn Met Lys Lys Leu Ala Ser Val Val Thr Cys Met Leu
1 5 10 15

Leu Ala Pro Met Phe Leu Asn Gly Asn Val Asn Ala Val Asn Ala Asp
20 25 30

Ser Lys Ile Asn Gln Ile Ser Thr Thr Gln Glu Asn Gln Gln Lys Glu
35 40 45

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Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys Asp Phe
50 55 60

Asn Asn Leu Thr Met Phe Ala Pro Thr Arg Asp Asn Thr Leu Met Tyr
65 70 75 80

Asp Gln Gln Thr Ala Asn Ala Leu Leu Asp Lys Lys Gln Gln Glu Tyr
85 90 95

Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln Arg Lys Glu Thr Gly Asp
100 105 110

Phe Thr Phe Asn Leu Ser Lys Asp Glu Gln Ala Ile Ile Glu Ile Asp
115 120 125

Gly Lys Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val His Leu
130 135 140

Glu Lys Glu Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser Asp Thr
145 150 155 160

Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu Phe Lys
165 170 175

Ile Asp Ser Gln Asn Gln Ser Gln Gln Val Gln Leu Arg Asn Pro Glu
180 185 190

Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Ala Ser Lys Thr
195 200 205

Asn Leu Phe Lys Gln Lys Met Lys Arg Asp Ile Asp Glu Asp Thr Asp
210 215 220

Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn Gly Tyr Thr
225 230 235 240

Ile Gln Asn Lys Val Ala Val Lys Trp Asp Asp Ser Leu Ala Ser Lys
245 250 255

Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Asp Ser His Thr Val Gly
260 265 270

Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu Asp Leu Ser
275 280 285

Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe Pro Ser Val
290 295 300

Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu Asn Leu Ser
305 310 315 320

Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr Thr Asn Thr
325 330 335

Glu Gly Ala Ser Ile Glu Ala Gly Gly Pro Leu Gly Leu Ser Phe

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340	345	350
Gly Val Ser Val Thr Tyr Gln His Ser Glu Thr Val Ala Gln Glu Trp		
355	360	365
Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala Ser Ala Gly		
370	375	380
Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr Gly Ala Ile		
385	390	395
Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn Asn Thr Ile		
405	410	415
Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Arg Ile Ser Pro		
420	425	430
Gly Asp Ser Tyr Pro Glu Ile Gly Glu Asn Ala Ile Ala Ile Thr Ser		
435	440	445
Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys Gln Gln Val		
450	455	460
Asn Gln Leu Ile Asn Asn Lys Pro Ile Met Leu Glu Thr Asp Gln Thr		
465	470	475
Asp Gly Val Tyr Lys Ile Arg Asp Thr His Gly Asn Ile Val Thr Gly		
485	490	495
Gly Glu Trp Asn Gly Val Thr Gln Gln Ile Lys Ala Lys Thr Ala Ser		
500	505	510
Ile Ile Val Asp Asp Gly Lys Gln Val Ala Glu Lys Arg Val Ala Ala		
515	520	525
Lys Asp Tyr Gly His Pro Glu Asp Lys Thr Pro Pro Leu Thr Leu Lys		
530	535	540
Asp Thr Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu Thr Asn Gly		
545	550	560
Leu Leu Tyr Tyr Asp Asp Lys Pro Ile Tyr Glu Ser Ser Val Met Thr		
565	570	575
Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Lys Lys Gln Ile Asn Asp		
580	585	590
Thr Thr Gly Lys Phe Lys Asp Val Asn His Leu Tyr Asp Val Lys Leu		
595	600	605
Thr Pro Lys Met Asn Phe Thr Ile Lys Met Ala Ser Leu Tyr Asp Gly		
610	615	620
Ala Glu Asn Asn His Asn Ser Leu Gly Thr Trp Tyr Leu Thr Tyr Asn		
625	630	640

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Val Ala Gly Gly Asn Thr Gly Lys Arg Gln Tyr Arg Ser Ala His Ser
645 650 655

Cys Ala His Val Ala Leu Ser Ser Glu Ala Lys Lys Lys Leu Asn Gln
660 665 670

Asn Ala Asn Tyr Tyr Leu Ser Met Tyr Met Lys Ala Asp Ser Thr Thr
675 680 685

Glu Pro Thr Ile Glu Val Ala Gly Glu Lys Ser Ala Ile Thr Ser Lys
690 695 700

Lys Val Lys Leu Asn Asn Gln Asn Tyr Gln Arg Val Asp Ile Leu Val
705 710 715 720

Lys Asn Ser Glu Arg Asn Pro Met Asp Lys Ile Tyr Ile Arg Gly Asn
725 730 735

Gly Thr Thr Asn Val Tyr Gly Asp Asp Val Thr Ile Pro Glu Val Ser
740 745 750

Ala Ile Asn Pro Ala Ser Leu Ser Asp Glu Glu Ile Gln Glu Ile Phe
755 760 765

Lys Asp Ser Thr Ile Glu Tyr Gly Asn Pro Ser Phe Val Ala Asp Ala
770 775 780

Val Thr Phe Lys Asn Ile Lys Pro Leu Gln Asn Tyr Val Lys Glu Tyr
785 790 795 800

Glu Ile Tyr His Lys Ser His Arg Tyr Glu Lys Lys Thr Val Phe Asp
805 810 815

Ile Met Gly Val His Tyr Glu Tyr Ser Ile Ala Arg Glu Gln Lys Lys
820 825 830

Ala Ala

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4041 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..4038
- (D) OTHER INFORMATION: /product= "VIP1A(a)/VIP2A(a) fusion"

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product"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

ATG AAA AGA ATG GAG GGA AAG TTG TTT ATG GTG TCA AAA AAA TTA CAA Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys Leu Gln 835 840 845 850	48
GTA GTT ACT AAA ACT GTA TTG CTT AGT ACA GTT TTC TCT ATA TCT TTA Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile Ser Leu 855 860 865	6
TTA AAT AAT GAA GTG ATA AAA GCT GAA CAA TTA AAT ATA AAT TCT CAA Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn Ser Gln 870 875 880	144
AGT AAA TAT ACT AAC TTG CAA AAT CTA AAA ATC ACT GAC AAG GTA GAG Ser Lys Tyr Thr Asn Leu Gln Leu Asn Leu Lys Ile Thr Asp Lys Val Glu 885 890 895	192
GAT TTT AAA GAA GAT AAG GAA AAA GCG AAA GAA TGG GGG AAA GAA AAA Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys 900 905 910	240
GAA AAA GAG TGG AAA CTA ACT GCT ACT GAA AAA GGA AAA ATG AAT AAT Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn 915 920 925 930	288
TTT TTA GAT AAT AAA AAT GAT ATA AAG ACA AAT TAT AAA GAA ATT ACT Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr 935 940 945	336
TTT TCT ATG GCA GGC TCA TTT GAA GAT GAA ATA AAA GAT TTA AAA GAA Phe Ser Met Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu 950 955 960	384
ATT GAT AAG ATG TTT GAT AAA ACC AAT CTA TCA AAT TCT ATT ATC ACC Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr 965 970 975	432
TAT AAA AAT GTG GAA CCG ACA ACA ATT GGA TTT AAT AAA TCT TTA ACA Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr 980 985 990	480
GAA GGT AAT ACG ATT AAT TCT GAT GCA ATG GCA CAG TTT AAA GAA CAA Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln 995 1000 1005 1010	528
TTT TTA GAT AGG GAT ATT AAG TTT GAT AGT TAT CTA GAT ACG CAT TTA Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu 1015 1020 1025	576
ACT GCT CAA CAA GTT TCC AGT AAA GAA AGA GTT ATT TTG AAG GTT ACG Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr	624

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1030	1035	1040	
GTT CCG AGT GGG AAA GGT TCT ACT ACT CCA ACA AAA GCA GGT GTC ATT Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile 1045	1050	1055	672
TTA AAT AAT AGT GAA TAC AAA ATG CTC ATT GAT AAT GGG TAT ATG GTC Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val 1060	1065	1070	720
CAT GTA GAT AAG GTA TCA AAA GTG GTG AAA AAA GGG GTG GAG TGC TTA His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu 1075	1080	1085	768
CAA ATT GAA GGG ACT TTA AAA AAG AGT CTT GAC TTT AAA AAT GAT ATA Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile 1095	1100	1105	816
AAT GCT GAA GCG CAT AGC TGG GGT ATG AAG AAT TAT GAA GAG TGG GCT Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala 1110	1115	1120	864
AAA GAT TTA ACC GAT TCG CAA AGG GAA GCT TTA GAT GGG TAT GCT AGG Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg 1125	1130	1135	912
CAA GAT TAT AAA GAA ATC AAT AAT TAT TTA AGA AAT CAA GGC GGA AGT Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Ser 1140	1145	1150	960
GGA AAT GAA AAA CTA GAT GCT CAA ATA AAA AAT ATT TCT GAT GCT TTA Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu 1155	1160	1165	1008
GGG AAG AAA CCA ATA CCG GAA AAT ATT ACT GTG TAT AGA TGG TGT GGC Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly 1175	1180	1185	1056
ATG CCG GAA TTT GGT TAT CAA ATT AGT GAT CCG TTA CCT TCT TTA AAA Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys 1190	1195	1200	1104
GAT TTT GAA GAA CAA TTT TTA AAT ACA ATC AAA GAA GAC AAA GGA TAT Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr 1205	1210	1215	1152
ATG AGT ACA AGC TTA TCG AGT GAA CGT CTT GCA GCT TTT GGA TCT AGA Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg 1220	1225	1230	1200
AAA ATT ATA TTA CGA TTA CAA GTT CCG AAA GGA AGT ACG GGT GCG TAT Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr 1235	1240	1245	1248
TTA AGT GCC ATT GGT GGA TTT GCA AGT GAA AAA GAG ATC CTA CTT GAT			1296

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Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp			
1255	1260	1265	
AAA GAT AGT AAA TAT CAT ATT GAT AAA GTA ACA GAG GTA ATT ATT AAA			1344
Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys			
1270	1275	1280	
GGT GTT AAG CGA TAT GTA GTG GAT GCA ACA TTA TTA ACA AAT ATG AAA			1392
Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn Met Lys			
1285	1290	1295	
AAT ATG AAG AAA AAG TTA GCA AGT GTT GTA ACG TGT ACG TTA TTA GCT			1440
Asn Met Lys Lys Leu Ala Ser Val Val Thr Cys Thr Leu Leu Ala			
1300	1305	1310	
CCT ATG TTT TTG AAT GGA AAT GTG AAT GCT GTT TAC GCA GAC AGC AAA			1488
Pro Met Phe Leu Asn Gly Asn Val Asn Ala Val Tyr Ala Asp Ser Lys			
1315	1320	1325	1330
ACA AAT CAA ATT TCT ACA ACA CAG AAA AAT CAA CAG AAA GAG ATG GAC			1536
Thr Asn Gln Ile Ser Thr Thr Gln Lys Asn Gln Gln Lys Glu Met Asp			
1335	1340	1345	
CGA AAA GGA TTA CTT GGG TAT TAT TTC AAA GGA AAA GAT TTT AGT AAT			1584
Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys Asp Phe Ser Asn			
1350	1355	1360	
CTT ACT ATG TTT GCA CCG ACA CGT GAT AGT ACT CTT ATT TAT GAT CAA			1632
Leu Thr Met Phe Ala Pro Thr Arg Asp Ser Thr Leu Ile Tyr Asp Gln			
1365	1370	1375	
CAA ACA GCA AAT AAA CTA TTA GAT AAA AAA CAA CAA GAA TAT CAG TCT			1680
Gln Thr Ala Asn Lys Leu Leu Asp Lys Lys Gln Gln Glu Tyr Gln Ser			
1380	1385	1390	
ATT CGT TGG ATT GGT TTG ATT CAG AGT AAA GAA ACG GGA GAT TTC ACA			1728
Ile Arg Trp Ile Gly Leu Ile Gln Ser Lys Glu Thr Gly Asp Phe Thr			
1395	1400	1405	1410
TTC AAC TTA TCT GAG GAT GAA CAG GCA ATT ATA GAA ATC AAT GGG AAA			1776
Phe Asn Leu Ser Glu Asp Glu Gln Ala Ile Ile Glu Ile Asn Gly Lys			
1415	1420	1425	
ATT ATT TCT AAT AAA GGG AAA GAA AAG CAA GAT GTT GTC CAT TTA GAA AAA			1824
Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val His Leu Glu Lys			
1430	1435	1440	
GGA AAA TTA GTT CCA ATC AAA ATA GAG TAT CAA TCA GAT ACA AAA TTT			1872
Gly Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser Asp Thr Lys Phe			
1445	1450	1455	
AAT ATT GAC AGT AAA ACA TTT AAA GAA CTT AAA TTA TTT AAA ATA GAT			1920
Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu Phe Lys Ile Asp			
1460	1465	1470	

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AGT CAA AAC CAA CCC CAG CAA GTC CAG CAA GAT GAA CTG AGA AAT CCT Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu Leu Arg Asn Pro 1475 1480 1485 1490	1968
GAA TTT AAC AAG AAA GAA TCA CAG GAA TTC TTA GCG AAA CCA TCG AAA Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Pro Ser Lys 1495 1500 1505	2016
ATA AAT CTT TTC ACT CAA AAA ATG AAA AGG GAA ATT GAT GAA GAC ACG Ile Asn Leu Phe Thr Gln Lys Met Lys Arg Glu Ile Asp Glu Asp Thr 1510 1515 1520	2064
GAT ACG GAT GGG GAC TCT ATT CCT GAC CTT TGG GAA GAA AAT GGG TAT Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn Gly Tyr 1525 1530 1535	2112
ACG ATT CAA AAT AGA ATC GCT GTA AAG TGG GAC GAT TCT CTA GCA AGT Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp Ser Leu Ala Ser 1540 1545 1550	2160
AAA GGG TAT ACG AAA TTT GTT TCA AAT CCA CTA GAA AGT CAC ACA GTT Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu Ser His Thr Val 1555 1560 1565 1570	2208
GGT GAT CCT TAT ACA GAT TAT GAA AAG GCA GCA AGA GAT CTA GAT TTG Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu Asp Leu 1575 1580 1585	2256
TCA AAT GCA AAG GAA ACG TTT AAC CCA TTG GTA GCT GCT TTT CCA AGT Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe Pro Ser 1590 1595 1600	2304
GTG AAT GTT AGT ATG GAA AAG GTG ATA TTA TCA CCA AAT GAA AAT TTA Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu Asn Leu 1605 1610 1615	2352
TCC AAT AGT GTA GAG TCT CAT TCA TCC ACG AAT TGG TCT TAT ACA AAT Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr Thr Asn 1620 1625 1630	2400
ACA GAA GGT GCT TCT GTT GAA GCG GGG ATT GGA CCA AAA GGT ATT TCG Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro Lys Gly Ile Ser 1635 1640 1645 1650	2448
TTC GGA GTT AGC GTA AAC TAT CAA CAC TCT GAA ACA GTT GCA CAA GAA Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr Val Ala Gln Glu 1655 1660 1665	2496
TGG GGA ACA TCT ACA GGA AAT ACT TCG CAA TTC AAT ACG GCT TCA GCG Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala Ser Ala 1670 1675 1680	2544
GGA TAT TTA AAT GCA AAT GTT CGA TAT AAC AAT GTA GGA ACT GGT GCC Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr Gly Ala 1685 1690 1695	2592

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ATC TAC GAT GTA AAA CCT ACA ACA AGT TTT GTA TTA AAT AAC GAT ACT Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn Asp Thr 1700 1705 1710	2640
ATC GCA ACT ATT ACG GCG AAA TCT AAT TCT ACA GCC TTA AAT ATA TCT Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Asn Ile Ser 1715 1720 1725 1730	2688
CCT GGA GAA AGT TAC CCG AAA AAA GGA CAA AAT GGA ATC GCA ATA ACA Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly Ile Ala Ile Thr 1735 1740 1745	2736
TCA ATG GAT GAT TTT AAT TCC CAT CCG ATT ACA TTA AAT AAA AAA CAA Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys Lys Gln 1750 1755 1760	2784
GTA GAT AAT CTG CTA AAT AAT AAA CCT ATG ATG TTG GAA ACA AAC CAA Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu Glu Thr Asn Gln 1765 1770 1775	2832
ACA GAT GGT GTT TAT AAG ATA AAA GAT ACA CAT GGA AAT ATA GTA ACT Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly Asn Ile Val Thr 1780 1785 1790	2880
GGC GGA GAA TGG AAT GGT GTC ATA CAA CAA ATC AAG GCT AAA ACA GCG Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys Ala Lys Thr Ala 1795 1800 1805 1810	2928
TCT ATT ATT GTG GAT GAT GGG GAA CGT GTA GCA GAA AAA CGT GTA GCG Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu Lys Arg Val Ala 1815 1820 1825	2976
GCA AAA GAT TAT GAA AAT CCA GAA GAT AAA ACA CCG TCT TTA ACT TTA Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro Ser Leu Thr Leu 1830 1835 1840	3024
AAA GAT GCC CTG AAG CTT TCA TAT CCA GAT GAA ATA AAA GAA ATA GAG Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu Ile Glu 1845 1850 1855	3072
GGA TTA TTA TAT TAT AAA AAC AAA CCG ATA TAC GAA TCG AGC GTT ATG Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu Ser Ser Val Met 1860 1865 1870	3120
ACT TAC TTA GAT GAA AAT ACA GCA AAA GAA GTG ACC AAA CAA TTA AAT Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr Lys Gln Leu Asn 1875 1880 1885 1890	3168
GAT ACC ACT GGG AAA TTT AAA GAT GTA AGT CAT TTA TAT GAT GTA AAA Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu Tyr Asp Val Lys 1895 1900 1905	3216
CTG ACT CCA AAA ATG AAT GTT ACA ATC AAA TTG TCT ATA CTT TAT GAT Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser Ile Leu Tyr Asp	3264

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1910	1915	1920	
AAT GCT GAG TCT AAT GAT AAC TCA ATT GGT AAA TGG ACA AAC ACA AAT Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp Thr Asn Thr Asn 1925	1930	1935	3312
ATT GTT TCA GGT GGA AAT AAC GGA AAA AAA CAA TAT TCT TCT AAT AAT Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr Ser Ser Asn Asn 1940	1945	1950	3360
CCG GAT GCT AAT TTG ACA TTA AAT ACA GAT GCT CAA GAA AAA TTA AAT Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln Glu Lys Leu Asn 1955	1960	1965	3408
AAA AAT CGT GAC TAT TAT ATA AGT TTA TAT ATG AAG TCA GAA AAA AAC Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys Ser Glu Lys Asn 1975	1980	1985	3456
ACA CAA TGT GAG ATT ACT ATA GAT GGG GAG ATT TAT CCG ATC ACT ACA Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr Pro Ile Thr Thr 1990	1995	2000	3504
AAA ACA GTG AAT GTG AAT AAA GAC AAT TAC AAA AGA TTA GAT ATT ATA Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg Leu Asp Ile Ile 2005	2010	2015	3552
GCT CAT AAT ATA AAA AGT AAT CCA ATT TCT TCA CTT CAT ATT AAA ACG Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu His Ile Lys Thr 2020	2025	2030	3600
AAT GAT GAA ATA ACT TTA TTT TGG GAT GAT ATT TCT ATA ACA GAT GTA Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser Ile Thr Asp Val 2035	2040	2045	3648
GCA TCA ATA AAA CCG GAA AAT TTA ACA GAT TCA GAA ATT AAA CAG ATT Ala Ser Ile Lys Pro Glu Asn Leu Thr Asp Ser Glu Ile Lys Gln Ile 2055	2060	2065	3696
TAT AGT AGG TAT GGT ATT AAG TTA GAA GAT GGA ATC CTT ATT GAT AAA Tyr Ser Arg Tyr Gly Ile Lys Leu Glu Asp Gly Ile Leu Ile Asp Lys 2070	2075	2080	3744
AAA GGT GGG ATT CAT TAT GGT GAA TTT ATT AAT GAA GCT AGT TTT AAT Lys Gly Gly Ile His Tyr Gly Glu Phe Ile Asn Glu Ala Ser Phe Asn 2085	2090	2095	3792
ATT GAA CCA TTG CAA AAT TAT GTG ACC AAA TAT GAA GTT ACT TAT AGT Ile Glu Pro Leu Gln Asn Tyr Val Thr Lys Tyr Glu Val Thr Tyr Ser 2100	2105	2110	3840
AGT GAG TTA GGA CCA AAC GTG AGT GAC ACA CTT GAA AGT GAT AAA ATT Ser Glu Leu Gly Pro Asn Val Ser Asp Thr Leu Glu Ser Asp Lys Ile 2115	2120	2125	3888
TAC AAG GAT GGG ACA ATT AAA TTT GAT TTT ACC AAA TAT AGT AAA AAT			3936

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Tyr Lys Asp Gly Thr Ile Lys Phe Asp Phe Thr Lys Tyr Ser Lys Asn
2135 2140 2145

GAA CAA GGA TTA TTT TAT GAC AGT GGA TTA AAT TGG GAC TTT AAA ATT 3984
Glu Gln Gly Leu Phe Tyr Asp Ser Gly Leu Asn Trp Asp Phe Lys Ile
2150 2155 2160

AAT GCT ATT ACT TAT GAT GGT AAA GAG ATG AAT GTT TTT CAT AGA TAT 4032
Asn Ala Ile Thr Tyr Asp Gly Lys Glu Met Asn Val Phe His Arg Tyr
2165 2170 2175

AAT AAA TAG 4041
Asn Lys
2180

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1346 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys Leu Gln
1 5 10 15

Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile Ser Leu
20 25 30

Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn Ser Gln
35 40 45

Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu
50 55 60

Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys
65 70 75 80

Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn
85 90 95

Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr
100 105 110

Phe Ser Met Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu
115 120 125

Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr
130 135 140

Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr

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145	150	155	160												
Glu	Gly	Asn	Thr	Ile	Asn	Ser	Asp	Ala	Met	Ala	Gln	Phe	Lys	Glu	Gln
				165					170				175		
Phe	Leu	Asp	Arg	Asp	Ile	Lys	Phe	Asp	Ser	Tyr	Leu	Asp	Thr	His	Leu
				180				185				190			
Thr	Ala	Gln	Gln	Val	Ser	Ser	Lys	Glu	Arg	Val	Ile	Leu	Lys	Val	Thr
				195			200				205				
Val	Pro	Ser	Gly	Lys	Gly	Ser	Thr	Thr	Pro	Thr	Lys	Ala	Gly	Val	Ile
				210			215			220					
Leu	Asn	Asn	Ser	Glu	Tyr	Lys	Met	Leu	Ile	Asp	Asn	Gly	Tyr	Met	Val
				225			230			235			240		
His	Val	Asp	Lys	Val	Ser	Lys	Val	Val	Lys	Lys	Gly	Val	Glu	Cys	Leu
				245			250			255					
Gln	Ile	Glu	Gly	Thr	Leu	Lys	Ser	Leu	Asp	Phe	Lys	Asn	Asp	Ile	
				260			265			270					
Asn	Ala	Glu	Ala	His	Ser	Trp	Gly	Met	Lys	Asn	Tyr	Glu	Glu	Trp	Ala
				275			280			285					
Lys	Asp	Leu	Thr	Asp	Ser	Gln	Arg	Glu	Ala	Leu	Asp	Gly	Tyr	Ala	Arg
				290			295			300					
Gln	Asp	Tyr	Lys	Glu	Ile	Asn	Asn	Tyr	Leu	Arg	Asn	Gln	Gly	Ser	
				305			310			315			320		
Gly	Asn	Glu	Lys	Leu	Asp	Ala	Gln	Ile	Lys	Asn	Ile	Ser	Asp	Ala	Leu
				325			330			335					
Gly	Lys	Lys	Pro	Ile	Pro	Glu	Asn	Ile	Thr	Val	Tyr	Arg	Trp	Cys	Gly
				340			345			350					
Met	Pro	Glu	Phe	Gly	Tyr	Gln	Ile	Ser	Asp	Pro	Leu	Pro	Ser	Leu	Lys
				355			360			365					
Asp	Phe	Glu	Glu	Gln	Phe	Leu	Asn	Thr	Ile	Lys	Glu	Asp	Lys	Gly	Tyr
				370			375			380					
Met	Ser	Thr	Ser	Leu	Ser	Ser	Glu	Arg	Leu	Ala	Ala	Phe	Gly	Ser	Arg
				385			390			395			400		
Lys	Ile	Ile	Leu	Arg	Leu	Gln	Val	Pro	Lys	Gly	Ser	Thr	Gly	Ala	Tyr
				405			410			415					
Leu	Ser	Ala	Ile	Gly	Gly	Phe	Ala	Ser	Glu	Lys	Glu	Ile	Leu	Leu	Asp
				420			425			430					
Lys	Asp	Ser	Lys	Tyr	His	Ile	Asp	Lys	Val	Thr	Glu	Val	Ile	Ile	Lys
				435			440			445					

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Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn Met Lys
450 455 460

Asn Met Lys Lys Lys Leu Ala Ser Val Val Thr Cys Thr Leu Leu Ala
465 470 475 480

Pro Met Phe Leu Asn Gly Asn Val Asn Ala Val Tyr Ala Asp Ser Lys
485 490 495

Thr Asn Gln Ile Ser Thr Thr Gln Lys Asn Gln Gln Lys Glu Met Asp
500 505 510

Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys Asp Phe Ser Asn
515 520 525

Leu Thr Met Phe Ala Pro Thr Arg Asp Ser Thr Leu Ile Tyr Asp Gln
530 535 540

Gln Thr Ala Asn Lys Leu Leu Asp Lys Lys Gln Gln Glu Tyr Gln Ser
545 550 555 560

Ile Arg Trp Ile Gly Leu Ile Gln Ser Lys Glu Thr Gly Asp Phe Thr
565 570 575

Phe Asn Leu Ser Glu Asp Glu Gln Ala Ile Ile Glu Ile Asn Gly Lys
580 585 590

Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val His Leu Glu Lys
595 600 605

Gly Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser Asp Thr Lys Phe
610 615 620

Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu Phe Lys Ile Asp
625 630 635 640

Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu Leu Arg Asn Pro
645 650 655

Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Pro Ser Lys
660 665 670

Ile Asn Leu Phe Thr Gln Lys Met Lys Arg Glu Ile Asp Glu Asp Thr
675 680 685

Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn Gly Tyr
690 695 700

Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp Ser Leu Ala Ser
705 710 715 720

Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu Ser His Thr Val
725 730 735

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Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu Asp Leu
740 745 750

Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe Pro Ser
755 760 765

Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu Asn Leu
770 775 780

Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr Thr Asn
785 790 795 800

Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro Lys Gly Ile Ser
805 810 815

Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr Val Ala Gln Glu
820 825 830

Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala Ser Ala
835 840 845

Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr Gly Ala
850 855 860

Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn Asp Thr
865 870 875 880

Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Asn Ile Ser
885 890 895

Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly Ile Ala Ile Thr
900 905 910

Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys Lys Gln
915 920 925

Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu Glu Thr Asn Gln
930 935 940

Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly Asn Ile Val Thr
945 950 955 960

Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys Ala Lys Thr Ala
965 970 975

Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu Lys Arg Val Ala
980 985 990

Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro Ser Leu Thr Leu
995 1000 1005

Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu Ile Glu
1010 1015 1020

Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu Ser Ser Val Met

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1025	1030	1035	1040
Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr Lys Gln Leu Asn			
1045	1050	1055	
Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu Tyr Asp Val Lys			
1060	1065	1070	
Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser Ile Leu Tyr Asp			
1075	1080	1085	
Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp Thr Asn Thr Asn			
1090	1095	1100	
Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr Ser Ser Asn Asn			
1105	1110	1115	1120
Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln Glu Lys Leu Asn			
1125	1130	1135	
Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys Ser Glu Lys Asn			
1140	1145	1150	
Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr Pro Ile Thr Thr			
1155	1160	1165	
Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg Leu Asp Ile Ile			
1170	1175	1180	
Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu His Ile Lys Thr			
1185	1190	1195	1200
Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser Ile Thr Asp Val			
1205	1210	1215	
Ala Ser Ile Lys Pro Glu Asn Leu Thr Asp Ser Glu Ile Lys Gln Ile			
1220	1225	1230	
Tyr Ser Arg Tyr Gly Ile Lys Leu Glu Asp Gly Ile Leu Ile Asp Lys			
1235	1240	1245	
Lys Gly Gly Ile His Tyr Gly Glu Phe Ile Asn Glu Ala Ser Phe Asn			
1250	1255	1260	
Ile Glu Pro Leu Gln Asn Tyr Val Thr Lys Tyr Glu Val Thr Tyr Ser			
1265	1270	1275	1280
Ser Glu Leu Gly Pro Asn Val Ser Asp Thr Leu Glu Ser Asp Lys Ile			
1285	1290	1295	
Tyr Lys Asp Gly Thr Ile Lys Phe Asp Phe Thr Lys Tyr Ser Lys Asn			
1300	1305	1310	
Glu Gln Gly Leu Phe Tyr Asp Ser Gly Leu Asn Trp Asp Phe Lys Ile			
1315	1320	1325	

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Asn Ala Ile Thr Tyr Asp Gly Lys Glu Met Asn Val Phe His Arg Tyr
1330 1335 1340

Asn Lys
1345

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1399 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..1386
- (D) OTHER INFORMATION: /note= "Maize optimized DNA sequence for VIP2A(a) protein from AB78"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

ATGAAGCGCA TGGAGGGCAA GCTGTTCATG GTGAGCAAGA AGCTCCAGGT GGTGACCAAG	60
ACCGTGCTGC TGAGCACCGT GTTCAGCATC AGCCTGCTGA ACAACGAGGT GATCAAGGCC	120
GAGCAGCTGA ACATCAACAG CCAGAGCAAG TACACCAACC TCCAGAACCT GAAGATCACC	180
GACAAGGTGG AGGACTTCAA GGAGGACAAG GAGAAGGCCA AGGAGTGGGG CAAGGAGAAAG	240
GAGAAGGAGT GGAAGCTTAC CGCCACCGAG AAGGGCAAGA TGAACAACTT CCTGGACAAC	300
AAGAACGACA TCAAGACCAA CTACAAGGAG ATCACCTTCA GCATGGCCGG CAGCTTCGAG	360
GACGAGATCA AGGACCTGAA GGAGATCGAC AAGATGTTCG ACAAGACCAA CCTGAGCAAC	420
AGCATCATCA CCTACAAGAA CGTGGAGCCC ACCACCATCG GCTTCAACAA GAGCCTGACC	480
GAGGGCAACA CCATCAACAG CGACGCCATG GCCCAGTTCA AGGAGCAGTT CCTGGACCGC	540
GACATCAAGT TCGACAGCTA CCTGGACACC CACCTGACCG CCCAGCAGGT GAGCAGCAAG	600
GAGCGCGTGA TCCTGAAGGT GACCGTCCCC AGCGGCAAGG GCAGCACCAC CCCCACCAAG	660
GCCGGCGTGA TCCTGAACAA CAGCGAGTAC AAGATGCTGA TCGACAAACGG CTACATGGTG	720
CACGTGGACA AGGTGAGCAA GGTGGTGAAG AAGGGCGTGG AGTGCCTCCA GATCGAGGGC	780
ACCCCTGAAGA AGAGTCTAGA CTTCAAGAAC GACATCAACG CCGAGGCCA CAGCTGGGGC	840

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ATGAAGAACT ACGAGGAGTG GGCCAAGGAC CTGACCGACA GCCAGCGCGA GGCCCTGGAC	900
GGCTACGCC C GCCAGGACTA CAAGGAGATC AACAACTACC TGCGCAACCA GGGCGGCAGC	960
GGCAACGAGA AGCTGGACGC CCAGATCAAG AACATCAGCG ACGCCCTGGG CAAGAAGCCC	1020
ATCCCCGAGA ACATCACCGT GTACCGCTGG TGCGGCATGC CCGAGTTCGG CTACCAGATC	1080
AGCGACCCCC TGCCCAGCCT GAAGGACTTC GAGGAGCAGT TCCTGAACAC CATCAAGGAG	1140
GACAAGGGCT ACATGAGCAC CAGCCTGAGC AGCGAGCGCC TGGCCGCCTT CGGCAGCCGC	1200
AAGATCATCC TGCGCCTGCA GGTGCCCAAG GGCAGCACCG GCGCCTACCT GAGCGCCATC	1260
GGCGGCTTCG CCAGCGAGAA GGAGATCCTG CTGGACAAGG ACAGCAAGTA CCACATCGAC	1320
AAGGTGACCG AGGTGATCAT CAAGGGCGTG AAGCGCTACG TGGTGGACGC CACCCCTGCTG	1380
ACCAACTAGA TCTGAGCTC	1399

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..19
- (D) OTHER INFORMATION: /note= "Secretion signal peptide to secrete VIP2 out of a cell"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Ala Ala Gly Val			
1	5	10	15

His Cys Leu

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2655 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

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(A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: misc_feature

(B) LOCATION: 1..2655

(D) OTHER INFORMATION: /note= "maize optimized DNA sequence encoding VIP1A(a)"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

ATGAAGAAC	TGAAGAAGAA	GCTGGCCAGC	GTGGTGACCT	GCACCCCTGCT	GGCCCCCATG	60
TTCCCTGAACG	GCAACGTGAA	CGCCGTGTAC	GCCGACAGCA	AGACCAACCA	GATCAGCACC	120
ACCCAGAAGA	ACCAGCAGAA	GGAGATGGAC	CGCAAGGGCC	TGCTGGGCTA	CTACTTCAAG	180
GGCAAGGACT	TCAGCAACCT	GACCATGTTTC	GCCCCCACGC	GTGACAGCAC	CCTGATCTAC	240
GACCAGCAGA	CCGCCAACAA	GCTGCTGGAC	AAGAAGCAGC	AGGAGTACCA	GAGCATCCGC	300
TGGATCGGCC	TGATCCAGAG	CAAGGAGACC	GGCGACTTCA	CCTTCAACCT	GAGCGAGGAC	360
GAGCAGGCCA	TCATCGAGAT	CAACGGCAAG	ATCATCAGCA	ACAAGGGCAA	GGAGAAGCAG	420
GTGGTGCACC	TGGAGAAGGG	CAAGCTGGTG	CCCATCAAGA	TCGAGTACCA	GAGCGACACC	480
AAGTTCAACA	TCGACAGCAA	GACTTCAAG	GAGCTGAAGC	TTTTCAAGAT	CGACAGCCAG	540
AACCAGCCCC	AGCAGGTGCA	GCAGGACGAG	CTGCGCAACC	CCGAGTTCAA	CAAGAAGGAG	600
AGCCAGGAGT	TCCTGGCAA	GCCCAGCAAG	ATCAACCTGT	TCACCCAGCA	GATGAAGCGC	660
GAGATCGACG	AGGACACCGA	CACCGACGGC	GACAGCATCC	CCGACCTGTG	GGAGGAGAAC	720
GGCTACACCA	TCCAGAACCG	CATCGCCGTG	AAAGTGGGACG	ACAGCCTGGC	TAGCAAGGGC	780
TACACCAAGT	TCGTGAGCAA	CCCCCTGGAG	AGCCACACCG	TGGGGACCCC	CTACACCGAC	840
TACGAGAAGG	CCGCCCGCGA	CCTGGACCTG	AGCAACGCCA	AGGAGACCTT	CAACCCCCTG	900
GTGGCCGCCT	TCCCCAGCGT	GAACGTGAGC	ATGGAGAAGG	TGATCCTGAG	CCCCAACGAG	960
AACCTGAGCA	ACAGCGTGGA	GAGCCACTCG	AGCACCAACT	GGAGCTACAC	CAACACCGAG	1020
GGCGCCAGCG	TGGAGGCCGG	CATCGGTCCC	AAGGGCATCA	GCTTCGGCGT	GAGCGTGAAC	1080
TACCAGCACA	GCGAGACCGT	GGCCCAGGAG	TGGGGCACCA	GCACCGGCAA	CACCAGCCAG	1140
TTCAACACCG	CCAGCGCCGG	CTACCTGAAC	GCCAACGTGC	GCTACAACAA	CGTGGGCACC	1200
GGCGCCATCT	ACGACGTGAA	GCCCACCAACC	AGCTTCGTGC	TGAACAAACGA	CACCATGCC	1260

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ACCATCACCG CCAAGTCGAA TTCCACCGCC CTGAACATCA GCCCCGGCGA GAGCTACCCC	1320
AAGAAGGGCC AGAACGGCAT CGCCATCACC AGCATGGACG ACTTCAACAG CCACCCCATC	1380
ACCCCTGAACA AGAACGCAGGT GGACAACCTG CTGAACAAACA AGCCCATGAT GCTGGAGACC	1440
AACCAGACCG ACGGCGTCTA CAAGATCAAG GACACCCACG GCAACATCGT GACGGGCGGC	1500
GAGTGGAACG GCGTGATCCA GCAGATCAAG GCCAAGACCG CCAGCATCAT CGTCGACGAC	1560
GGCGAGCGCG TGGCCGAGAA GCGCGTGGCC GCCAAGGACT ACGAGAACCC CGAGGACAAG	1620
ACCCCCAGCC TGACCCCTGAA GGACGCCCTG AAGCTGAGCT ACCCCGACGA GATCAAGGAG	1680
ATCGAGGGCT TGCTGTACTA CAAGAACAAAG CCCATCTACG AGAGCAGCGT GATGACCTAT	1740
CTAGACGAGA ACACCGCCAA GGAGGTGACC AAGCAGCTGA ACGACACCAAC CGGCAAGTTC	1800
AAGGACGTGA GCCACCTGTA CGACGTGAAG CTGACCCCCA AGATGAACGT GACCATCAAG	1860
CTGAGCATCC TGTACGACAA CGCCGAGAGC AACGACAACA GCATCGGCAA GTGGACCAAC	1920
ACCAACATCG TGAGCGCGG CAACAACGGC AAGAACAGT ACAGCAGCAA CAACCCGAC	1980
GCCAACCTGA CCCTGAACAC CGACGCCAG GAGAAGCTGA ACAAGAACCG CGACTACTAC	2040
ATCAGCCTGT ACATGAAGAG CGAGAAGAAC ACCCAGTGCAG AGATCACCAC CGACGGCGAG	2100
ATATAACCCCA TCACCACCAA GACCGTGAAC GTGAACAAGG ACAACTACAA GCGCCTGGAC	2160
ATCATCGCCC ACAACATCAA GAGCAACCCC ATCAGCAGCC TGCACATCAA GACCAACGAC	2220
GAGATCACCC TGTCTGGGA CGACATATCG ATTACCGACG TCGCCAGCAT CAAGCCCGAG	2280
AACCTGACCG ACAGCGAGAT CAAGCAGATA TACAGTCGCT ACGGCATCAA GCTGGAGGAC	2340
GGCATCCTGA TCGACAAGAA AGGCGGCATC CACTACGGCG AGTTCATCAA CGAGGCCAGC	2400
TTCAACATCG AGCCCCCTGCA GAACTACGTG ACCAAGTACG AGGTGACCTA CAGCAGCGAG	2460
CTGGGCCCCA ACGTGAGCGA CACCCCTGGAG AGCGACAAGA TTTACAAGGA CGGCACCATC	2520
AAGTTCGACT TCACCAAGTA CAGCAAGAAC GAGCAGGGCC TGTCTACGA CAGCGGCCTG	2580
AACTGGGACT TCAAGATCAA CGCCATCACC TACGACGGCA AGGAGATGAA CGTGTCCAC	2640
CGCTACAACA AGTAG	2655

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1389 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: misc_feature
 (B) LOCATION: 1..1389
 (D) OTHER INFORMATION: /note= "maize optimized DNA sequence encoding VIP2A(a)"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

ATGAAGCGCA	TGGAGGGCAA	GCTGTTCATG	GTGAGCAAGA	AGCTCCAGGT	GGTGACCAAG	60
ACCGTGCTGC	TGAGCACCGT	GTTCAGCATT	AGCCTGCTGA	ACAACGAGGT	GATCAAGGCC	120
GAGCAGCTGA	ACATCAACAG	CCAGAGCAAG	TACACCAACC	TCCAGAACCT	GAAGATCACC	180
GACAAGGTGG	AGGACTTCAA	GGAGGACAAG	GAGAAGGCCA	AGGAGTGGGG	CAAGGAGAAG	240
GAGAAGGAGT	GGAAGCTTAC	CGCCACCGAG	AAGGGCAAGA	TGAACAACTT	CCTGGACAAC	300
AAGAACGACA	TCAAGACCAA	CTACAAGGAG	ATCACCTTCA	GCATAGCCGG	CAGCTTCGAG	360
GACGAGATCA	AGGACCTGAA	GGAGATCGAC	AAGATGTTCG	ACAAGACCAA	CCTGAGCAAC	420
AGCATCATCA	CCTACAAGAA	CGTGGAGCCC	ACCACCATCG	GCTTCAACAA	GAGCCTGACC	480
GAGGGCAACA	CCATCAACAG	CGACGCCATG	GCCCAGTTCA	AGGAGCAGTT	CCTGGACCGC	540
GACATCAAGT	TCGACAGCTA	CCTGGACACC	CACCTGACCG	CCCAGCAGGT	GAGCAGCAAG	600
GAGCGCGTGA	TCCTGAAGGT	GACCGTCCCC	AGCGGCAAGG	GCAGCACCAC	CCCCACCAAG	660
GCCGGCGTGA	TCCTGAACAA	CAGCGAGTAC	AAGATGCTGA	TCGACAACGG	CTACATGGTG	720
CACGTGGACA	AGGTGAGCAA	GGTGGTGAAG	AAGGGCGTGG	AGTGCCTCCA	GATCGAGGGC	780
ACCTGAAAGA	AGAGTCTAGA	CTTCAAGAAC	GACATCAACG	CCGAGGCCCA	CAGCTGGGGC	840
ATGAAGAACT	ACGAGGGAGTG	GGCCAAGGAC	CTGACCGACA	GCCAGCGCGA	GGCCCTGGAC	900
GGCTACGCC	GCCAGGACTA	CAAGGAGATC	AACAACTACC	TGCGCAACCA	GGGCGGCAGC	960
GGCAACGAGA	AGCTGGACGC	CCAGATCAAG	AACATCAGCG	ACGCCCTGGG	CAAGAAGCCC	1020
ATCCCCGAGA	ACATCACCGT	GTACCGCTGG	TGCGGCATGC	CCGAGTTCGG	CTACCAAGATC	1080
AGCGACCCCC	TGCCAGCCT	GAAGGACTTC	GAGGAGCAGT	TCCTGAACAC	CATCAAGGAG	1140

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GACAAGGGCT ACATGAGCAC CAGCCTGAGC AGCGAGCGCC TGGCCGCCCTT CGGCAGCCGC	1200
AAGATCATCC TGCGCCTGCA GGTGCCCAAG GGCAGCACTG GTGCCTACCT GAGGCCATC	1260
GGCGGCTTCG CCAGCGAGAA GGAGATCCTG CTGGATAAGG ACAGCAAGTA CCACATCGAC	1320
AAGGTGACCG AGGTGATCAT CAAGGGCGTG AAGCGCTACG TGGTGGACGC CACCCGTG	1380
ACCAACTAG	1389

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2378 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: CDS	
(B) LOCATION: 9..2375	
(D) OTHER INFORMATION: /note= "Native DNA sequence encoding VIP3A(a) protein from AB88 as contained in pCIB7104"	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

AGATGAAC ATG AAC AAG AAT AAT ACT AAA TTA AGC ACA AGA GCC TTA CCA	50
Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro	
1 5 10	
AGT TTT ATT GAT TAT TTT AAT GGC ATT TAT GGA TTT GCC ACT GGT ATC	98
Ser Phe Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile	
15 20 25 30	
AAA GAC ATT ATG AAC ATG ATT TTT AAA ACG GAT ACA GGT GGT GAT CTA	146
Lys Asp Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu	
35 40 45	
ACC CTA GAC GAA ATT TTA AAG AAT CAG CAG TTA CTA AAT GAT ATT TCT	194
Thr Leu Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn Asp Ile Ser	
50 55 60	
GGT AAA TTG GAT GGG GTG AAT GGA AGC TTA AAT GAT CTT ATC GCA CAG	242
Gly Lys Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln	
65 70 75	
GGA AAC TTA AAT ACA GAA TTA TCT AAG GAA ATA TTA AAA ATT GCA AAT	290
Gly Asn Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn	
80 85 90	

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GAA CAA AAT CAA GTT TTA AAT GAT GTT AAT AAC AAA CTC GAT GCG ATA Glu Gln Asn Gln Val Leu Asn Asp Val Asn Lys Leu Asp Ala Ile 95 100 105 110	338
AAT ACG ATG CTT CGG GTA TAT CTA CCT AAA ATT ACC TCT ATG TTG AGT Asn Thr Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser 115 120 125	386
GAT GTA ATG AAA CAA AAT TAT GCG CTA AGT CTG CAA ATA GAA TAC TTA Asp Val Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu 130 135 140	434
AGT AAA CAA TTG CAA GAG ATT TCT GAT AAG TTG GAT ATT ATT AAT GTA Ser Lys Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val 145 150 155	482
AAT GTA CTT ATT AAC TCT ACA CTT ACT GAA ATT ACA CCT GCG TAT CAA Asn Val Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln 160 165 170	530
AGG ATT AAA TAT GTG AAC GAA AAA TTT GAG GAA TTA ACT TTT GCT ACA Arg Ile Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr 175 180 185 190	578
GAA ACT AGT TCA AAA GTA AAA AAG GAT GGC TCT CCT GCA GAT ATT CTT Glu Thr Ser Ser Lys Val Lys Asp Gly Ser Pro Ala Asp Ile Leu 195 200 205	626
GAT GAG TTA ACT GAG TTA ACT GAA CTA GCG AAA AGT GTA ACA AAA AAT Asp Glu Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn 210 215 220	674
GAT GTG GAT GGT TTT GAA TTT TAC CTT AAT ACA TTC CAC GAT GTA ATG Asp Val Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met 225 230 235	722
GTA GGA AAT AAT TTA TTC GGG CGT TCA GCT TTA AAA ACT GCA TCG GAA Val Gly Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu 240 245 250	770
TTA ATT ACT AAA GAA AAT GTG AAA ACA AGT GGC AGT GAG GTC GGA AAT Leu Ile Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu Val Gly Asn 255 260 265 270	818
GTT TAT AAC TTC TTA ATT GTA TTA ACA GCT CTG CAA GCC CAA GCT TTT Val Tyr Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala Gln Ala Phe 275 280 285	866
CTT ACT TTA ACA ACA TGC CGA AAA TTA TTA GGC TTA GCA GAT ATT GAT Leu Thr Leu Thr Thr Cys Arg Lys Leu Leu Gly Leu Ala Asp Ile Asp 290 295 300	914
TAT ACT TCT ATT ATG AAT GAA CAT TTA AAT AAG GAA AAA GAG GAA TTT Tyr Thr Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys Glu Glu Phe	962

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305	310	315	
AGA GTA AAC ATC CTC CCT ACA CTT TCT AAT ACT TTT TCT AAT CCT AAT Arg Val Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser Asn Pro Asn	320	325	1010
TAT GCA AAA GTT AAA GGA AGT GAT GAA GAT GCA AAG ATG ATT GTG GAA Tyr Ala Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met Ile Val Glu	335	340	1058
GCT AAA CCA GGA CAT GCA TTG ATT GGG TTT GAA ATT AGT AAT GAT TCA Ala Lys Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser Asn Asp Ser	355	360	1106
ATT ACA GTA TTA AAA GTA TAT GAG GCT AAG CTA AAA CAA AAT TAT CAA Ile Thr Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln Asn Tyr Gln	370	375	1154
GTC GAT AAG GAT TCC TTA TCG GAA GTT ATT TAT GGT GAT ATG GAT AAA Val Asp Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp Met Asp Lys	385	390	1202
TTA TTG TGC CCA GAT CAA TCT GAA CAA ATC TAT TAT ACA AAT AAC ATA Leu Leu Cys Pro Asp Gln Ser Glu Gln Ile Tyr Thr Asn Asn Ile	400	405	1250
GTA TTT CCA AAT GAA TAT GTA ATT ACT AAA ATT GAT TTC ACT AAA AAA Val Phe Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe Thr Lys Lys	415	420	1298
ATG AAA ACT TTA AGA TAT GAG GTA ACA GCG AAT TTT TAT GAT TCT TCT Met Lys Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr Asp Ser Ser	435	440	1346
ACA GGA GAA ATT GAC TTA AAT AAG AAA AAA GTA GAA TCA AGT GAA GCG Thr Gly Glu Ile Asp Leu Asn Lys Lys Val Glu Ser Ser Glu Ala	450	455	1394
GAG TAT AGA ACG TTA AGT GCT AAT GAT GAT GGG GTG TAT ATG CCG TTA Glu Tyr Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr Met Pro Leu	465	470	1442
GGT GTC ATC AGT GAA ACA TTT TTG ACT CCG ATT AAT GGG TTT GGC CTC Gly Val Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly Phe Gly Leu	480	485	1490
CAA GCT GAT GAA AAT TCA AGA TTA ATT ACT TTA ACA TGT AAA TCA TAT Gln Ala Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys Lys Ser Tyr	495	500	1538
TTA AGA GAA CTA CTG CTA GCA ACA GAC TTA AGC AAT AAA GAA ACT AAA Leu Arg Glu Leu Leu Ala Thr Asp Leu Ser Asn Lys Glu Thr Lys	515	520	1586
TTG ATC GTC CCG CCA AGT GGT TTT ATT AGC AAT ATT GTA GAG AAC GGG			1634

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Leu Ile Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val Glu Asn Gly		
530	535	540
TCC ATA GAA GAG GAC AAT TTA GAG CCG TGG AAA GCA AAT AAT AAG AAT		1682
Ser Ile Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn Asn Lys Asn		
545	550	555
GCG TAT GTA GAT CAT ACA GGC GGA GTG AAT GGA ACT AAA GCT TTA TAT		1730
Ala Tyr Val Asp His Thr Gly Gly Val Asn Gly Thr Lys Ala Leu Tyr		
560	565	570
GTT CAT AAG GAC GGA GGA ATT TCA CAA TTT ATT GGA GAT AAG TTA AAA		1778
Val His Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp Lys Leu Lys		
575	580	585
CCG AAA ACT GAG TAT GTA ATC CAA TAT ACT GTT AAA GGA AAA CCT TCT		1826
Pro Lys Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly Lys Pro Ser		
595	600	605
ATT CAT TTA AAA GAT GAA AAT ACT GGA TAT ATT CAT TAT GAA GAT ACA		1874
Ile His Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr Glu Asp Thr		
610	615	620
AAT AAT AAT TTA GAA GAT TAT CAA ACT ATT AAT AAA CGT TTT ACT ACA		1922
Asn Asn Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg Phe Thr Thr		
625	630	635
GGA ACT GAT TTA AAG GGA GTG TAT TTA ATT TTA AAA AGT CAA AAT GGA		1970
Gly Thr Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser Gln Asn Gly		
640	645	650
GAT GAA GCT TGG GGA GAT AAC TTT ATT ATT TTG GAA ATT AGT CCT TCT		2018
Asp Glu Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile Ser Pro Ser		
655	660	665
670		
GAA AAG TTA TTA AGT CCA GAA TTA ATT AAT ACA AAT AAT TGG ACG AGT		2066
Glu Lys Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn Trp Thr Ser		
675	680	685
ACG GGA TCA ACT AAT ATT AGC GGT AAT ACA CTC ACT CTT TAT CAG GGA		2114
Thr Gly Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu Tyr Gln Gly		
690	695	700
GGA CGA GGG ATT CTA AAA CAA AAC CTT CAA TTA GAT AGT TTT TCA ACT		2162
Gly Arg Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser Phe Ser Thr		
705	710	715
TAT AGA GTG TAT TTT TCT GTG TCC GGA GAT GCT AAT GTA AGG ATT AGA		2210
Tyr Arg Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val Arg Ile Arg		
720	725	730
AAT TCT AGG GAA GTG TTA TTT GAA AAA AGA TAT ATG AGC GGT GCT AAA		2258
Asn Ser Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser Gly Ala Lys		
735	740	745
		750

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GAT GTT TCT GAA ATG TTC ACT ACA AAA TTT GAG AAA GAT AAC TTT TAT Asp Val Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp Asn Phe Tyr 755 760 765	2306
ATA GAG CTT TCT CAA GGG AAT AAT TTA TAT GGT GGT CCT ATT GTA CAT Ile Glu Leu Ser Gln Gly Asn Asn Leu Tyr Gly Gly Pro Ile Val His 770 775 780	2354
TTT TAC GAT GTC TCT ATT AAG TAA Phe Tyr Asp Val Ser Ile Lys 785	2378

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 789 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe 1 5 10 15
Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp 20 25 30
Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Thr Leu 35 40 45
Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn Asp Ile Ser Gly Lys 50 55 60
Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln Gly Asn 65 70 75 80
Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn Glu Gln 85 90 95
Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu Asp Ala Ile Asn Thr 100 105 110
Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser Asp Val 115 120 125
Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu Ser Lys 130 135 140
Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val Asn Val 145 150 155 160
Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile

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165	170	175
Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr Glu Thr 180	185	190
Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala Asp Ile Leu Asp Glu 195	200	205
Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn Asp Val 210	215	220
Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met Val Gly 225	230	235
Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu Leu Ile 245	250	255
Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu Val Gly Asn Val Tyr 260	265	270
Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala Gln Ala Phe Leu Thr 275	280	285
Leu Thr Thr Cys Arg Lys Leu Leu Gly Leu Ala Asp Ile Asp Tyr Thr 290	295	300
Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys Glu Glu Phe Arg Val 305	310	315
Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser Asn Pro Asn Tyr Ala 325	330	335
Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met Ile Val Glu Ala Lys 340	345	350
Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser Asn Asp Ser Ile Thr 355	360	365
Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln Asn Tyr Gln Val Asp 370	375	380
Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp Met Asp Lys Leu Leu 385	390	395
Cys Pro Asp Gln Ser Glu Gln Ile Tyr Tyr Thr Asn Asn Ile Val Phe 405	410	415
Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe Thr Lys Lys Met Lys 420	425	430
Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr Asp Ser Ser Thr Gly 435	440	445
Glu Ile Asp Leu Asn Lys Lys Val Glu Ser Ser Glu Ala Glu Tyr 450	455	460

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Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr Met Pro Leu Gly Val
465 470 475 480

Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly Phe Gly Leu Gln Ala
485 490 495

Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys Lys Ser Tyr Leu Arg
500 505 510

Glu Leu Leu Leu Ala Thr Asp Leu Ser Asn Lys Glu Thr Lys Leu Ile
515 520 525

Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val Glu Asn Gly Ser Ile
530 535 540

Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn Asn Lys Asn Ala Tyr
545 550 555 560

Val Asp His Thr Gly Gly Val Asn Gly Thr Lys Ala Leu Tyr Val His
565 570 575

Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp Lys Leu Lys Pro Lys
580 585 590

Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly Lys Pro Ser Ile His
595 600 605

Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr Glu Asp Thr Asn Asn
610 615 620

Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg Phe Thr Thr Gly Thr
625 630 635 640

Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser Gln Asn Gly Asp Glu
645 650 655

Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile Ser Pro Ser Glu Lys
660 665 670

Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn Trp Thr Ser Thr Gly
675 680 685

Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu Tyr Gln Gly Arg
690 695 700

Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser Phe Ser Thr Tyr Arg
705 710 715 720

Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val Arg Ile Arg Asn Ser
725 730 735

Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser Gly Ala Lys Asp Val
740 745 750

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Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp Asn Phe Tyr Ile Glu
 755 760 765

Leu Ser Gln Gly Asn Asn Leu Tyr Gly Gly Pro Ile Val His Phe Tyr
 770 775 780

Asp Val Ser Ile Lys
 785

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2403 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 11..2389
- (D) OTHER INFORMATION: /note= "maize optimized DNA sequence encoding VIP3A(a)"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGATCCACCA ATGAACATGA ACAAGAACAA CACCAAGCTG AGCACCCGCG CCCTGCCGAG	60
CTTCATCGAC TACTTCAACG GCATCTACGG CTTCGCCACC GGCACTCAAGG ACATCATGAA	120
CATGATCTTC AAGACCGACA CGGGCGGCGA CCTGACCCCTG GACGAGATCC TGAAGAACCA	180
GCAGCTGCTG AACGACATCA GCGGCAAGCT GGACGGCGTG AACGGCAGCC TGAACGACCT	240
GATGCCAGGG GGCAACCTGA ACACCGAGCT GAGCAAGGAG ATCCTTAAGA TCGCCAACGA	300
GCAGAACCAAG GTGCTGAACG ACGTGAACAA CAAGCTGGAC GCCATCAACA CCATGCTGCG	360
CGTGTACCTG CCGAAGATCA CCAGCATGCT GAGCGACGTG ATGAAGCAGA ACTACGCCCT	420
GAGCCTGCAG ATCGAGTACC TGAGCAAGCA GCTGCAGGAG ATCAGCGACA AGCTGGACAT	480
CATCAACGTG AACGTCTGA TCAACAGCAC CCTGACCGAG ATCACCCCGG CCTACCAAGCG	540
CATCAAGTAC GTGAACGAGA AGTTCGAAGA GCTGACCTTC GCCACCGAGA CCAGCAGCAA	600
GGTGAAGAAG GACGGCAGCC CGGCCGACAT CCTGGACGAG CTGACCGAGC TGACCGAGCT	660
GGCCAAGAGC GTGACCAAGA ACGACGTGGA CGGCTTCGAG TTCTACCTGA ACACCTTCCA	720

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CGACGTGATG	GTGGGCAACA	ACCTGTTCGG	CCGCAGCGCC	CTGAAGACCG	CCAGCGAGCT	780
GATCACCAAG	GAGAACGTGA	AGACCAGCGG	CAGCGAGGTG	GGCAACGTGT	ACAACITCCT	840
GATCGTGCTG	ACCGCCCTGC	AGGCCAGGC	CTTCCTGACC	CTGACCACCT	GTCGCAAGCT	900
GCTGGGCCTG	GCCGACATCG	ACTACACCAG	CATCATGAAC	GAGCACTTGA	ACAAGGAGAA	960
GGAGGAGTTC	CGCGTGAACA	TCCTGCCGAC	CCTGAGCAAC	ACCTTCAGCA	ACCCGAACTA	1020
CGCCAAGGTG	AAGGGCAGCG	ACGAGGACGC	CAAGATGATC	GTGGAGGCTA	AGCCGGGCCA	1080
CGCGTTGATC	GGCTTCGAGA	TCAGCAACGA	CAGCATCACC	GTGCTGAAGG	TGTACGAGGC	1140
CAAGCTGAAG	CAGAACTACC	AGGTGGACAA	GGACAGCTTG	AGCGAGGTGA	TCTACGGCGA	1200
CATGGACAAG	CTGCTGTGTC	CGGACCAGAG	CGAGCAAATC	TACTACACCA	ACAACATCGT	1260
GTTCCCGAAC	GAGTACGTGA	TCACCAAGAT	CGACTTCACC	AAGAAGATGA	AGACCCCTGCG	1320
CTACGAGGTG	ACCGCCAATC	TCTACGACAG	CAGCACCGGC	GAGATCGACC	TGAACAAGAA	1380
GAAGGTGGAG	AGCAGCGAGG	CGAGTACCG	CACCTGAGC	GCGAACGACG	ACGGCGTCTA	1440
CATGCCACTG	GGCGTGATCA	GCGAGACCTT	CCTGACCCCCG	ATCAACGGCT	TTGGCCTGCA	1500
GGCCGACGAG	AACAGCCGCC	TGATCACCCCT	GACCTGTAAG	AGCTACCTGC	GCGAGCTGCT	1560
GCTAGCCACC	GACCTGAGCA	ACAAGGAGAC	CAAGCTGATC	GTGCCACCGA	GCGGCTTCAT	1620
CAGCAACATC	GTGGAGAACG	GCAGCATCGA	GGAGGACAAC	CTGGAGCCGT	GGAAGGCCAA	1680
CAACAAGAAC	GCCTACGTGG	ACCACACCGG	CGGCGTGAAC	GGCACCAAGG	CCCTGTACGT	1740
GCACAAGGAC	GGCGGCATCA	GCCAGTTCAT	CGGOGACAAG	CTGAAGCCGA	AGACCGAGTA	1800
CGTGATCCAG	TACACCGTGA	AGGGCAAGCC	ATCGATTAC	CTGAAGGACG	AGAACACCGG	1860
CTACATCCAC	TACGAGGACA	CCAACAACAA	CCTGGAGGAC	TACCAGACCA	TCAACAAGCG	1920
CTTCACCACC	GGCACCGACC	TGAAGGGCGT	GTACCTGATC	CTGAAGAGCC	AGAACGGCGA	1980
CGAGGCCTGG	GGCGACAACT	TCATCATCCT	GGAGATCAGC	CCGAGCGAGA	AGCTGCTGAG	2040
CCCGGAGCTG	ATCAACACCA	ACAACGGAC	CAGCACCGGC	AGCACCAACA	TCAGCGGCAA	2100
CACCCGTAC	GTGTACCGAG	GCGGCCGCGG	CATCCTGAAG	CAGAACCTGC	AGCTGGACAG	2160
CTTCAGCACC	TACCGCGTGT	ACTTCAGCGT	GAGCGCGAC	GCCAACGTGC	GCATCCGCAA	2220
CAGCCGCGAG	GTGCTGTTCG	AGAAGAGGTA	CATGAGCGGC	GCCAAGGACG	TGAGCGAGAT	2280
GTTCACCACC	AAGTCGAGA	AGGACAACCT	CTACATCGAG	CTGAGCCAGG	GCAACAAACCT	2340

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GTACGGCGGC CCGATCGTGC ACTTCTACGA CGTGAGGCATC AAGTTAACGT AGAGCTCAGA 2400
TCT 2403

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2612 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(ix) **FEATURE:**

- (A) NAME/KEY: CDS
(B) LOCATION: 118..2484
(D) OTHER INFORMATION: /note= "Native DNA sequence
encoding VIP3A(b) from AB424"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

ATTTGAAATTG ATAAAAAAGT ATGAGTGTTT AATAATCACT AATTACCAAT AAAGAATTAA 60

GAATACAAGT TTACAAGAAA TAAGTGTAC AAAAAATAGC TGAAAAGGAA GATGAAC 117

ATG AAC AAG AAT AAT ACT AAA TTA AGC ACA AGA GCC TTA CCA AGT TTT
 Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe
 790 795 800 805 165

ATT GAT TAT TTC AAT GGC ATT TAT GGA TTT GCC ACT GGT ATC AAA GAC
 Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp
 810 815 820

ATT ATG AAC ATG ATT TTT AAA ACG GAT ACA GGT GGT GAT CTA ACC CTA
 Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Thr Leu
 825 830 835

GAC GAA ATT TTA AAG AAT CAG CAG CTA CTA AAT GAT ATT TCT GGT AAA
Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn Asp Ile Ser Gly Lys
840 845 850

TTG GAT GGG GTG AAT GGA AGC TTA AAT GAT CTT ATC GCA CAG GGA AAC
 Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln Gly Asn
 855 860 865

TTA AAT ACA GAA TTA TCT AAG GAA ATA TTA AAA ATT GCA AAT GAA CAA 405
 Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn Glu Gln
 870 875 880 885

AAT CAA GTT TTA AAT GAT GTT AAT AAC AAA CTC GAT GCG ATA AAT ACG 453

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Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu Asp Ala Ile Asn Thr	890	895	900
ATG CTT CGG GTA TAT CTA CCT AAA ATT ACC TCT ATG TTG AGT GAT GTA			501
Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser Asp Val	905	910	915
ATG AAA CAA AAT TAT GCG CTA AGT CTG CAA ATA GAA TAC TTA AGT AAA			549
Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu Ser Lys	920	925	930
CAA TTG CAA GAG ATT TCT GAT AAG TTG GAT ATT ATT AAT GTA AAT GTA			597
Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val Asn Val	935	940	945
CTT ATT AAC TCT ACA CTT ACT GAA ATT ACA CCT GCG TAT CAA AGG ATT			645
Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile	950	955	960
AAA TAT GTG AAC GAA AAA TTT GAG GAA TTA ACT TTT GCT ACA GAA ACT			693
Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr Glu Thr	970	975	980
AGT TCA AAA GTA AAA AAG GAT GGC TCT CCT GCA GAT ATT CGT GAT GAG			741
Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala Asp Ile Arg Asp Glu	985	990	995
TTA ACT GAG TTA ACT GAA CTA GCG AAA AGT GTA ACA AAA AAT GAT GTG			789
Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn Asp Val	1000	1005	1010
GAT GGT TTT GAA TTT TAC CTT AAT ACA TTC CAC GAT GTA ATG GTA GGA			837
Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met Val Gly	1015	1020	1025
AAT AAT TTA TTC GGG CGT TCA GCT TTA AAA ACT GCA TCG GAA TTA ATT			885
Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu Leu Ile	1030	1035	1040
1045			
ACT AAA GAA AAT GTG AAA ACA AGT GGC AGT GAG GTC GGA AAT GTT TAT			933
Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu Val Gly Asn Val Tyr	1050	1055	1060
AAC TTC CTA ATT GTA TTA ACA GCT CTG CAA GCA AAA GCT TTT CTT ACT			981
Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala Lys Ala Phe Leu Thr	1065	1070	1075
TTA ACA CCA TGC CGA AAA TTA TTA GGC TTA GCA GAT ATT GAT TAT ACT			1029
Leu Thr Pro Cys Arg Lys Leu Leu Gly Leu Ala Asp Ile Asp Tyr Thr	1080	1085	1090
TCT ATT ATG AAT GAA CAT TTA AAT AAG GAA AAA GAG GAA TTT AGA GTA			1077
Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys Glu Glu Phe Arg Val	1095	1100	1105

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AAC ATC CTC CCT ACA CTT TCT AAT ACT TTT TCT AAT CCT AAT TAT GCA Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser Asn Pro Asn Tyr Ala 1110 1115 1120 1125	1125
AAA GTT AAA GGA AGT GAT GAA GAT GCA AAG ATG ATT GTG GAA GCT AAA Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met Ile Val Glu Ala Lys 1130 1135 1140	1173
CCA GGA CAT GCA TTG ATT GGG TTT GAA ATT AGT AAT GAT TCA ATT ACA Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser Asn Asp Ser Ile Thr 1145 1150 1155	1221
GTA TTA AAA GTA TAT GAG GCT AAG CTA AAA CAA AAT TAT CAA GTC GAT Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln Asn Tyr Gln Val Asp 1160 1165 1170	1269
AAG GAT TCC TTA TCG GAA GTT ATT TAT GGC GAT ATG GAT AAA TTA TTG Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp Met Asp Lys Leu Leu 1175 1180 1185	1317
TGC CCA GAT CAA TCT GGA CAA ATC TAT TAT ACA AAT AAC ATA GTA TTT Cys Pro Asp Gln Ser Gly Gln Ile Tyr Tyr Thr Asn Asn Ile Val Phe 1190 1195 1200 1205	1365
CCA AAT GAA TAT GTA ATT ACT AAA ATT GAT TTC ACT AAA AAA ATG AAA Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe Thr Lys Lys Met Lys 1210 1215 1220	1413
ACT TTA AGA TAT GAG GTA ACA GCG AAT TTT TAT GAT TCT TCT ACA GGA Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr Asp Ser Ser Thr Gly 1225 1230 1235	1461
GAA ATT GAC TTA AAT AAG AAA AAA GTA GAA TCA AGT GAA GCG GAG TAT Glu Ile Asp Leu Asn Lys Lys Val Glu Ser Ser Glu Ala Glu Tyr 1240 1245 1250	1509
AGA ACG TTA AGT GCT AAT GAT GAT GGG GTG TAT ATG CCG TTA GGT GTC Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr Met Pro Leu Gly Val 1255 1260 1265	1557
ATC AGT GAA ACA TTT TTG ACT CCG ATT AAT GGG TTT GGC CTC CAA GCT Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly Phe Gly Leu Gln Ala 1270 1275 1280 1285	1605
GAT GAA AAT TCA AGA TTA ATT ACT TTA ACA TGT AAA TCA TAT TTA AGA Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys Lys Ser Tyr Leu Arg 1290 1295 1300	1653
GAA CTA CTG CTA GCA ACA GAC TTA AGC AAT AAA GAA ACT AAA TTG ATC Glu Leu Leu Leu Ala Thr Asp Leu Ser Asn Lys Glu Thr Lys Leu Ile 1305 1310 1315	1701
GTC CCG CCA AGT GGT TTT ATT AGC AAT ATT GTA GAG AAC GGG TCC ATA Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val Glu Asn Gly Ser Ile 1320 1325 1330	1749

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GAA GAG GAC AAT TTA GAG CCG TGG AAA GCA AAT AAT AAG AAT GCG TAT Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn Asn Lys Asn Ala Tyr 1335 1340 1345	1797
GTA GAT CAT ACA GGC GGA GTG AAT GGA ACT AAA GCT TTA TAT GTT CAT Val Asp His Thr Gly Gly Val Asn Gly Thr Lys Ala Leu Tyr Val His 1350 1355 1360 1365	1845
AAG GAC GGA GGA ATT TCA CAA TTT ATT GGA GAT AAG TTA AAA CCG AAA Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp Lys Leu Lys Pro Lys 1370 1375 1380	1893
ACT GAG TAT GTA ATC CAA TAT ACT GTT AAA GGA AAA CCT TCT ATT CAT Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly Lys Pro Ser Ile His 1385 1390 1395	1941
TTA AAA GAT GAA AAT ACT GGA TAT ATT CAT TAT GAA GAT ACA AAT AAT Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr Glu Asp Thr Asn Asn 1400 1405 1410	1989
AAT TTA GAA GAT TAT CAA ACT ATT AAT AAA CGT TTT ACT ACA GGA ACT Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg Phe Thr Thr Gly Thr 1415 1420 1425	2037
GAT TTA AAG GGA GTG TAT TTA ATT TTA AAA AGT CAA AAT GGA GAT GAA Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser Gln Asn Gly Asp Glu 1430 1435 1440 1445	2085
GCT TGG GGA GAT AAC TTT ATT ATT TTG GAA ATT AGT CCT TCT GAA AAG Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile Ser Pro Ser Glu Lys 1450 1455 1460	2133
TTA TTA AGT CCA GAA TTA ATT AAT ACA AAT AAT TGG ACG AGT ACG GGA Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn Trp Thr Ser Thr Gly 1465 1470 1475	2181
TCA ACT AAT ATT AGC GGT AAT ACA CTC ACT CTT TAT CAG GGA GGA CGA Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu Tyr Gln Gly Gly Arg 1480 1485 1490	2229
GGG ATT CTA AAA CAA AAC CTT CAA TTA GAT AGT TTT TCA ACT TAT AGA Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser Phe Ser Thr Tyr Arg 1495 1500 1505	2277
GTG TAT TTC TCT GTG TCC GGA GAT GCT AAT GTA AGG ATT AGA AAT TCT Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val Arg Ile Arg Asn Ser 1510 1515 1520 1525	2325
AGG GAA GTG TTA TTT GAA AAA AGA TAT ATG AGC GGT GCT AAA GAT GTT Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser Gly Ala Lys Asp Val 1530 1535 1540	2373
TCT GAA ATG TTC ACT ACA AAA TTT GAG AAA GAT AAC TTC TAT ATA GAG Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp Asn Phe Tyr Ile Glu	2421

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1545	1550	1555	
GGG AAT AAT TTA TAT GGT GGT CCT ATT GTA CAT TTT TAC Gly Asn Asn Leu Tyr Gly Gly Pro Ile Val His Phe Tyr)			2469
	1565	1570	
ATT AAG TAAGATCGGG ATCTAATATT AACAGTTTT AGAAGCTAAT Ile Lys			2524
TGTCTTGAT TATGGAAAAA CACAATTTG TTTGCTAAGA TGTATATATA TAAAAGGCCAA TCAAGCTT			2584
			2612

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 789 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Met	Asn	Lys	Asn	Asn	Thr	Lys	Leu	Ser	Thr	Arg	Ala	Leu	Pro	Ser	Phe
1					5					10					15
Ile	Asp	Tyr	Phe	Asn	Gly	Ile	Tyr	Gly	Phe	Ala	Thr	Gly	Ile	Lys	Asp
						20				25					30
Ile	Met	Asn	Met	Ile	Phe	Lys	Thr	Asp	Thr	Gly	Gly	Asp	Leu	Thr	Leu
						35			40						45
Asp	Glu	Ile	Leu	Lys	Asn	Gln	Gln	Leu	Leu	Asn	Asp	Ile	Ser	Gly	Lys
						50			55						60
Leu	Asp	Gly	Val	Asn	Gly	Ser	Leu	Asn	Asp	Leu	Ile	Ala	Gln	Gly	Asn
						65			70						80
Leu	Asn	Thr	Glu	Leu	Ser	Lys	Glu	Ile	Leu	Lys	Ile	Ala	Asn	Glu	Gln
						85				90					95
Asn	Gln	Val	Leu	Asn	Asp	Val	Asn	Asn	Lys	Leu	Asp	Ala	Ile	Asn	Thr
						100				105					110
Met	Leu	Arg	Val	Tyr	Leu	Pro	Lys	Ile	Thr	Ser	Met	Leu	Ser	Asp	Val
						115				120					125
Met	Lys	Gln	Asn	Tyr	Ala	Leu	Ser	Leu	Gln	Ile	Glu	Tyr	Leu	Ser	Lys
						130			135						140
Gln	Leu	Gln	Glu	Ile	Ser	Asp	Lys	Leu	Asp	Ile	Ile	Asn	Val	Asn	Val
						145			150						160

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Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile
165 170 175

Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr Glu Thr
180 185 190

Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala Asp Ile Arg Asp Glu
195 200 205

Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn Asp Val
210 215 220

Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met Val Gly
225 230 235 240

Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu Leu Ile
245 250 255

Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu Val Gly Asn Val Tyr
260 265 270

Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala Lys Ala Phe Leu Thr
275 280 285

Leu Thr Pro Cys Arg Lys Leu Leu Gly Leu Ala Asp Ile Asp Tyr Thr
290 295 300

Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys Glu Glu Phe Arg Val
305 310 315 320

Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser Asn Pro Asn Tyr Ala
325 330 335

Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met Ile Val Glu Ala Lys
340 345 350

Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser Asn Asp Ser Ile Thr
355 360 365

Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln Asn Tyr Gln Val Asp
370 375 380

Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp Met Asp Lys Leu Leu
385 390 395 400

Cys Pro Asp Gln Ser Gly Gln Ile Tyr Tyr Thr Asn Asn Ile Val Phe
405 410 415

Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe Thr Lys Lys Met Lys
420 425 430

Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr Asp Ser Ser Thr Gly
435 440 445

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Glu Ile Asp Leu Asn Lys Lys Lys Val Glu Ser Ser Glu Ala Glu Tyr
450 455 460

Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr Met Pro Leu Gly Val
465 470 475 480

Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly Phe Gly Leu Gln Ala
485 490 495

Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys Lys Ser Tyr Leu Arg
500 505 510

Glu Leu Leu Leu Ala Thr Asp Leu Ser Asn Lys Glu Thr Lys Leu Ile
515 520 525

Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val Glu Asn Gly Ser Ile
530 535 540

Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn Asn Lys Asn Ala Tyr
545 550 555 560

Val Asp His Thr Gly Gly Val Asn Gly Thr Lys Ala Leu Tyr Val His
565 570 575

Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp Lys Leu Lys Pro Lys
580 585 590

Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly Lys Pro Ser Ile His
595 600 605

Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr Glu Asp Thr Asn Asn
610 615 620

Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg Phe Thr Thr Gly Thr
625 630 635 640

Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser Gln Asn Gly Asp Glu
645 650 655

Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile Ser Pro Ser Glu Lys
660 665 670

Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn Trp Thr Ser Thr Gly
675 680 685

Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu Tyr Gln Gly Gly Arg
690 695 700

Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser Phe Ser Thr Tyr Arg
705 710 715 720

Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val Arg Ile Arg Asn Ser
725 730 735

Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser Gly Ala Lys Asp Val

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740 745 750
Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp Asn Phe Tyr Ile Glu
755 760 765
Leu Ser Gln Gly Asn Asn Leu Tyr Gly Gly Pro Ile Val His Phe Tyr
770 775 780
Asp Val Ser Ile Lys
785

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "forward primer used to make
pCIB5526"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGATCCACCA TGAAGACCAA CCAGATCAGC

30

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "reverse primer used to make
pCIB5526"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

AAGCTTCAGC TCCTT

15

(2) INFORMATION FOR SEQ ID NO:35:

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2576 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 9..2564

(D) OTHER INFORMATION: /note= "Maize optimized sequence encoding VIP1A(a) with the Bacillus secretion signal removed as contained in pCIB5526"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

GATCCACC ATG AAG ACC AAC CAG ATC AGC ACC ACC CAG AAG AAC CAG CAG		50
Met Lys Thr Asn Gln Ile Ser Thr Thr Gln Lys Asn Gln Gln		
825	830	835
AAG GAG ATG GAC CGC AAG GGC CTG CTG GGC TAC TAC TTC AAG GGC AAG		98
Lys Glu Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr Phe Lys Gly Lys		
840	845	850
GAC TTC AGC AAC CTG ACC ATG TTC GCC CCC ACG CGT GAC AGC ACC CTG		146
Asp Phe Ser Asn Leu Thr Met Phe Ala Pro Thr Arg Asp Ser Thr Leu		
855	860	865
ATC TAC GAC CAG CAG ACC GCC AAC AAG CTG CTG GAC AAG AAG CAG CAG		194
Ile Tyr Asp Gln Gln Thr Ala Asn Lys Leu Leu Asp Lys Lys Gln Gln		
870	875	880
GAG TAC CAG AGC ATC CGC TGG ATC GGC CTG ATC CAG AGC AAG GAG ACC		242
Glu Tyr Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln Ser Lys Glu Thr		
885	890	895
GGC GAC TTC ACC TTC AAC CTG AGC GAG GAC GAG CAG GCC ATC ATC GAG		290
Gly Asp Phe Thr Phe Asn Leu Ser Glu Asp Glu Gln Ala Ile Ile Glu		
900	905	910
ATC AAC GGC AAG ATC ATC AGC AAC AAG GGC AAG GAG AAG CAG GTG GTG		338
Ile Asn Gly Lys Ile Ile Ser Asn Lys Gly Lys Glu Lys Gln Val Val		
920	925	930
CAC CTG GAG AAG GGC AAG CTG GTG CCC ATC AAG ATC GAG TAC CAG AGC		386
His Leu Glu Lys Gly Lys Leu Val Pro Ile Lys Ile Glu Tyr Gln Ser		
935	940	945
GAC ACC AAG TTC AAC ATC GAC AGC AAG ACC TTC AAG GAG CTG AAG CTT		434

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Asp Thr Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys Glu Leu Lys Leu			
950	955	960	
TTC AAG ATC GAC AGC CAG AAC CAG CCC CAG CAG GTG CAG CAG GAC GAG			482
Phe Lys Ile Asp Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu			
965	970	975	
CTG CGC AAC CCC GAG TTC AAC AAG AAG GAG AGC CAG GAG TTC CTG GCC			530
Leu Arg Asn Pro Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala			
980	985	990	995
AAG CCC AGC AAG ATC AAC CTG TTC ACC CAG CAG ATG AAG CGC GAG ATC			578
Lys Pro Ser Lys Ile Asn Leu Phe Thr Gln Gln Met Lys Arg Glu Ile			
1000	1005	1010	
GAC GAG GAC ACC GAC ACC GAC GGC GAC AGC ATC CCC GAC CTG TGG GAG			626
Asp Glu Asp Thr Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu			
1015	1020	1025	
GAG AAC GGC TAC ACC ATC CAG AAC CGC ATC GCC GTG AAG TGG GAC GAC			674
Glu Asn Gly Tyr Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp			
1030	1035	1040	
AGC CTG GCT AGC AAG GGC TAC ACC AAG TTC GTG AGC AAC CCC CTG GAG			722
Ser Leu Ala Ser Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu			
1045	1050	1055	
AGC CAC ACC GTG GGC GAC CCC TAC ACC GAC TAC GAG AAG GCC GCC CGC			770
Ser His Thr Val Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg			
1060	1065	1070	1075
GAC CTG GAC CTG AGC AAC GCC AAG GAG ACC TTC AAC CCC CTG GTG GCC			818
Asp Leu Asp Leu Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala			
1080	1085	1090	
GCC TTC CCC AGC GTG AAC GTG AGC ATG GAG AAG GTG ATC CTG AGC CCC			866
Ala Phe Pro Ser Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro			
1095	1100	1105	
AAC GAG AAC CTG AGC AAC AGC GTG GAG AGC CAC TCG AGC ACC AAC TGG			914
Asn Glu Asn Leu Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp			
1110	1115	1120	
AGC TAC ACC AAC ACC GAG GGC GCC AGC GTG GAG GCC GGC ATC GGT CCC			962
Ser Tyr Thr Asn Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro			
1125	1130	1135	
AAG GGC ATC AGC TTC GGC GTG AGC GTG AAC TAC CAG CAC AGC GAG ACC			1010
Lys Gly Ile Ser Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr			
1140	1145	1150	1155
GTG GCC CAG GAG TGG GGC ACC AGC ACC GGC AAC ACC AGC CAG TTC AAC			1058
Val Ala Gln Glu Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn			
1160	1165	1170	

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ACC GCC AGC GCC GGC TAC CTG AAC GCC AAC GTG CGC TAC AAC AAC GTG Thr Ala Ser Ala Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val 1175 1180 1185	1106
GGC ACC GGC GCC ATC TAC GAC GTG AAG CCC ACC ACC AGC TTC GTG CTG Gly Thr Gly Ala Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu 1190 1195 1200	1154
AAC AAC GAC ACC ATC GCC ACC ATC ACC GCC AAG TCG AAT TCC ACC GCC Asn Asn Asp Thr Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala 1205 1210 1215	1202
CTG AAC ATC AGC CCC GGC GAG AGC TAC CCC AAG AAG GGC CAG AAC GGC Leu Asn Ile Ser Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly 1220 1225 1230 1235	1250
ATC GCC ATC ACC AGC ATG GAC GAC TTC AAC AGC CAC CCC ATC ACC CTG Ile Ala Ile Thr Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu 1240 1245 1250	1298
AAC AAG AAG CAG GTG GAC AAC CTG CTG AAC AAC AAG CCC ATG ATG CTG Asn Lys Lys Gln Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu 1255 1260 1265	1346
GAG ACC AAC CAG ACC GAC GGC GTC TAC AAG ATC AAG GAC ACC CAC GGC Glu Thr Asn Gln Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly 1270 1275 1280	1394
AAC ATC GTG ACG GGC GGC GAG TGG AAC GGC GTG ATC CAG CAG ATC AAG Asn Ile Val Thr Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys 1285 1290 1295	1442
GCC AAG ACC GCC AGC ATC ATC GTC GAC GAC GGC GAG CGC GTG GCC GAG Ala Lys Thr Ala Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu 1300 1305 1310 1315	1490
AAG CGC GTG GCC GCC AAG GAC TAC GAG AAC CCC GAG GAC AAG ACC CCC Lys Arg Val Ala Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro 1320 1325 1330	1538
AGC CTG ACC CTG AAG GAC GCC CTG AAG CTG AGC TAC CCC GAC GAG ATC Ser Leu Thr Leu Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile 1335 1340 1345	1586
AAG GAG ATC GAG GGC TTG CTG TAC TAC AAG AAC AAG CCC ATC TAC GAG Lys Glu Ile Glu Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu 1350 1355 1360	1634
AGC AGC GTG ATG ACC TAT CTA GAC GAG AAC ACC GCC AAG GAG GTG ACC Ser Ser Val Met Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr 1365 1370 1375	1682
AAG CAG CTG AAC GAC ACC ACC GGC AAG TTC AAG GAC GTG AGC CAC CTG Lys Gln Leu Asn Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu 1380 1385 1390 1395	1730

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TAC GAC GTG AAG CTG ACC CCC AAG ATG AAC GTG ACC ATC AAG CTG AGC Tyr Asp Val Lys Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser 1400 1405 1410	1778
ATC CTG TAC GAC AAC GCC GAG AGC AAC GAC AAC AGC ATC GGC AAG TGG Ile Leu Tyr Asp Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp 1415 1420 1425	1826
ACC AAC ACC AAC ATC GTG AGC GGC GGC AAC AAC GGC AAG AAG CAG TAC Thr Asn Thr Asn Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr 1430 1435 1440	1874
AGC AGC AAC AAC CCC GAC GCC AAC CTG ACC CTG AAC ACC GAC GCC CAG Ser Ser Asn Asn Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln 1445 1450 1455	1922
GAG AAG CTG AAC AAG AAC CGC GAC TAC TAC ATC AGC CTG TAC ATG AAG Glu Lys Leu Asn Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys 1460 1465 1470 1475	1970
AGC GAG AAC ACC CAG TGC GAG ATC ACC ATC GAC GGC GAG ATA TAC Ser Glu Lys Asn Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr 1480 1485 1490	2018
CCC ATC ACC ACC AAG ACC GTG AAC AAC GTG AAC AAG GAC AAC TAC AAG CGC Pro Ile Thr Thr Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg 1495 1500 1505	2066
CTG GAC ATC ATC GCC CAC AAC ATC AAG AGC AAC CCC ATC AGC AGC CTG Leu Asp Ile Ile Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu 1510 1515 1520	2114
CAC ATC AAG ACC AAC GAC GAG ATC ACC CTG TTC TGG GAC GAC ATA TCG His Ile Lys Thr Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser 1525 1530 1535	2162
ATT ACC GAC GTC GCC AGC ATC AAG CCC GAG AAC CTG ACC GAC AGC GAG Ile Thr Asp Val Ala Ser Ile Lys Pro Glu Asn Leu Thr Asp Ser Glu 1540 1545 1550 1555	2210
ATC AAG CAG ATA TAC AGT CGC TAC GGC ATC AAG CTG GAG GAC GGC ATC Ile Lys Gln Ile Tyr Ser Arg Tyr Gly Ile Lys Leu Glu Asp Gly Ile 1560 1565 1570	2258
CTG ATC GAC AAG AAA GGC GGC ATC CAC TAC GGC GAG TTC ATC AAC GAG Leu Ile Asp Lys Lys Gly Gly Ile His Tyr Gly Glu Phe Ile Asn Glu 1575 1580 1585	2306
GCC AGC TTC AAC ATC GAG CCC CTG CAG AAC TAC GTG ACC AAG TAC GAG Ala Ser Phe Asn Ile Glu Pro Leu Gln Asn Tyr Val Thr Lys Tyr Glu 1590 1595 1600	2354
GTG ACC TAC AGC AGC GAG CTG GGC CCC AAC GTG AGC GAC ACC CTG GAG Val Thr Tyr Ser Ser Glu Leu Gly Pro Asn Val Ser Asp Thr Leu Glu	2402

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1605	1610	1615	
AGC GAC AAG ATT TAC AAG GAC GGC ACC ATC AAG TTC GAC TTC ACC AAG Ser Asp Lys Ile Tyr Lys Asp Gly Thr Ile Lys Phe Asp Phe Thr Lys			2450
1620	1625	1630	1635
TAC AGC AAG AAC GAG CAG GGC CTG TTC TAC GAC AGC GGC CTG AAC TGG Tyr Ser Lys Asn Glu Gln Gly Leu Phe Tyr Asp Ser Gly Leu Asn Trp			2498
1640	1645	1650	
GAC TTC AAG ATC AAC GCC ATC ACC TAC GAC GGC AAG GAG ATG AAC GTG Asp Phe Lys Ile Asn Ala Ile Thr Tyr Asp Gly Lys Glu Met Asn Val			2546
1655	1660	1665	
TTC CAC CGC TAC AAC AAG TAGATCTGAG CT Phe His Arg Tyr Asn Lys			2576
1670			

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 852 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Met	Lys	Thr	Asn	Gln	Ile	Ser	Thr	Thr	Gln	Lys	Asn	Gln	Gln	Lys	Glu
1															15
Met	Asp	Arg	Lys	Gly	Leu	Leu	Gly	Tyr	Tyr	Phe	Lys	Gly	Lys	Asp	Phe
															30
Ser	Asn	Leu	Thr	Met	Phe	Ala	Pro	Thr	Arg	Asp	Ser	Thr	Leu	Ile	Tyr
															45
Asp	Gln	Gln	Thr	Ala	Asn	Lys	Leu	Leu	Asp	Lys	Lys	Gln	Gln	Glu	Tyr
															50
Gln	Ser	Ile	Arg	Trp	Ile	Gly	Leu	Ile	Gln	Ser	Lys	Glu	Thr	Gly	Asp
															55
Phe	Thr	Phe	Asn	Leu	Ser	Glu	Asp	Glu	Gln	Ala	Ile	Ile	Glu	Ile	Asn
															60
Gly	Lys	Ile	Ile	Ser	Asn	Lys	Gly	Lys	Glu	Lys	Gln	Val	Val	His	Leu
															65
Glu	Lys	Gly	Lys	Leu	Val	Pro	Ile	Lys	Ile	Glu	Tyr	Gln	Ser	Asp	Thr
															70
Lys	Phe	Asn	Ile	Asp	Ser	Lys	Thr	Phe	Lys	Glu	Leu	Lys	Leu	Phe	Lys
															75
															80
															85
															90
															95
															100
															105
															110
															115
															120
															125

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130	135	140
Ile Asp Ser Gln Asn Gln Pro Gln Gln Val Gln Gln Asp Glu Leu Arg		
145	150	155
160		
Asn Pro Glu Phe Asn Lys Lys Glu Ser Gln Glu Phe Leu Ala Lys Pro		
165	170	175
Ser Lys Ile Asn Leu Phe Thr Gln Gln Met Lys Arg Glu Ile Asp Glu		
180	185	190
Asp Thr Asp Thr Asp Gly Asp Ser Ile Pro Asp Leu Trp Glu Glu Asn		
195	200	205
Gly Tyr Thr Ile Gln Asn Arg Ile Ala Val Lys Trp Asp Asp Ser Leu		
210	215	220
Ala Ser Lys Gly Tyr Thr Lys Phe Val Ser Asn Pro Leu Glu Ser His		
225	230	235
240		
Thr Val Gly Asp Pro Tyr Thr Asp Tyr Glu Lys Ala Ala Arg Asp Leu		
245	250	255
Asp Leu Ser Asn Ala Lys Glu Thr Phe Asn Pro Leu Val Ala Ala Phe		
260	265	270
Pro Ser Val Asn Val Ser Met Glu Lys Val Ile Leu Ser Pro Asn Glu		
275	280	285
Asn Leu Ser Asn Ser Val Glu Ser His Ser Ser Thr Asn Trp Ser Tyr		
290	295	300
320		
Thr Asn Thr Glu Gly Ala Ser Val Glu Ala Gly Ile Gly Pro Lys Gly		
305	310	315
320		
Ile Ser Phe Gly Val Ser Val Asn Tyr Gln His Ser Glu Thr Val Ala		
325	330	335
Gln Glu Trp Gly Thr Ser Thr Gly Asn Thr Ser Gln Phe Asn Thr Ala		
340	345	350
Ser Ala Gly Tyr Leu Asn Ala Asn Val Arg Tyr Asn Asn Val Gly Thr		
355	360	365
Gly Ala Ile Tyr Asp Val Lys Pro Thr Thr Ser Phe Val Leu Asn Asn		
370	375	380
Asp Thr Ile Ala Thr Ile Thr Ala Lys Ser Asn Ser Thr Ala Leu Asn		
385	390	395
400		
Ile Ser Pro Gly Glu Ser Tyr Pro Lys Lys Gly Gln Asn Gly Ile Ala		
405	410	415
Ile Thr Ser Met Asp Asp Phe Asn Ser His Pro Ile Thr Leu Asn Lys		
420	425	430

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Lys Gln Val Asp Asn Leu Leu Asn Asn Lys Pro Met Met Leu Glu Thr
435 440 445

Asn Gln Thr Asp Gly Val Tyr Lys Ile Lys Asp Thr His Gly Asn Ile
450 455 460

Val Thr Gly Gly Glu Trp Asn Gly Val Ile Gln Gln Ile Lys Ala Lys
465 470 475 480

Thr Ala Ser Ile Ile Val Asp Asp Gly Glu Arg Val Ala Glu Lys Arg
485 490 495

Val Ala Ala Lys Asp Tyr Glu Asn Pro Glu Asp Lys Thr Pro Ser Leu
500 505 510

Thr Leu Lys Asp Ala Leu Lys Leu Ser Tyr Pro Asp Glu Ile Lys Glu
515 520 525

Ile Glu Gly Leu Leu Tyr Tyr Lys Asn Lys Pro Ile Tyr Glu Ser Ser
530 535 540

Val Met Thr Tyr Leu Asp Glu Asn Thr Ala Lys Glu Val Thr Lys Gln
545 550 555 560

Leu Asn Asp Thr Thr Gly Lys Phe Lys Asp Val Ser His Leu Tyr Asp
565 570 575

Val Lys Leu Thr Pro Lys Met Asn Val Thr Ile Lys Leu Ser Ile Leu
580 585 590

Tyr Asp Asn Ala Glu Ser Asn Asp Asn Ser Ile Gly Lys Trp Thr Asn
595 600 605

Thr Asn Ile Val Ser Gly Gly Asn Asn Gly Lys Lys Gln Tyr Ser Ser
610 615 620

Asn Asn Pro Asp Ala Asn Leu Thr Leu Asn Thr Asp Ala Gln Glu Lys
625 630 635 640

Leu Asn Lys Asn Arg Asp Tyr Tyr Ile Ser Leu Tyr Met Lys Ser Glu
645 650 655

Lys Asn Thr Gln Cys Glu Ile Thr Ile Asp Gly Glu Ile Tyr Pro Ile
660 665 670

Thr Thr Lys Thr Val Asn Val Asn Lys Asp Asn Tyr Lys Arg Leu Asp
675 680 685

Ile Ile Ala His Asn Ile Lys Ser Asn Pro Ile Ser Ser Leu His Ile
690 695 700

Lys Thr Asn Asp Glu Ile Thr Leu Phe Trp Asp Asp Ile Ser Ile Thr
705 710 715 720

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Asp	Val	Ala	Ser	Ile	Lys	Pro	Glu	Asn	Leu	Thr	Asp	Ser	Glu	Ile	Lys
				725					730					735	
Gln	Ile	Tyr	Ser	Arg	Tyr	Gly	Ile	Lys	Leu	Glu	Asp	Gly	Ile	Leu	Ile
				740				745					750		
Asp	Lys	Lys	Gly	Gly	Ile	His	Tyr	Gly	Glu	Phe	Ile	Asn	Glu	Ala	Ser
					755			760				765			
Phe	Asn	Ile	Glu	Pro	Leu	Gln	Asn	Tyr	Val	Thr	Lys	Tyr	Glu	Val	Thr
					770			775			780				
Tyr	Ser	Ser	Glu	Leu	Gly	Pro	Asn	Val	Ser	Asp	Thr	Leu	Glu	Ser	Asp
					785			790			795			800	
Lys	Ile	Tyr	Lys	Asp	Gly	Thr	Ile	Lys	Phe	Asp	Phe	Thr	Lys	Tyr	Ser
					805			810					815		
Lys	Asn	Glu	Gln	Gly	Leu	Phe	Tyr	Asp	Ser	Gly	Leu	Asn	Trp	Asp	Phe
					820			825			830				
Lys	Ile	Asn	Ala	Ile	Thr	Tyr	Asp	Gly	Lys	Glu	Met	Asn	Val	Phe	His
					835			840			845				
Arg	Tyr	Asn	Lys												
			850												

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "forward primer used to make pCIB5527"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GGATCCACCA TGCTGCAGAA CCTGAAGATC AC

32

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "reverse primer used to make pCIB5527"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

AAGCTTCCAC TCCTTCTC

18

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1241 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 9..1238

(D) OTHER INFORMATION: /note= "Maize optimized DNA sequence encoding VIP2A(a) with the Bacillus secretion signal removed as contained in pCIB5527"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

GATCCACC ATG CTG CAG AAC CTG AAG ATC ACC GAC AAG GTG GAG GAC TTC
 Met Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu Asp Phe
 855 860 865

50

AAG GAG GAC AAG GAG AAG GCC AAG GAG TGG GGC AAG GAG AAG GAG AAG
 Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys Glu Lys
 870 875 880

98

GAG TGG AAG CTT ACC GCC ACC GAG AAG GGC AAG ATG AAC AAC TTC CTG
 Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn Phe Leu
 885 890 895

146

GAC AAC AAG AAC GAC ATC AAG ACC AAC TAC AAG GAG ATC ACC TTC AGC
 Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr Phe Ser
 900 905 910

194

ATA GCC GGC AGC TTC GAG GAC GAG ATC AAG GAC CTG AAG GAG ATC GAC

242

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Ile Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu Ile Asp		915		
	920		'925	930
AAG ATG TTC GAC AAG ACC AAC CTG AGC AAC AGC ATC ATC ACC TAC AAG				290
Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr Tyr Lys		935	940	945
AAC GTG GAG CCC ACC ACC ATC GGC TTC AAC AAG AGC CTG ACC GAG GGC				338
Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr Glu Gly		950	955	960
AAC ACC ATC AAC AGC GAC GCC ATG GCC CAG TTC AAG GAG CAG TTC CTG				386
Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln Phe Leu		965	970	975
GAC CGC GAC ATC AAG TTC GAC AGC TAC CTG GAC ACC CAC CTG ACC GCC				434
Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu Thr Ala		980	985	990
CAG CAG GTG AGC AGC AAG GAG CGC GTG ATC CTG AAG GTG ACC GTC CCC				482
Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr Val Pro		995	1000	1005
AGC GGC AAG GGC AGC ACC ACC CCC ACC AAG GCC GGC GTG ATC CTG AAC				530
Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile Leu Asn		1015	1020	1025
AAC AGC GAG TAC AAG ATG CTG ATC GAC AAC GGC TAC ATG GTG CAC GTG				578
Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val His Val		1030	1035	1040
GAC AAG GTG AGC AAG GTG GTG AAG AAG GGC GTG GAG TGC CTC CAG ATC				626
Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu Gln Ile		1045	1050	1055
GAG GGC ACC CTG AAG AAG AGT CTA GAC TTC AAG AAC GAC ATC AAC GCC				674
Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile Asn Ala		1060	1065	1070
GAG GCC CAC AGC TGG GGC ATG AAG AAC TAC GAG GAG TGG GCC AAG GAC				722
Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala Lys Asp		1075	1080	1085
CTG ACC GAC AGC CAG CGC GAG GCC CTG GAC GGC TAC GCC CGC CAG GAC				770
Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg Gln Asp		1095	1100	1105
TAC AAG GAG ATC AAC AAC TAC CTG CGC AAC CAG GGC GGC AGC GGC AAC				818
Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Ser Gly Asn		1110	1115	1120
GAG AAG CTG GAC GCC CAG AAG AAC ATC AGC GAC GCC CTG GGC AAG				866
Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu Gly Lys		1125	1130	1135

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AAG CCC ATC CCC GAG AAC ATC ACC GTG TAC CGC TGG TGC GGC ATG CCC Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly Met Pro 1140 1145 1150	914
GAG TTC GGC TAC CAG ATC AGC GAC CCC CTG CCC AGC CTG AAG GAC TTC Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys Asp Phe 1155 1160 1165 1170	962
GAG GAG CAG TTC CTG AAC ACC ATC AAG GAG GAC AAG GGC TAC ATG AGC Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr Met Ser 1175 1180 1185	1010
ACC AGC CTG AGC AGC GAG CGC CTG GCC GCC TTC GGC AGC CGC AAG ATC Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg Lys Ile 1190 1195 1200	1058
ATC CTG CGC CTG CAG GTG CCC AAG GGC AGC ACT GGT GCC TAC CTG AGC Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr Leu Ser 1205 1210 1215	1106
GCC ATC GGC GGC TTC GCC AGC GAG AAG GAG ATC CTG CTG GAT AAG GAC Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp Lys Asp 1220 1225 1230	1154
AGC AAG TAC CAC ATC GAC AAG GTG ACC GAG GTG ATC ATC AAG GGC GTG Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys Gly Val 1235 1240 1245 1250	1202
AAG CGC TAC GTG GTG GAC GCC ACC CTG CTG ACC AAC TAG Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn 1255 1260	1241

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 410 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Met Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu Asp Phe Lys Glu 1 5 10 15
Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys Glu Lys Glu Trp 20 25 30
Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn Phe Leu Asp Asn 35 40 45
Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr Phe Ser Ile Ala 50 55 60

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Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu Ile Asp Lys Met
65 70 75 80

Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr Tyr Lys Asn Val
85 90 95

Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr Glu Gly Asn Thr
100 105 110

Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln Phe Leu Asp Arg
115 120 125

Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu Thr Ala Gln Gln
130 135 140

Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr Val Pro Ser Gly
145 150 155 160

Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile Leu Asn Asn Ser
165 170 175

Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val His Val Asp Lys
180 185 190

Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu Gln Ile Glu Gly
195 200 205

Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile Asn Ala Glu Ala
210 215 220

His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala Lys Asp Leu Thr
225 230 235 240

Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg Gln Asp Tyr Lys
245 250 255

Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Ser Gly Asn Glu Lys
260 265 270

Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu Gly Lys Lys Pro
275 280 285

Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly Met Pro Glu Phe
290 295 300

Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys Asp Phe Glu Glu
305 310 315 320

Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr Met Ser Thr Ser
325 330 335

Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg Lys Ile Ile Leu
340 345 350

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Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr Leu Ser Ala Ile
355 360 365

Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp Lys Asp Ser Lys
370 375 380

Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys Gly Val Lys Arg
385 390 395 400

Tyr Val Val Asp Ala Thr Leu Leu Thr Asn
405 410

(2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 72 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "oligonucleotide encoding eukaryotic secretion signal used to construct pCIB5527"

- (iii) HYPOTHETICAL: NO

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

GGATCCACCA TGGCTGAG CTGGATCTTC CTGTTCCTGC TGAGCGGCGC CGCGGGCGTG 60

CACTGGCTGC AG 72

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1241 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "Synthetic DNA"

- (iii) HYPOTHETICAL: NO

- (ix) FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 9..1238

(D) OTHER INFORMATION: /note= "Maize optimized DNA sequence encoding VIP2A(a) with the *Bacillus* secretion signal removed and the eukaryotic secretion signal inserted as

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contained in pCIB5528"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

GATCCACC ATG CTG CAG AAC CTG AAG ATC ACC GAC AAG GTG GAG GAC TTC Met Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu Asp Phe 415	420	50
AAG GAG GAC AAG GAG AAG GCC AAG GAG TGG GGC AAG GAG AAG GAG AAG Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys Glu Lys 425	430	440
GAG TGG AAG CTT ACC GCC ACC GAG AAG GGC AAG ATG AAC AAC TTC CTG Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn Phe Leu 445	450	455
GAC AAC AAG AAC GAC ATC AAG ACC AAC TAC AAG GAG ATC ACC ACC TTC AGC Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr Phe Ser 460	465	470
ATA GCC GGC AGC TTC GAG GAC GAG ATC AAG GAC CTG AAG GAG ATC GAC Ile Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu Ile Asp 475	480	485
AAG ATG TTC GAC AAG ACC AAC CTG AGC AAC AGC ATC ATC ACC TAC AAG Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr Tyr Lys 490	495	500
AAC GTG GAG CCC ACC ACC ATC GGC TTC AAC AAG AGC CTG ACC GAG GGC Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr Glu Gly 505	510	520
AAC ACC ATC AAC AGC GAC GCC ATG GCC CAG TTC AAG GAG CAG TTC CTG Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln Phe Leu 525	530	535
GAC CGC GAC ATC AAG TTC GAC AGC TAC CTG GAC ACC CAC CTG ACC GCC Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu Thr Ala 540	545	550
CAG CAG GTG AGC AGC AAG GAG CGC GTG ATC CTG AAG GTG ACC GTC CCC Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr Val Pro 555	560	565
AGC GGC AAG GGC AGC ACC ACC CCC ACC AAG GCC GGC GTG ATC CTG AAC Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile Leu Asn 570	575	580
AAC AGC GAG TAC AAG ATG CTG ATC GAC AAC GGC TAC ATG GTG CAC GTG Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val His Val 585	590	595
GAC AAG GTG AGC AAG GTG GTG AAG AAG GGC GTG GAG TGC CTC CAG ATC Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu Gln Ile 600	600	626

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605	610	615	
GAG GGC ACC CTG AAG AAG AGT CTA GAC TTC AAG AAC GAC ATC AAC GCC Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile Asn Ala 620 625 630 674			
GAG GCC CAC AGC TGG GGC ATG AAG AAC TAC GAG GAG TGG GCC AAG GAC Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala Lys Asp 635 640 645 722			
CTG ACC GAC AGC CAG CGC GAG GCC CTG GAC GGC TAC GCC CGC CAG GAC Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg Gln Asp 650 655 660 770			
TAC AAG GAG ATC AAC AAC TAC CTG CGC AAC CAG GGC GGC AGC GGC AAC Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser Gly Asn 665 670 675 680 818			
GAG AAG CTG GAC GCC CAG ATC AAG AAC ATC AGC GAC GCC CTG GGC AAG Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu Gly Lys 685 690 695 866			
AAG CCC ATC CCC GAG AAC ATC ACC GTG TAC CGC TGG TGC GGC ATG CCC Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly Met Pro 700 705 710 914			
GAG TTC GGC TAC CAG ATC AGC GAC CCC CTG CCC AGC CTG AAG GAC TTC Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys Asp Phe 715 720 725 962			
GAG GAG CAG TTC CTG AAC ACC ATC AAG GAG GAC AAG GGC TAC ATG AGC Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr Met Ser 730 735 740 1010			
ACC AGC CTG AGC AGC GAG CGC CTG GCC TTC GGC AGC CGC AAG ATC Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg Lys Ile 745 750 755 760 1058			
ATC CTG CGC CTG CAG GTG CCC AAG GGC AGC ACT GGT GCC TAC CTG AGC Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr Leu Ser 765 770 775 1106			
GCC ATC GGC GGC TTC GCC AGC GAG AAG GAG ATC CTG CTG GAT AAG GAC Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp Lys Asp 780 785 790 1154			
AGC AAG TAC CAC ATC GAC AAG GTG ACC GAG GTG ATC ATC AAG GGC GTG Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys Gly Val 795 800 805 1202			
AAG CGC TAC GTG GTG GAC GCC ACC CTG CTG ACC AAC TAG Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn 810 815 820 1241			

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(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 410 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Met Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu Asp Phe Lys Glu
1 5 10 15

Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys Glu Lys Glu Trp
20 25 30

Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn Phe Leu Asp Asn
35 40 45

Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr Phe Ser Ile Ala
50 55 60

Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu Ile Asp Lys Met
65 70 75 80

Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr Tyr Lys Asn Val
85 90 95

Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr Glu Gly Asn Thr
100 105 110

Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln Phe Leu Asp Arg
115 120 125

Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu Thr Ala Gln Gln
130 135 140

Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr Val Pro Ser Gly
145 150 155 160

Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile Leu Asn Asn Ser
165 170 175

Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val His Val Asp Lys
180 185 190

Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu Gln Ile Glu Gly
195 200 205

Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile Asn Ala Glu Ala
210 215 220

His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala Lys Asp Leu Thr
225 230 235 240

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Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg Gln Asp Tyr Lys			
245	250	255	
Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser Gly Asn Glu Lys			
260	265	270	
Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu Gly Lys Lys Pro			
275	280	285	
Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly Met Pro Glu Phe			
290	295	300	
Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys Asp Phe Glu Glu			
305	310	315	320
Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr Met Ser Thr Ser			
325	330	335	
Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg Lys Ile Ile Leu			
340	345	350	
Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr Leu Ser Ala Ile			
355	360	365	
Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp Lys Asp Ser Lys			
370	375	380	
Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys Gly Val Lys Arg			
385	390	395	400
Tyr Val Val Asp Ala Thr Leu Leu Thr Asn			
405	410		

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 86 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "oligonucleotide encoding
vacuolar targetting peptide used to construct pCIB5533"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

CCGCAGGGCGT GCACTGCCTC AGCAGCAGCA GCTTCGCCGA CAGCAACCCC ATCCGCGTGA

60

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CCGACCGCGC CGCCAGCACC CTGCAG

86

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1358 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 9..1355

(D) OTHER INFORMATION: /note= "Maize optimized VIP2A(a)
with the Bacillus secretion signal removed and the vacuolar
targetting signal inserted as contained in pCIB5533"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

GATCCACC ATG GGC TGG AGC TGG ATC TTC CTG TTC CTG CTG AGC GGC GCC	50
Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Ala	
415	420

GCG GGC GTG CAC TGC CTC AGC AGC AGC TTC GCC GAC AGC AAC CCC	98
Ala Gly Val His Cys Leu Ser Ser Ser Phe Ala Asp Ser Asn Pro	
425	430
435	440

ATC CGC GTG ACC GAC CGC GCC AGC ACC CTG CAG AAC CTG AAG ATC	146
Ile Arg Val Thr Asp Arg Ala Ala Ser Thr Leu Gln Asn Leu Lys Ile	
445	450
455	

ACC GAC AAG GTG GAG GAC TTC AAG GAG GAC AAG GAG AAG GCC AAG GAG	194
Thr Asp Lys Val Glu Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu	
460	465
470	

TGG GGC AAG GAG AAG GAG TGG AAG CTT ACC GCC ACC GAG AAG	242
Trp Gly Lys Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys	
475	480
485	

GGC AAG ATG AAC AAC TTC CTG GAC AAC AAG AAC GAC ATC AAG ACC AAC	290
Gly Lys Met Asn Asn Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn	
490	495
500	

TAC AAG GAG ATC ACC TTC AGC ATA GCC GGC AGC TTC GAG GAC GAG ATC	338
Tyr Lys Glu Ile Thr Phe Ser Ile Ala Gly Ser Phe Glu Asp Glu Ile	
505	510
515	520

AAG GAC CTG AAG GAG ATC GAC AAG ATG TTC GAC AAG ACC AAC CTG AGC	386
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Lys Asp Leu Lys Glu Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser			
525	530	535	
AAC AGC ATC ATC ACC TAC AAG AAC GTG GAG CCC ACC ACC ATC GGC TTC			434
Asn Ser Ile Ile Thr Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe			
540	545	550	
AAC AAG AGC CTG ACC GAG GGC AAC ACC ATC AAC AGC GAC GCC ATG GCC			482
Asn Lys Ser Leu Thr Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala			
555	560	565	
CAG TTC AAG GAG CAG TTC CTG GAC CGC GAC ATC AAG TTC GAC AGC TAC			530
Gln Phe Lys Glu Gln Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr			
570	575	580	
CTG GAC ACC CAC CTG ACC GCC CAG CAG GTG AGC AGC AAG GAG CGC GTG			578
Leu Asp Thr His Leu Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val			
585	590	595	600
ATC CTG AAG GTG ACC GTC CCC AGC GGC AAG GGC AGC ACC ACC CCC ACC			626
Ile Leu Lys Val Thr Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr			
605	610	615	
AAG GCC GGC GTG ATC CTG AAC AAC AGC GAG TAC AAG ATG CTG ATC GAC			674
Lys Ala Gly Val Ile Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp			
620	625	630	
AAC GGC TAC ATG GTG CAC GTG GAC AAG GTG AGC AAG GTG GTG AAG AAG			722
Asn Gly Tyr Met Val His Val Asp Lys Val Ser Lys Val Val Lys Lys			
635	640	645	
GGC GTG GAG TGC CTC CAG ATC GAG GGC ACC CTG AAG AAG AGT CTA GAC			770
Gly Val Glu Cys Leu Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp			
650	655	660	
TTC AAG AAC GAC ATC AAC GCC GAG GCC CAC AGC TGG GGC ATG AAG AAC			818
Phe Lys Asn Asp Ile Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn			
665	670	675	680
TAC GAG GAG TGG GCC AAG GAC CTG ACC GAC AGC CAG CGC GAG GCC CTG			866
Tyr Glu Glu Trp Ala Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu			
685	690	695	
GAC GGC TAC GCC CGC CAG GAC TAC AAG GAG ATC AAC AAC TAC CTG CGC			914
Asp Gly Tyr Ala Arg Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg			
700	705	710	
AAC CAG GGC GGC AGC GGC AAC GAG AAG CTG GAC GCC CAG ATC AAG AAC			962
Asn Gln Gly Gly Ser Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn			
715	720	725	
ATC AGC GAC GCC CTG GGC AAG AAG CCC ATC CCC GAG AAC ATC ACC GTG			1010
Ile Ser Asp Ala Leu Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val			
730	735	740	

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TAC CGC TGG TGC GGC ATG CCC GAG TTC GGC TAC CAG ATC AGC GAC CCC Tyr Arg Trp Cys Gly Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro 745 750 755 760	1058
CTG CCC AGC CTG AAG GAC TTC GAG GAG CAG TTC CTG AAC ACC ATC AAG Leu Pro Ser Leu Lys Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys 765 770 775	1106
GAG GAC AAG GGC TAC ATG AGC ACC AGC CTG AGC AGC GAG CGC CTG GCC Glu Asp Lys Gly Tyr Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala 780 785 790	1154
GCC TTC GGC AGC CGC AAG ATC ATC CTG CGC CTG CAG GTG CCC AAG GGC Ala Phe Gly Ser Arg Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly 795 800 805	1202
AGC ACT GGT GCC TAC CTG AGC GCC ATC GGC GGC TTC GCC AGC GAG AAG Ser Thr Gly Ala Tyr Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys 810 815 820	1250
GAG ATC CTG CTG GAT AAG GAC AGC AAG TAC CAC ATC GAC AAG GTG ACC Glu Ile Leu Leu Asp Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr 825 830 835 840	1298
GAG GTG ATC ATC AAG GGC GTG AAG CGC TAC GTG GTG GAC GCC ACC CTG Glu Val Ile Ile Lys Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu 845 850 855	1346
CTG ACC AAC TAG Leu Thr Asn	1358

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 449 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Ala Ala Gly 1 5 10 15
Val His Cys Leu Ser Ser Ser Phe Ala Asp Ser Asn Pro Ile Arg 20 25 30
Val Thr Asp Arg Ala Ala Ser Thr Leu Gln Asn Leu Lys Ile Thr Asp 35 40 45
Lys Val Glu Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly 50 55 60

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Lys Glu Lys Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys
65 70 75 80

Met Asn Asn Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys
85 90 95

Glu Ile Thr Phe Ser Ile Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp
100 105 110

Leu Lys Glu Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser
115 120 125

Ile Ile Thr Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys
130 135 140

Ser Leu Thr Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe
145 150 155 160

Lys Glu Gln Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp
165 170 175

Thr His Leu Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu
180 185 190

Lys Val Thr Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala
195 200 205

Gly Val Ile Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly
210 215 220

Tyr Met Val His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val
225 230 235 240

Glu Cys Leu Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys
245 250 255

Asn Asp Ile Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu
260 265 270

Glu Trp Ala Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly
275 280 285

Tyr Ala Arg Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln
290 295 300

Gly Gly Ser Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser
305 310 315 320

Asp Ala Leu Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg
325 330 335

Trp Cys Gly Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro
340 345 350

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Ser Leu Lys Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp
355 360 365

Lys Gly Tyr Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe
370 375 380

Gly Ser Arg Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr
385 390 395 400

Gly Ala Tyr Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile
405 410 415

Leu Leu Asp Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val
420 425 430

Ile Ile Lys Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr
435 440 445

Asn

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1..16
- (D) OTHER INFORMATION: /note= "linker peptide for fusion
of VIP1A(a) and VIP2A(a) used to construct pCIB5533"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Thr Pro Pro Pro Thr Pro Ser
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:48:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 66 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

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(A) DESCRIPTION: /desc = "DNA encoding linker peptide used to construct pCIB5533"

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

CCCGGGCCTT CTACTCCCC AACTCCCTCT CCTAGCACGC CTCCGACACC TAGCGATATC	60
GGATCC	66

(2) INFORMATION FOR SEQ ID NO:49:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4031 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "Synthetic DNA"

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 6..4019

(D) OTHER INFORMATION: /note= "Maize optimized DNA sequence encoding a VIP2A(a) - VIPIA(a) fusion protein as contained in pCIB5531"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

GATCC ATG AAG CGC ATG GAG GGC AAG CTG TTC ATG GTG AGC AAG AAG Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys	47
450 455 460	

CTC CAG GTG GTG ACC AAG ACC GTG CTG CTG AGC ACC GTG TTC AGC ATC Leu Gln Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile	95
465 470 475	

AGC CTG CTG AAC AAC GAG GTG ATC AAG GCC GAG CAG CTG AAC ATC AAC Ser Leu Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn	143
480 485 490 495	

AGC CAG AGC AAG TAC ACC AAC CTC CAG AAC CTG AAG ATC ACC GAC AAG Ser Gln Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Thr Asp Lys	191
500 505 510	

GTG GAG GAC TTC AAG GAG GAC AAG GAG AAG GCC AAG GAG TGG GGC AAG	239
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Val Glu Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys		
515	520	525
GAG AAG GAG AAG GAG TGG AAG CTT ACC GCC ACC GAG AAG GGC AAG ATG		287
Glu Lys Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met		
530	535	540
AAC AAC TTC CTG GAC AAC AAG AAC GAC ATC AAG ACC AAC TAC AAG GAG		335
Asn Asn Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu		
545	550	555
ATC ACC TTC AGC ATA GCC GGC AGC TTC GAG GAC GAG ATC AAG GAC CTG		383
Ile Thr Phe Ser Ile Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu		
560	565	570
575		
AAG GAG ATC GAC AAG ATG TTC GAC AAG ACC AAC CTG AGC AAC AGC ATC		431
Lys Glu Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile		
580	585	590
ATC ACC TAC AAG AAC GTG GAG CCC ACC ACC ATC GGC TTC AAC AAG AGC		479
Ile Thr Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser		
595	600	605
CTG ACC GAG GGC AAC ACC ATC AAC AGC GAC GCC ATG GCC CAG TTC AAG		527
Leu Thr Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys		
610	615	620
GAG CAG TTC CTG GAC CGC GAC ATC AAG TTC GAC AGC TAC CTG GAC ACC		575
Glu Gln Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr		
625	630	635
CAC CTG ACC GCC CAG CAG GTG AGC AGC AAG GAG CGC GTG ATC CTG AAG		623
His Leu Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys		
640	645	650
655		
GTG ACC GTC CCC AGC GGC AAG GGC AGC ACC ACC CCC ACC AAG GCC GGC		671
Val Thr Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly		
660	665	670
GTG ATC CTG AAC AAC AGC GAG TAC AAG ATG CTG ATC GAC AAC GGC TAC		719
Val Ile Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr		
675	680	685
ATG GTG CAC GTG GAC AAG GTG AGC AAG GTG GTG AAG AAG GGC GTG GAG		767
Met Val His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu		
690	695	700
TGC CTC CAG ATC GAG GGC ACC CTG AAG AAG AGT CTA GAC TTC AAG AAC		815
Cys Leu Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn		
705	710	715
GAC ATC AAC GCC GAG GCC CAC AGC TGG GGC ATG AAG AAC TAC GAG GAG		863
Asp Ile Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu		
720	725	730
735		

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TGG GCC AAG GAC CTG ACC GAC AGC CAG CGC GAG GCC CTG GAC GGC TAC Trp Ala Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr 740 745 750	911
GCC CGC CAG GAC TAC AAG GAG ATC AAC AAC TAC CTG CGC AAC CAG GGC Ala Arg Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly 755 760 765	959
GGC AGC GGC AAC GAG AAG CTG GAC GCC CAG ATC AAG AAC ATC AGC GAC Gly Ser Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp 770 775 780	1007
GCC CTG GGC AAG AAG CCC ATC CCC GAG AAC ATC ACC GTG TAC CGC TGG Ala Leu Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp 785 790 795	1055
TGC GGC ATG CCC GAG TTC GGC TAC CAG ATC AGC GAC CCC CTG CCC AGC Cys Gly Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser 800 805 810 815	1103
CTG AAG GAC TTC GAG GAG CAG TTC CTG AAC ACC ATC AAG GAG GAC AAG Leu Lys Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys 820 825 830	1151
GGC TAC ATG AGC ACC AGC CTG AGC AGC GAG CGC CTG GCC GCC TTC GGC Gly Tyr Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly 835 840 845	1199
AGC CGC AAG ATC ATC CTG CGC CTG CAG GTG CCC AAG GGC AGC ACT GGT Ser Arg Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly 850 855 860	1247
GCC TAC CTG AGC GCC ATC GGC GGC TTC GCC AGC GAG AAG GAG ATC CTG Ala Tyr Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu 865 870 875	1295
CTG GAT AAG GAC AGC AAG TAC CAC ATC GAC AAG GTG ACC GAG GTG ATC Leu Asp Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile 880 885 890 895	1343
ATC AAG GGC GTG AAG CGC TAC GTG GTG GAC GCC ACC CTG CTG ACC AAC Ile Lys Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn 900 905 910	1391
TCC CGG GGG CCT TCT ACT CCC CCA ACT CCC TCT CCT AGC ACG CCT CCG Ser Arg Gly Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Thr Pro Pro 915 920 925	1439
ACA CCT AGC GAT ATC GGA TCC ACC ATG AAG ACC AAC CAG ATC AGC ACC Thr Pro Ser Asp Ile Gly Ser Thr Met Lys Thr Asn Gln Ile Ser Thr 930 935 940	1487
ACC CAG AAG AAC CAG CAG AAG GAG ATG GAC CGC AAG GGC CTG CTG GGC Thr Gln Lys Asn Gln Gln Lys Glu Met Asp Arg Lys Gly Leu Leu Gly 945 950 955	1535

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TAC TAC TTC AAG GGC AAG GAC TTC AGC AAC CTG ACC ATG TTC GCC CCC Tyr Tyr Phe Lys Gly Lys Asp Phe Ser Asn Leu Thr Met Phe Ala Pro 960 965 970 975	1583
ACG CGT GAC AGC ACC CTG ATC TAC GAC CAG CAG ACC GCC AAC AAG CTG Thr Arg Asp Ser Thr Leu Ile Tyr Asp Gln Gln Thr Ala Asn Lys Leu 980 985 990	1631
CTG GAC AAG AAG CAG CAG GAG TAC CAG AGC ATC CGC TGG ATC GGC CTG Leu Asp Lys Lys Gln Gln Glu Tyr Gln Ser Ile Arg Trp Ile Gly Leu 995 1000 1005	1679
ATC CAG AGC AAG GAG ACC GGC GAC TTC ACC AAC CTG AGC GAG GAC Ile Gln Ser Lys Glu Thr Gly Asp Phe Thr Phe Asn Leu Ser Glu Asp 1010 1015 1020	1727
GAG CAG GCC ATC ATC GAG ATC AAC GGC AAG ATC ATC AGC AAC AAG GGC Glu Gln Ala Ile Ile Glu Ile Asn Gly Lys Ile Ile Ser Asn Lys Gly 1025 1030 1035	1775
AAG GAG AAG CAG GTG GTG CAC CTG GAG AAG GGC AAG CTG GTG CCC ATC Lys Glu Lys Gln Val Val His Leu Glu Lys Gly Lys Leu Val Pro Ile 1040 1045 1050 1055	1823
AAG ATC GAG TAC CAG AGC GAC ACC AAG TTC AAC ATC GAC AGC AAG ACC Lys Ile Glu Tyr Gln Ser Asp Thr Lys Phe Asn Ile Asp Ser Lys Thr 1060 1065 1070	1871
TTC AAG GAG CTG AAG CTT TTC AAG ATC GAC AGC CAG AAC CAG CCC CAG Phe Lys Glu Leu Lys Leu Phe Lys Ile Asp Ser Gln Asn Gln Pro Gln 1075 1080 1085	1919
CAG GTG CAG CAG GAC GAG CTG CGC AAC CCC GAG TTC AAC AAG AAG GAG Gln Val Gln Gln Asp Glu Leu Arg Asn Pro Glu Phe Asn Lys Lys Glu 1090 1095 1100	1967
AGC CAG GAG TTC CTG GCC AAG CCC AGC AAG ATC AAC CTG TTC ACC CAG Ser Gln Glu Phe Leu Ala Lys Pro Ser Lys Ile Asn Leu Phe Thr Gln 1105 1110 1115	2015
CAG ATG AAG CGC GAG ATC GAC GAG GAC ACC GAC ACC GAC GGC GAC AGC Gln Met Lys Arg Glu Ile Asp Glu Asp Thr Asp Thr Asp Gly Asp Ser 1120 1125 1130 1135	2063
ATC CCC GAC CTG TGG GAG GAG AAC GGC TAC ACC ATC CAG AAC CGC ATC Ile Pro Asp Leu Trp Glu Glu Asn Gly Tyr Thr Ile Gln Asn Arg Ile 1140 1145 1150	2111
GCC GTG AAG TGG GAC GAC AGC CTG GCT AGC AAG GGC TAC ACC AAG TTC Ala Val Lys Trp Asp Asp Ser Leu Ala Ser Lys Gly Tyr Thr Lys Phe 1155 1160 1165	2159
GTG AGC AAC CCC CTG GAG AGC CAC ACC GTG GGC GAC CCC TAC ACC GAC Val Ser Asn Pro Leu Glu Ser His Thr Val Gly Asp Pro Tyr Thr Asp	2207

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1170	1175	1180	
TAC GAG AAG GCC GCC CGC GAC CTG GAC CTG AGC AAC GCC AAG GAG ACC Tyr Glu Lys Ala Ala Arg Asp Leu Asp Leu Ser Asn Ala Lys Glu Thr 1185	1190	1195	2255
TTC AAC CCC CTG GTG GCC GCC TTC CCC AGC GTG AAC GTG AGC ATG GAG Phe Asn Pro Leu Val Ala Ala Phe Pro Ser Val Asn Val Ser Met Glu 1200	1205	1210	2303
AAG GTG ATC CTG AGC CCC AAC GAG AAC CTG AGC AAC AGC GTG GAG AGC Lys Val Ile Leu Ser Pro Asn Glu Asn Leu Ser Asn Ser Val Glu Ser 1220	1225	1230	2351
CAC TCG AGC ACC AAC TGG AGC TAC ACC AAC ACC GAG GGC GCC AGC GTG His Ser Ser Thr Asn Trp Ser Tyr Thr Asn Thr Glu Gly Ala Ser Val 1235	1240	1245	2399
GAG GCC GGC ATC GGT CCC AAG GGC ATC AGC TTC GGC GTG AGC GTG AAC Glu Ala Gly Ile Gly Pro Lys Gly Ile Ser Phe Gly Val Ser Val Asn 1250	1255	1260	2447
TAC CAG CAC AGC GAG ACC GTG GCC CAG GAG TGG GGC ACC AGC ACC GGC Tyr Gln His Ser Glu Thr Val Ala Gln Glu Trp Gly Thr Ser Thr Gly 1265	1270	1275	2495
AAC ACC AGC CAG TTC AAC ACC GCC AGC GCC GGC TAC CTG AAC GCC AAC Asn Thr Ser Gln Phe Asn Thr Ala Ser Ala Gly Tyr Leu Asn Ala Asn 1280	1285	1290	2543
GTG CGC TAC AAC AAC GTG GGC ACC GGC GCC ATC TAC GAC GTG AAG CCC Val Arg Tyr Asn Asn Val Gly Thr Gly Ala Ile Tyr Asp Val Lys Pro 1300	1305	1310	2591
ACC ACC AGC TTC GTG CTG AAC AAC GAC ACC ATC GCC ACC ATC ACC GCC Thr Thr Ser Phe Val Leu Asn Asn Asp Thr Ile Ala Thr Ile Thr Ala 1315	1320	1325	2639
AAG TCG AAT TCC ACC GCC CTG AAC ATC AGC CCC GGC GAG AGC TAC CCC Lys Ser Asn Ser Thr Ala Leu Asn Ile Ser Pro Gly Glu Ser Tyr Pro 1330	1335	1340	2687
AAG AAG GGC CAG AAC GGC ATC GCC ATC ACC AGC ATG GAC GAC TTC AAC Lys Lys Gly Gln Asn Gly Ile Ala Ile Thr Ser Met Asp Asp Phe Asn 1345	1350	1355	2735
AGC CAC CCC ATC ACC CTG AAC AAG AAG CAG GTG GAC AAC CTG CTG AAC Ser His Pro Ile Thr Leu Asn Lys Lys Gln Val Asp Asn Leu Leu Asn 1360	1365	1370	2783
AAC AAG CCC ATG ATG CTG GAG ACC AAC CAG ACC GAC GGC GTC TAC AAG Asn Lys Pro Met Met Leu Glu Thr Asn Gln Thr Asp Gly Val Tyr Lys 1380	1385	1390	2831
ATC AAG GAC ACC CAC GGC AAC ATC GTG ACG GGC GGC GAG TGG AAC GGC			2879

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Ile Lys Asp Thr His Gly Asn Ile Val Thr Gly Gly Glu Trp Asn Gly 1395 1400 1405		
GTG ATC CAG CAG ATC AAG GCC AAG ACC GCC AGC ATC ATC GTC GAC GAC Val Ile Gln Gln Ile Lys Ala Lys Thr Ala Ser Ile Ile Val Asp Asp 1410 1415 1420		2927
GGC GAG CGC GTG GCC GAG AAG CGC GTG GCC GCC AAG GAC TAC GAG AAC Gly Glu Arg Val Ala Glu Lys Arg Val Ala Ala Lys Asp Tyr Glu Asn 1425 1430 1435		2975
CCC GAG GAC AAG ACC CCC AGC CTG ACC CTG AAG GAC GCC CTG AAG CTG Pro Glu Asp Lys Thr Pro Ser Leu Thr Leu Lys Asp Ala Leu Lys Leu 1440 1445 1450		3023
AGC TAC CCC GAC GAG ATC AAG GAG ATC GAG GGC TTG CTG TAC TAC AAG Ser Tyr Pro Asp Glu Ile Lys Glu Ile Glu Gly Leu Leu Tyr Tyr Lys 1460 1465 1470		3071
AAC AAG CCC ATC TAC GAG AGC AGC GTG ATG ACC TAT CTA GAC GAG AAC Asn Lys Pro Ile Tyr Glu Ser Ser Val Met Thr Tyr Leu Asp Glu Asn 1475 1480 1485		3119
ACC GCC AAG GAG GTG ACC AAG CAG CTG AAC GAC ACC ACC GGC AAG TTC Thr Ala Lys Glu Val Thr Lys Gln Leu Asn Asp Thr Thr Gly Lys Phe 1490 1495 1500		3167
AAG GAC GTG AGC CAC CTG TAC GAC GTG AAG CTG ACC CCC AAG ATG AAC Lys Asp Val Ser His Leu Tyr Asp Val Lys Leu Thr Pro Lys Met Asn 1505 1510 1515		3215
GTG ACC ATC AAG CTG AGC ATC CTG TAC GAC AAC GCC GAG AGC AAC GAC Val Thr Ile Lys Leu Ser Ile Leu Tyr Asp Asn Ala Glu Ser Asn Asp 1520 1525 1530		3263
AAC AGC ATC GGC AAG TGG ACC AAC ACC AAC ATC GTG AGC GGC GGC AAC Asn Ser Ile Gly Lys Trp Thr Asn Thr Asn Ile Val Ser Gly Gly Asn 1540 1545 1550		3311
AAC GGC AAG AAG CAG TAC AGC AGC AAC AAC CCC GAC GCC AAC CTG ACC Asn Gly Lys Gln Tyr Ser Ser Asn Asn Pro Asp Ala Asn Leu Thr 1555 1560 1565		3359
CTG AAC ACC GAC GCC CAG GAG AAG CTG AAC AAG AAC CGC GAC TAC TAC Leu Asn Thr Asp Ala Gln Glu Lys Leu Asn Lys Asn Arg Asp Tyr Tyr 1570 1575 1580		3407
ATC AGC CTG TAC ATG AAG AGC GAG AAG AAC ACC CAG TGC GAG ATC ACC Ile Ser Leu Tyr Met Lys Ser Glu Lys Asn Thr Gln Cys Glu Ile Thr 1585 1590 1595		3455
ATC GAC GGC GAG ATA TAC CCC ATC ACC ACC AAG ACC GTG AAC GTG AAC Ile Asp Gly Glu Ile Tyr Pro Ile Thr Thr Lys Thr Val Asn Val Asn 1600 1605 1610		3503
	1615	

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AAG GAC AAC TAC AAG CGC CTG GAC ATC ATC GCC CAC AAC ATC AAG AGC Lys Asp Asn Tyr Lys Arg Leu Asp Ile Ile Ala His Asn Ile Lys Ser 1620 1625 1630	3551
AAC CCC ATC AGC AGC CTG CAC ATC AAG ACC AAC GAC GAG ATC ACC CTG Asn Pro Ile Ser Ser Leu His Ile Lys Thr Asn Asp Glu Ile Thr Leu 1635 1640 1645	3599
TTC TGG GAC GAC ATA TCG ATT ACC GAC GTC GCC AGC ATC AAG CCC GAG Phe Trp Asp Asp Ile Ser Ile Thr Asp Val Ala Ser Ile Lys Pro Glu 1650 1655 1660	3647
AAC CTG ACC GAC AGC GAG ATC AAG CAG ATA TAC AGT CGC TAC GGC ATC Asn Leu Thr Asp Ser Glu Ile Lys Gln Ile Tyr Ser Arg Tyr Gly Ile 1665 1670 1675	3695
AAG CTG GAG GAC GGC ATC CTG ATC GAC AAG AAA GGC GGC ATC CAC TAC Lys Leu Glu Asp Gly Ile Leu Ile Asp Lys Lys Gly Gly Ile His Tyr 1680 1685 1690 1695	3743
GGC GAG TTC ATC AAC GAG GCC AGC TTC AAC ATC GAG CCC CTG CAG AAC Gly Glu Phe Ile Asn Glu Ala Ser Phe Asn Ile Glu Pro Leu Gln Asn 1700 1705 1710	3791
TAC GTG ACC AAG TAC GAG GTG ACC TAC AGC AGC GAG CTG GGC CCC AAC Tyr Val Thr Lys Tyr Glu Val Thr Tyr Ser Ser Glu Leu Gly Pro Asn 1715 1720 1725	3839
GTG AGC GAC ACC CTG GAG AGC GAC AAG ATT TAC AAG GAC GGC ACC ATC Val Ser Asp Thr Leu Glu Ser Asp Lys Ile Tyr Lys Asp Gly Thr Ile 1730 1735 1740	3887
AAG TTC GAC TTC ACC AAG TAC AGC AAG AAC GAG CAG GGC CTG TTC TAC Lys Phe Asp Phe Thr Lys Tyr Ser Lys Asn Glu Gln Gly Leu Phe Tyr 1745 1750 1755	3935
GAC AGC GGC CTG AAC TGG GAC TTC AAG ATC AAC GCC ATC ACC TAC GAC Asp Ser Gly Leu Asn Trp Asp Phe Lys Ile Asn Ala Ile Thr Tyr Asp 1760 1765 1770 1775	3983
GGC AAG GAG ATG AAC GTG TTC CAC CGC TAC AAC AAG TAGATCTGAG Gly Lys Glu Met Asn Val Phe His Arg Tyr Asn Lys 1780 1785	4029
CT	4031

(2) INFORMATION FOR SEQ ID NO:50:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1338 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

Met Lys Arg Met Glu Gly Lys Leu Phe Met Val Ser Lys Lys Leu Gln
1 5 10 15

Val Val Thr Lys Thr Val Leu Leu Ser Thr Val Phe Ser Ile Ser Leu
20 25 30

Leu Asn Asn Glu Val Ile Lys Ala Glu Gln Leu Asn Ile Asn Ser Gln
35 40 45

Ser Lys Tyr Thr Asn Leu Gln Asn Leu Lys Ile Thr Asp Lys Val Glu
50 55 60

Asp Phe Lys Glu Asp Lys Glu Lys Ala Lys Glu Trp Gly Lys Glu Lys
65 70 75 80

Glu Lys Glu Trp Lys Leu Thr Ala Thr Glu Lys Gly Lys Met Asn Asn
85 90 95

Phe Leu Asp Asn Lys Asn Asp Ile Lys Thr Asn Tyr Lys Glu Ile Thr
100 105 110

Phe Ser Ile Ala Gly Ser Phe Glu Asp Glu Ile Lys Asp Leu Lys Glu
115 120 125

Ile Asp Lys Met Phe Asp Lys Thr Asn Leu Ser Asn Ser Ile Ile Thr
130 135 140

Tyr Lys Asn Val Glu Pro Thr Thr Ile Gly Phe Asn Lys Ser Leu Thr
145 150 155 160

Glu Gly Asn Thr Ile Asn Ser Asp Ala Met Ala Gln Phe Lys Glu Gln
165 170 175

Phe Leu Asp Arg Asp Ile Lys Phe Asp Ser Tyr Leu Asp Thr His Leu
180 185 190

Thr Ala Gln Gln Val Ser Ser Lys Glu Arg Val Ile Leu Lys Val Thr
195 200 205

Val Pro Ser Gly Lys Gly Ser Thr Thr Pro Thr Lys Ala Gly Val Ile
210 215 220

Leu Asn Asn Ser Glu Tyr Lys Met Leu Ile Asp Asn Gly Tyr Met Val
225 230 235 240

His Val Asp Lys Val Ser Lys Val Val Lys Lys Gly Val Glu Cys Leu
245 250 255

Gln Ile Glu Gly Thr Leu Lys Lys Ser Leu Asp Phe Lys Asn Asp Ile
260 265 270

Asn Ala Glu Ala His Ser Trp Gly Met Lys Asn Tyr Glu Glu Trp Ala

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275	280	285
Lys Asp Leu Thr Asp Ser Gln Arg Glu Ala Leu Asp Gly Tyr Ala Arg		
290	295	300
Gln Asp Tyr Lys Glu Ile Asn Asn Tyr Leu Arg Asn Gln Gly Gly Ser		
305	310	315
320		
Gly Asn Glu Lys Leu Asp Ala Gln Ile Lys Asn Ile Ser Asp Ala Leu		
325	330	335
Gly Lys Lys Pro Ile Pro Glu Asn Ile Thr Val Tyr Arg Trp Cys Gly		
340	345	350
Met Pro Glu Phe Gly Tyr Gln Ile Ser Asp Pro Leu Pro Ser Leu Lys		
355	360	365
Asp Phe Glu Glu Gln Phe Leu Asn Thr Ile Lys Glu Asp Lys Gly Tyr		
370	375	380
Met Ser Thr Ser Leu Ser Ser Glu Arg Leu Ala Ala Phe Gly Ser Arg		
385	390	395
400		
Lys Ile Ile Leu Arg Leu Gln Val Pro Lys Gly Ser Thr Gly Ala Tyr		
405	410	415
Leu Ser Ala Ile Gly Gly Phe Ala Ser Glu Lys Glu Ile Leu Leu Asp		
420	425	430
Lys Asp Ser Lys Tyr His Ile Asp Lys Val Thr Glu Val Ile Ile Lys		
435	440	445
Gly Val Lys Arg Tyr Val Val Asp Ala Thr Leu Leu Thr Asn Ser Arg		
450	455	460
Gly Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Thr Pro Pro Thr Pro		
465	470	475
480		
Ser Asp Ile Gly Ser Thr Met Lys Thr Asn Gln Ile Ser Thr Thr Gln		
485	490	495
Lys Asn Gln Gln Lys Glu Met Asp Arg Lys Gly Leu Leu Gly Tyr Tyr		
500	505	510
Phe Lys Gly Lys Asp Phe Ser Asn Leu Thr Met Phe Ala Pro Thr Arg		
515	520	525
Asp Ser Thr Ile Ile Tyr Asp Gln Gln Thr Ala Asn Lys Leu Leu Asp		
530	535	540
Lys Lys Gln Gln Glu Tyr Gln Ser Ile Arg Trp Ile Gly Leu Ile Gln		
545	550	555
560		
Ser Lys Glu Thr Gly Asp Phe Thr Phe Asn Leu Ser Glu Asp Glu Gln		
565	570	575

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Ala Ile Ile Glu Ile Asn Gly Lys Ile Ile Ser Asn Lys Gly Lys Glu
580 585 590

Lys Gln Val Val His Leu Glu Lys Gly Lys Leu Val Pro Ile Lys Ile
595 600 605

Glu Tyr Gln Ser Asp Thr Lys Phe Asn Ile Asp Ser Lys Thr Phe Lys
610 615 620

Glu Leu Lys Leu Phe Lys Ile Asp Ser Gln Asn Gln Pro Gln Gln Val
625 630 635 640

Gln Gln Asp Glu Leu Arg Asn Pro Glu Phe Asn Lys Lys Glu Ser Gln
645 650 655

Glu Phe Leu Ala Lys Pro Ser Lys Ile Asn Leu Phe Thr Gln Gln Met
660 665 670

Lys Arg Glu Ile Asp Glu Asp Thr Asp Thr Asp Gly Asp Ser Ile Pro
675 680 685

Asp Leu Trp Glu Glu Asn Gly Tyr Thr Ile Gln Asn Arg Ile Ala Val
690 695 700

Lys Trp Asp Asp Ser Leu Ala Ser Lys Gly Tyr Thr Lys Phe Val Ser
705 710 715 720

Asn Pro Leu Glu Ser His Thr Val Gly Asp Pro Tyr Thr Asp Tyr Glu
725 730 735

Lys Ala Ala Arg Asp Leu Asp Leu Ser Asn Ala Lys Glu Thr Phe Asn
740 745 750

Pro Leu Val Ala Ala Phe Pro Ser Val Asn Val Ser Met Glu Lys Val
755 760 765

Ile Leu Ser Pro Asn Glu Asn Leu Ser Asn Ser Val Glu Ser His Ser
770 775 780

Ser Thr Asn Trp Ser Tyr Thr Asn Thr Glu Gly Ala Ser Val Glu Ala
785 790 795 800

Gly Ile Gly Pro Lys Gly Ile Ser Phe Gly Val Ser Val Asn Tyr Gln
805 810 815

His Ser Glu Thr Val Ala Gln Glu Trp Gly Thr Ser Thr Gly Asn Thr
820 825 830

Ser Gln Phe Asn Thr Ala Ser Ala Gly Tyr Leu Asn Ala Asn Val Arg
835 840 845

Tyr Asn Asn Val Gly Thr Gly Ala Ile Tyr Asp Val Lys Pro Thr Thr
850 855 860

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Ser Phe Val Leu Asn Asn Asp Thr Ile Ala Thr Ile Thr Ala Lys Ser
865 870 875 880

Asn Ser Thr Ala Leu Asn Ile Ser Pro Gly Glu Ser Tyr Pro Lys Lys
885 890 895

Gly Gln Asn Gly Ile Ala Ile Thr Ser Met Asp Asp Phe Asn Ser His
900 905 910

Pro Ile Thr Leu Asn Lys Lys Gln Val Asp Asn Leu Leu Asn Asn Lys
915 920 925

Pro Met Met Leu Glu Thr Asn Gln Thr Asp Gly Val Tyr Lys Ile Lys
930 935 940

Asp Thr His Gly Asn Ile Val Thr Gly Gly Glu Trp Asn Gly Val Ile
945 950 955 960

Gln Gln Ile Lys Ala Lys Thr Ala Ser Ile Ile Val Asp Asp Gly Glu
965 970 975

Arg Val Ala Glu Lys Arg Val Ala Ala Lys Asp Tyr Glu Asn Pro Glu
980 985 990

Asp Lys Thr Pro Ser Leu Thr Leu Lys Asp Ala Leu Lys Leu Ser Tyr
995 1000 1005

Pro Asp Glu Ile Lys Glu Ile Glu Gly Leu Leu Tyr Tyr Lys Asn Lys
1010 1015 1020

Pro Ile Tyr Glu Ser Ser Val Met Thr Tyr Leu Asp Glu Asn Thr Ala
1025 1030 1035 1040

Lys Glu Val Thr Lys Gln Leu Asn Asp Thr Thr Gly Lys Phe Lys Asp
1045 1050 1055

Val Ser His Leu Tyr Asp Val Lys Leu Thr Pro Lys Met Asn Val Thr
1060 1065 1070

Ile Lys Leu Ser Ile Leu Tyr Asp Asn Ala Glu Ser Asn Asp Asn Ser
1075 1080 1085

Ile Gly Lys Trp Thr Asn Thr Asn Ile Val Ser Gly Gly Asn Asn Gly
1090 1095 1100

Lys Lys Gln Tyr Ser Ser Asn Asn Pro Asp Ala Asn Leu Thr Leu Asn
1105 1110 1115 1120

Thr Asp Ala Gln Glu Lys Leu Asn Lys Asn Arg Asp Tyr Tyr Ile Ser
1125 1130 1135

Leu Tyr Met Lys Ser Glu Lys Asn Thr Gln Cys Glu Ile Thr Ile Asp
1140 1145 1150

Gly Glu Ile Tyr Pro Ile Thr Thr Lys Thr Val Asn Val Asn Lys Asp

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1155	1160	1165
Asn Tyr Lys Arg Leu Asp Ile Ile Ala His Asn Ile Lys Ser Asn Pro		
1170	1175	1180
Ile Ser Ser Leu His Ile Lys Thr Asn Asp Glu Ile Thr Leu Phe Trp		
1185	1190	1195
Asp Asp Ile Ser Ile Thr Asp Val Ala Ser Ile Lys Pro Glu Asn Leu		
1205	1210	1215
Thr Asp Ser Glu Ile Lys Gln Ile Tyr Ser Arg Tyr Gly Ile Lys Leu		
1220	1225	1230
Glu Asp Gly Ile Leu Ile Asp Lys Lys Gly Gly Ile His Tyr Gly Glu		
1235	1240	1245
Phe Ile Asn Glu Ala Ser Phe Asn Ile Glu Pro Leu Gln Asn Tyr Val		
1250	1255	1260
Thr Lys Tyr Glu Val Thr Tyr Ser Ser Glu Leu Gly Pro Asn Val Ser		
1265	1270	1275
Asp Thr Leu Glu Ser Asp Lys Ile Tyr Lys Asp Gly Thr Ile Lys Phe		
1285	1290	1295
Asp Phe Thr Lys Tyr Ser Lys Asn Glu Gln Gly Leu Phe Tyr Asp Ser		
1300	1305	1310
Gly Leu Asn Trp Asp Phe Lys Ile Asn Ala Ile Thr Tyr Asp Gly Lys		
1315	1320	1325
Glu Met Asn Val Phe His Arg Tyr Asn Lys		
1330	1335	

(2) INFORMATION FOR SEQ ID NO:51:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2444 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 17..2444
- (D) OTHER INFORMATION: /product= "3A(a) synthetic:native fusion"

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

GGATCCACCA ATGAAAC ATG AAC AAG AAC ACC AAG CTG AGC ACC CGC Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg 1 5 10	49
GCC CTG CCG AGC TTC ATC GAC TAC TTC AAC GGC ATC TAC GGC TTC GCC Ala Leu Pro Ser Phe Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala 15 20 25	97
ACC GGC ATC AAG GAC ATC ATG AAC ATG ATC TTC AAG ACC GAC ACC GGC Thr Gly Ile Lys Asp Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly 30 35 40	145
GGC GAC CTG ACC CTG GAC GAG ATC CTG AAG AAC CAG CAG CTG CTG AAC Gly Asp Leu Thr Leu Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn 45 50 55	193
GAC ATC AGC GGC AAG CTG GAC GGC GTG AAC GGC AGC CTG AAC GAC CTG Asp Ile Ser Gly Lys Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu 60 65 70 75	241
ATC GCC CAG GGC AAC CTG AAC ACC GAG CTG AGC AAG GAG ATC CTT AAG Ile Ala Gln Gly Asn Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys 80 85 90	289
ATC GCC AAC GAG CAG AAC CAG GTG CTG AAC GAC GTG AAC AAC AAG CTG Ile Ala Asn Glu Gln Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu 95 100 105	337
GAC GCC ATC AAC ACC ATG CTG CGC GTG TAC CTG CCG AAG ATC ACC ACC AGC Asp Ala Ile Asn Thr Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser 110 115 120	385
ATG CTG AGC GAC GTG ATG AAG CAG AAC TAC GCC CTG AGC CTG CAG ATC Met Leu Ser Asp Val Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile 125 130 135	433
GAG TAC CTG AGC AAG CAG CTG CAG GAG ATC AGC GAC AAG CTG GAC ATC Glu Tyr Leu Ser Lys Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile 140 145 150 155	481
ATC AAC GTG AAC GTC CTG ATC AAC AGC ACC CTG ACC GAG ATC ACC ACC CCG Ile Asn Val Asn Val Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro 160 165 170	529
GCC TAC CAG CGC ATC AAG TAC GTG AAC GAG AAG TTC GAA GAG CTG ACC Ala Tyr Gln Arg Ile Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr 175 180 185	577
TTC GCC ACC GAG ACC AGC AGC AAG GTG AAG AAG GAC GGC AGC CCG GCC Phe Ala Thr Glu Thr Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala 190 195 200	625
GAC ATC CTG GAC GAG CTG ACC GAG CTG ACC GAG CTG GCC AAG AGC GTG	673

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Asp Ile Leu Asp Glu Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val			
205	210	215	
ACC AAG AAC GAC GTG GAC GGC TTC GAG TTC TAC CTG AAC ACC TTC CAC			721
Thr Lys Asn Asp Val Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His			
220	225	230	235
GAC GTG ATG GTG GGC AAC AAC CTG TTC GGC CGC AGC GCC CTG AAG ACC			769
Asp Val Met Val Gly Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr			
240	245	250	
GCC AGC GAG CTG ATC ACC AAG GAG AAC GTG AAG ACC AGC GGC AGC GAG			817
Ala Ser Glu Leu Ile Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu			
255	260	265	
GTG GGC AAC GTG TAC AAC TTC CTG ATC GTG CTG ACC GCC CTG CAG GCC			865
Val Gly Asn Val Tyr Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala			
270	275	280	
CAG GCC TTC CTG ACC CTG ACC ACC TGT CGC AAG CTG CTG GGC CTG GCC			913
Gln Ala Phe Leu Thr Leu Thr Cys Arg Lys Leu Leu Gly Leu Ala			
285	290	295	
GAC ATC GAC TAC ACC AGC ATC ATG AAC GAG CAC TTG AAC AAG GAG AAG			961
Asp Ile Asp Tyr Thr Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys			
300	305	310	315
GAG GAG TTC CCG GTG AAC ATC CTG CCG ACC CTG AGC AAC ACC TTC AGC			1009
Glu Glu Phe Arg Val Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser			
320	325	330	
AAC CCG AAC TAC GCC AAG GTG AAG GGC AGC GAC GAG GAC GCC AAG ATG			1057
Asn Pro Asn Tyr Ala Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met			
335	340	345	
ATC GTG GAG GCT AAG CCG GGC CAC GCG TTG ATC GGC TTC GAG ATC AGC			1105
Ile Val Glu Ala Lys Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser			
350	355	360	
AAC GAC AGC ATC ACC GTG CTG AAG GTG TAC GAG GCC AAG CTG AAG CAG			1153
Asn Asp Ser Ile Thr Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln			
365	370	375	
AAC TAC CAG GTG GAC AAG GAC AGC TTG AGC GAG GTG ATC TAC GGC GAC			1201
Asn Tyr Gln Val Asp Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp			
380	385	390	395
ATG GAC AAG CTG CTG TGT CCG GAC CAG AGC GAG CAA ATC TAC TAC ACC			1249
Met Asp Lys Leu Leu Cys Pro Asp Gln Ser Glu Gln Ile Tyr Tyr Thr			
400	405	410	
AAC AAC ATC GTG TTC CCG AAC GAG TAC GTG ATC ACC AAG ATC GAC TTC			1297
Asn Asn Ile Val Phe Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe			
415	420	425	

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ACC AAG AAG ATG AAG ACC CTG CGC TAC GAG GTG ACC GCC AAC TTC TAC Thr Lys Lys Met Lys Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr 430	435	440	1345
GAC AGC AGC ACC GGC GAG ATC GAC CTG AAC AAG AAG GTG GAG AGC Asp Ser Ser Thr Gly Glu Ile Asp Leu Asn Lys Lys Lys Val Glu Ser 445	450	455	1393
AGC GAG GCC GAG TAC CGC ACC CTG AGC GCG AAC GAC GAC GGC GTC TAC Ser Glu Ala Glu Tyr Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr 460	465	470	475
ATG CCA CTG GGC GTG ATC AGC GAG ACC TTC CTG ACC CCG ATC AAC GGC Met Pro Leu Gly Val Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly 480	485	490	1489
TTT GGC CTG CAG GCC GAC GAG AAC AGC CGC CTG ATC ACC CTG ACC TGT Phe Gly Leu Gln Ala Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys 495	500	505	1537
AAG AGC TAC CTG CGC GAG CTG CTG CTA GCC ACC GAC CTG AGC AAC AAG Lys Ser Tyr Leu Arg Glu Leu Leu Ala Thr Asp Leu Ser Asn Lys 510	515	520	1585
GAG ACC AAG CTG ATC GTG CCA CCG AGC GGC TTC ATC AGC AAC ATC GTG Glu Thr Lys Leu Ile Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val 525	530	535	1633
GAG AAC GGC AGC ATC GAG GAG GAC AAC CTG GAG CCG TGG AAG GCC AAC Glu Asn Gly Ser Ile Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn 540	545	550	555
AAC AAG AAC GCC TAC GTG GAC CAC ACC GGC GGC GTG AAC GGC ACC AAG Asn Lys Asn Ala Tyr Val Asp His Thr Gly Gly Val Asn Gly Thr Lys 560	565	570	1729
GCC CTG TAC GTG CAC AAG GAC GGC GGC ATC AGC CAG TTC ATC GGC GAC Ala Leu Tyr Val His Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp 575	580	585	1777
AAG CTG AAG CCG AAG ACC GAG TAC GTG ATC CAG TAC ACC GTG AAG GGC Lys Leu Lys Pro Lys Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly 590	595	600	1825
AAG CCA TCG ATT CAC CTG AAG GAC GAG AAC ACC GGC TAC ATC CAC TAC Lys Pro Ser Ile His Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr 605	610	615	1873
GAG GAC ACC AAC AAC AAC CTG GAG GAC TAC CAG ACC ATC AAC AAG CGC Glu Asp Thr Asn Asn Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg 620	625	630	635
TTC ACC ACC GGC ACC GAC CTG AAG GGC GTG TAC CTG ATC CTG AAG AGC Phe Thr Thr Gly Thr Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser 640	645	650	1969

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CAG AAC GGC GAC GAG GCC TGG GGC GAC AAC TTC ATC ATC CTG GAG ATC Gln Asn Gly Asp Glu Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile 655 660 665	2017
AGC CCG AGC GAG AAG CTG CTG AGC CCG GAG CTG ATC AAC ACC AAC AAC Ser Pro Ser Glu Lys Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn 670 675 680	2065
TGG ACC AGC ACC GGC AGC ACC AAC ATC AGC GGC AAC ACC CTG ACC CTG Trp Thr Ser Thr Gly Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu 685 690 695	2113
TAC CAG GGC GGC CGG GGG ATT CTA AAA CAA AAC CTT CAA TTA GAT AGT Tyr Gln Gly Gly Arg Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser 700 705 710 715	2161
TTT TCA ACT TAT AGA GTG TAT TTT TCT GTG TCC GGA GAT GCT AAT GTA Phe Ser Thr Tyr Arg Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val 720 725 730	2209
AGG ATT AGA AAT TCT AGG GAA GTG TTA TTT GAA AAA AGA TAT ATG AGC Arg Ile Arg Asn Ser Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser 735 740 745	2257
GGT GCT AAA GAT GTT TCT GAA ATG TTC ACT ACA AAA TTT GAG AAA GAT Gly Ala Lys Asp Val Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp 750 755 760	2305
AAC TTT TAT ATA GAG CTT TCT CAA GGG AAT AAT TTA TAT GGT GGT CCT Asn Phe Tyr Ile Glu Leu Ser Gln Gly Asn Asn Leu Tyr Gly Gly Pro 765 770 775	2353
ATT GTA CAT TTT TAC GAT GTC TCT ATT AAG NAA GAT CGG GAT CTA ATA Ile Val His Phe Tyr Asp Val Ser Ile Lys Xaa Asp Arg Asp Leu Ile 780 785 790 795	2401
TTA ACA GTT TTT AAA AGC NAA TTC TTG TAT AAT GTC CTT GAT T Leu Thr Val Phe Lys Ser Xaa Phe Leu Tyr Asn Val Leu Asp 800 805	2444

(2) INFORMATION FOR SEQ ID NO:52:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 809 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

Met Asn Lys Asn Asn Thr Lys Leu Ser Thr Arg Ala Leu Pro Ser Phe
1 5 10 15

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Ile Asp Tyr Phe Asn Gly Ile Tyr Gly Phe Ala Thr Gly Ile Lys Asp
20 25 30

Ile Met Asn Met Ile Phe Lys Thr Asp Thr Gly Gly Asp Leu Thr Leu
35 40 45

Asp Glu Ile Leu Lys Asn Gln Gln Leu Leu Asn Asp Ile Ser Gly Lys
50 55 60

Leu Asp Gly Val Asn Gly Ser Leu Asn Asp Leu Ile Ala Gln Gly Asn
65 70 75 80

Leu Asn Thr Glu Leu Ser Lys Glu Ile Leu Lys Ile Ala Asn Glu Gln
85 90 95

Asn Gln Val Leu Asn Asp Val Asn Asn Lys Leu Asp Ala Ile Asn Thr
100 105 110

Met Leu Arg Val Tyr Leu Pro Lys Ile Thr Ser Met Leu Ser Asp Val
115 120 125

Met Lys Gln Asn Tyr Ala Leu Ser Leu Gln Ile Glu Tyr Leu Ser Lys
130 135 140

Gln Leu Gln Glu Ile Ser Asp Lys Leu Asp Ile Ile Asn Val Asn Val
145 150 155 160

Leu Ile Asn Ser Thr Leu Thr Glu Ile Thr Pro Ala Tyr Gln Arg Ile
165 170 175

Lys Tyr Val Asn Glu Lys Phe Glu Glu Leu Thr Phe Ala Thr Glu Thr
180 185 190

Ser Ser Lys Val Lys Lys Asp Gly Ser Pro Ala Asp Ile Leu Asp Glu
195 200 205

Leu Thr Glu Leu Thr Glu Leu Ala Lys Ser Val Thr Lys Asn Asp Val
210 215 220

Asp Gly Phe Glu Phe Tyr Leu Asn Thr Phe His Asp Val Met Val Gly
225 230 235 240

Asn Asn Leu Phe Gly Arg Ser Ala Leu Lys Thr Ala Ser Glu Leu Ile
245 250 255

Thr Lys Glu Asn Val Lys Thr Ser Gly Ser Glu Val Gly Asn Val Tyr
260 265 270

Asn Phe Leu Ile Val Leu Thr Ala Leu Gln Ala Phe Leu Thr
275 280 285

Leu Thr Thr Cys Arg Lys Leu Leu Gly Leu Ala Asp Ile Asp Tyr Thr
290 295 300

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Ser Ile Met Asn Glu His Leu Asn Lys Glu Lys Glu Glu Phe Arg Val
305 310 315 320

Asn Ile Leu Pro Thr Leu Ser Asn Thr Phe Ser Asn Pro Asn Tyr Ala
325 330 335

Lys Val Lys Gly Ser Asp Glu Asp Ala Lys Met Ile Val Glu Ala Lys
340 345 350

Pro Gly His Ala Leu Ile Gly Phe Glu Ile Ser Asn Asp Ser Ile Thr
355 360 365

Val Leu Lys Val Tyr Glu Ala Lys Leu Lys Gln Asn Tyr Gln Val Asp
370 375 380

Lys Asp Ser Leu Ser Glu Val Ile Tyr Gly Asp Met Asp Lys Leu Leu
385 390 395 400

Cys Pro Asp Gln Ser Glu Gln Ile Tyr Tyr Thr Asn Asn Ile Val Phe
405 410 415

Pro Asn Glu Tyr Val Ile Thr Lys Ile Asp Phe Thr Lys Lys Met Lys
420 425 430

Thr Leu Arg Tyr Glu Val Thr Ala Asn Phe Tyr Asp Ser Ser Thr Gly
435 440 445

Glu Ile Asp Leu Asn Lys Lys Val Glu Ser Ser Glu Ala Glu Tyr
450 455 460

Arg Thr Leu Ser Ala Asn Asp Asp Gly Val Tyr Met Pro Leu Gly Val
465 470 475 480

Ile Ser Glu Thr Phe Leu Thr Pro Ile Asn Gly Phe Gly Leu Gln Ala
485 490 495

Asp Glu Asn Ser Arg Leu Ile Thr Leu Thr Cys Lys Ser Tyr Leu Arg
500 505 510

Glu Leu Leu Leu Ala Thr Asp Leu Ser Asn Lys Glu Thr Lys Leu Ile
515 520 525

Val Pro Pro Ser Gly Phe Ile Ser Asn Ile Val Glu Asn Gly Ser Ile
530 535 540

Glu Glu Asp Asn Leu Glu Pro Trp Lys Ala Asn Asn Lys Asn Ala Tyr
545 550 555 560

Val Asp His Thr Gly Gly Val Asn Gly Thr Lys Ala Leu Tyr Val His
565 570 575

Lys Asp Gly Gly Ile Ser Gln Phe Ile Gly Asp Lys Leu Lys Pro Lys
580 585 590

Thr Glu Tyr Val Ile Gln Tyr Thr Val Lys Gly Lys Pro Ser Ile His

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595	600	605
Leu Lys Asp Glu Asn Thr Gly Tyr Ile His Tyr Glu Asp Thr Asn Asn		
610	615	620
Asn Leu Glu Asp Tyr Gln Thr Ile Asn Lys Arg Phe Thr Thr Gly Thr		
625	630	635
640		
Asp Leu Lys Gly Val Tyr Leu Ile Leu Lys Ser Gln Asn Gly Asp Glu		
645	650	655
Ala Trp Gly Asp Asn Phe Ile Ile Leu Glu Ile Ser Pro Ser Glu Lys		
660	665	670
Leu Leu Ser Pro Glu Leu Ile Asn Thr Asn Asn Trp Thr Ser Thr Gly		
675	680	685
Ser Thr Asn Ile Ser Gly Asn Thr Leu Thr Leu Tyr Gln Gly Gly Arg		
690	695	700
Gly Ile Leu Lys Gln Asn Leu Gln Leu Asp Ser Phe Ser Thr Tyr Arg		
705	710	715
720		
Val Tyr Phe Ser Val Ser Gly Asp Ala Asn Val Arg Ile Arg Asn Ser		
725	730	735
Arg Glu Val Leu Phe Glu Lys Arg Tyr Met Ser Gly Ala Lys Asp Val		
740	745	750
Ser Glu Met Phe Thr Thr Lys Phe Glu Lys Asp Asn Phe Tyr Ile Glu		
755	760	765
Leu Ser Gln Gly Asn Asn Leu Tyr Gly Gly Pro Ile Val His Phe Tyr		
770	775	780
Asp Val Ser Ile Lys Xaa Asp Arg Asp Leu Ile Leu Thr Val Phe Lys		
785	790	795
800		
Ser Xaa Phe Leu Tyr Asn Val Leu Asp		
805		

What is claimed is:

1. A substantially purified *Bacillus* strain which produces a pesticidal protein during vegetative growth wherein said *Bacillus* is not *B. sphaericus* SSII-1.
2. A *Bacillus* strain which produces a pesticidal protein during vegetative growth, wherein said *Bacillus* is *Bacillus cereus* having Accession No. NRRL B-21058.
3. A *Bacillus* strain which produces a pesticidal protein during vegetative growth, wherein said *Bacillus* is *Bacillus thuringiensis* having Accession No. NRRL B-21060
4. A *Bacillus* strain which produces a pesticidal protein during vegetative growth, wherein said *Bacillus* is a *Bacillus* selected from Accession Numbers NRRL B-21224, NRRL B-21225, NRRL B-21226, NRRL B-21227, NRRL B-21228, NRRL B-21229, NRRL B-21230, and NRRL B-21439.
5. An insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1.
6. The insect-specific protein of claim 5 wherein said *Bacillus* is selected from a *Bacillus thuringiensis* and *B. cereus*.
7. The insect-specific protein of claim 5 wherein said protein is toxic to Coleoptera or Lepidoptera.
8. The insect-specific protein of claim 5 wherein the spectrum of insecticidal activity includes an activity against *Agrotis* and/or *Spodoptera* species, but preferably a black cutworm [*Agrotis ipsilon* ; BCW] and/or fall armyworm [*Spodoptera frugiperda*] and/or beet armyworm [*Spodoptera exigua*] and/or tobacco budworm and/or corn earworm [*Helicoverpa zea*] activity.
9. The insect-specific protein of claim 5, wherein said *Bacillus* is *Bacillus cereus* having Accession No. NRRL B-21058.
10. The insect-specific protein of claim 5, wherein said *Bacillus* is *Bacillus thuringiensis* having Accession No. NRRL B-21060.

11. The insect-specific protein of claim 5, wherein said *Bacillus* is a *Bacillus* selected from Accession Numbers NRRL B-21224, NRRL B-21225, NRRL B-21226, NRRL B-21227, NRRL B-21228, NRRL B-21229, NRRL B-21230, and NRRL B-21439.
12. The insect-specific protein of claim 5 wherein said protein has a molecular weight of about 30 kDa or greater.
13. The insect-specific protein of claim 12 wherein said protein has a molecular weight of about 60 to about 100 kDa.
14. The insect-specific protein of claim 13, wherein said protein has a molecular weight of about 80 kDa.
15. The insect-specific protein of claim 5, wherein said protein comprises a sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:7, including homologues thereof.
16. The insect-specific protein of claim 5, wherein said protein has the sequence selected from the group consisting of SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:29 SEQ ID NO:32 and SEQ ID NO:2 including homologues thereof.
17. The insect-specific protein of claim 8, wherein said protein has the sequence selected from the group consisting of SEQ ID NO:29 and SEQ ID NO:32 including homologues thereof.
18. An insect-specific protein according to any one of claims 5 to 15, wherein the sequences representing the secretion signal have been removed or inactivated.
19. An auxiliary protein which enhances the insect-specific activity of an insect-specific protein.
20. The auxiliary protein of claim 19 wherein said auxiliary protein has a molecular weight of about 50 kDa.
21. The auxiliary protein of claim 19 wherein said auxiliary protein is from *Bacillus cereus*.
22. The auxiliary protein of any one of claims 19 to 21 wherein both the said auxiliary protein as well as said insect-specific protein is from strain AB78.

23. An auxiliary protein according to any one claims 19 to 22, wherein the sequences representing the secretion signal have been removed or inactivated.
24. A multimeric pesticidal protein, which comprises more than one polypeptide chain and wherein at least one of the said polypeptide chains represents an insect-specific protein of any one of claims 5 to 18 and at least one of the said polypeptide chains represents an auxiliary protein according to any one of claims 19 to 23, which activates or enhances the pesticidal activity of the said insect-specific protein.
25. The multimeric pesticidal protein according to claim 24 having a molecular weight of about 50 kDa to about 200 kDa.
26. The multimeric pesticidal protein of claim 25 comprising an insect-specific protein of any one of claims 5 to 18 and an auxiliary protein according to any one of claims 19 to 23, which activates or enhances the pesticidal activity of the said insect-specific protein.
27. A fusion protein comprising several protein domains including at least an insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 produced by in frame genetic fusions, which, when translated by ribosomes, produce a fusion protein with at least the combined attributes of the insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 and, optionally, of the other components used in the fusion.
28. A fusion protein according to claim 27, comprising a ribonuclease S-protein, an insect-specific protein of any one of claims 5 to 18 and an auxiliary protein according to any one of claims 19 to 23.
29. A fusion protein according to claim 27 comprising an insect-specific protein according to claim 5 and an auxiliary protein according to claim 19 having either the insect-specific protein or the auxiliary protein at the N-terminal end of the said fusion protein.
30. A fusion protein according to claim 29, comprising an insect-specific protein as given in SEQ ID NO:5 and an auxiliary protein as given in SEQ ID NO: 2 resulting in the protein given in SEQ ID NO: 23 including homologues thereof.

31. A fusion protein according to claim 29, comprising an insect-specific protein as given in SEQ ID NO:35 and an auxiliary protein as given in SEQ ID NO: 27 resulting in the protein given in SEQ ID NO: 50 including homologues thereof.
32. A fusion protein according to claim 28 comprising an insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 fused to a signal sequence, which is of heterologous origin with respect to the recipient protein.
33. A fusion protein according to claim 32, wherein the said signal sequence is a secretion signal.
34. A fusion protein according to claim 32, wherein the said signal sequence is a targeting sequence that directs the transgene product to a specific organelle or cell compartment.
35. A fusion protein according to claim 33 wherein the said protein has a sequence as given in SEQ ID NO: 43 including homologues thereof.
36. A fusion protein according to claim 34 wherein the said protein has a sequence as given in SEQ ID NO: 46 including homologues thereof.
37. A DNA molecule comprising a nucleotide sequence which encodes the protein of any one of claims 5-7, 9, 10, 12-15, and 19-22.
38. A DNA molecule comprising a nucleotide sequence which encodes the protein of any one of claims 8, 11, 16-18 and 23 to 36.
39. A DNA molecule comprising a nucleotide sequence which encodes an insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1.
40. The DNA molecule of claim 39, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO: 4, or SEQ ID NO: 6 including homologues thereof.
41. The DNA molecule of claim 39, wherein the said molecule comprises a nucleotide sequence as given SEQ ID NO:19, SEQ ID NO:28, SEQ ID NO:31, or SEQ ID NO:1 including homologues thereof.

42. A DNA molecule comprising a nucleotide sequence which encodes an auxiliary protein which enhances the insect-specific activity of an insect-specific protein.
43. The DNA molecule of claim 42 wherein the said molecule comprises a nucleotide sequence as given SEQ ID NO:19 including homologues thereof.
44. The DNA molecule according to any one of claims 37, 39, 40 or 42 which comprises a nucleotide sequence that has been optimized for expression in a microorganism.
45. The DNA molecule according to claim 37, 39, 40 or 42 which comprises a nucleotide sequence that has been optimized for expression in a plant.
46. The DNA molecule according to any one of claims 38, 41, or 43 which comprises a nucleotide sequence that has been wholly or partially optimized for expression in a microorganism.
47. The DNA molecule according to claim 38, 41 or 43 which comprises a nucleotide sequence that has been optimized for expression in a plant.
48. The DNA molecule of claim 45, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:17 or SEQ ID NO:18 including homologues thereof.
49. The DNA molecule of claim 47, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:27, or SEQ ID NO:30 including homologues thereof.
50. A DNA molecule which comprises a nucleotide sequence encoding a multimeric pesticidal protein, which comprises more than one polypeptide chains and wherein at least one of the said polypeptide chains represents an insect-specific protein of any one of claims 5 to 18 and at least one of the said polypeptide chains represents an auxiliary protein according to any one of claims 19 to 23, which activates or enhances the pesticidal activity of the said insect-specific protein.
51. The DNA molecule of claim 50 comprising a nucleotide sequence encoding an insect-specific protein of any one of claims 5 to 18 and an auxiliary protein according to any one of claims 19 to 23, which activates or enhances the pesticidal activity of the said insect-specific protein.

52. The DNA molecule of claim 51, wherein said molecule comprises a nucleotide sequence as given in SEQ ID NO:1 or SEQ ID NO:19 including homologues thereof.
53. A DNA molecule which encodes a fusion protein comprising several protein domains including at least an insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 produced by in frame genetic fusions, which, when translated by ribosomes, produce a fusion protein with at least the combined attributes of the insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 and, optionally, of the other components used in the fusion.
54. The DNA molecule of claim 53 which encodes a fusion protein comprising an insect-specific protein according to claim 5 and an auxiliary protein according to claim 19 having either the insect-specific protein or the auxiliary protein at the N-terminal end of the said fusion protein.
55. The DNA molecule of claim 53, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:22 including homologues thereof.
56. The DNA molecule of claim 53 which encodes a fusion protein comprising an insect-specific protein of any one of claims 5 to 18 and/or an auxiliary protein according to any one of claims 19 to 23 fused to a signal sequence, which is of heterologous origin respective to the recipient DNA.
57. The DNA molecule of claim 56, wherein the said signal sequence is a secretion signal.
58. The DNA molecule of claim 56, wherein the said signal sequence is a targeting sequence that directs the transgene product to a specific organelle or cell compartment.
59. The DNA molecule according to any one of claims 53 to 58, wherein at least one of its component sequences comprises a nucleotide sequence that has been optimized for expression in a microorganism.
60. The DNA molecule according to any one of claims 53 to 58, wherein at least one of its component sequences comprises a nucleotide sequence that has been optimized for expression in a plant.

61. The DNA molecule of claim 60, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO:42, SEQ ID NO:45, or SEQ ID NO:49 including homologues thereof.
62. The DNA molecule of claim 45, wherein the sequences encoding the secretion signal have been removed from its 5' end.
63. The DNA molecule of claim 62, wherein the said molecule comprises a nucleotide sequence as given in SEQ ID NO: 35 or SEQ ID NO:39 including homologues thereof.
64. A DNA molecule which hybridizes to a DNA molecule according to any one of claims 37-63 under moderately stringent conditions and which molecule has insect-specific activity.
65. The DNA molecule of claim 64, wherein hybridization occurs at 65°C in a buffer comprising 7% SDS and 0.5 M sodium phosphate.
66. An insect specific protein wherein the said protein is encoded by a DNA molecule according to claims 64 or 65.
67. An expression cassette comprising a DNA molecule according to any one of claims 37, 39, 40, 42, 44, 45 or 48 operably linked to plant expression sequences including the transcriptional and translational regulatory signals necessary for expression of the associated DNA constructs in a host organism and optionally further regulatory sequences.
68. An expression cassette comprising a DNA molecule according to any one of claims 38, 41, 43, 46, 47 or 49-65 operably linked to plant expression sequences including the transcriptional and translational regulatory signals necessary for expression of the associated DNA constructs in a host organism and optionally further regulatory sequences.
69. An expression cassette according to claim 67, wherein the said host organism is a plant.
70. An expression cassette according to claim 68, wherein the said host organism is a plant.
71. A vector molecule comprising an expression cassette according to claim 67 or 69.
72. A vector molecule comprising an expression cassette according to claim 68 or 70.

73. An expression cassette according to claims 69 or 70 or a vector molecule according to claims 71 or 73 which is part of the plant genome.
74. A host organism comprising a DNA molecule according to any one of claims 37, 39, 40, 42, 44, 45 or 48, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the genome of the host organism..
75. A host organism comprising a DNA molecule according to any one of claims 38, 41, 43, 46, 47 or 49-65, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the genome of the host organism..
76. A host organism according to claim 74 or 75, selected from the group consisting of plant and insect cells, bacteria, yeast, baculoviruses, protozoa, nematodes and algae.
77. A transgenic plant including parts as well as progeny and seed thereof comprising a DNA molecule according to any one of claims 37, 39, 40, 42, 44, 45 or 48, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the plant genome.
78. A transgenic plant including parts as well as progeny and seed thereof comprising a DNA molecule according to any one of claims 38, 41, 43, 46, 47 or 49-65, an expression cassette comprising the said DNA molecule or a vector molecule comprising the said expression cassette, preferably stably incorporated into the plant genome.
79. A transgenic plant including parts as well as progeny and seed thereof which has been stably transformed with a DNA molecule according to any one of claims 38, 41, 43, 46, 47 or 49-65.
80. A transgenic plant including parts as well as progeny and seed thereof which expresses an insect-specific protein according to any one of claims 5, 7, 9, 10, 12-15, or 19-22.
81. A transgenic plant including parts as well as progeny and seed thereof which expresses an insect-specific protein according to any one of claims 8, 11, 16-18, 23-36 or 66.

82. The transgenic plant according to claim 80 or 81, which further expresses a second distinct insect control principle.
83. The transgenic plant of claim 82, wherein said second insect control principle is a *Bt δ-endotoxin*.
84. A transgenic plant according to any one of claims 77-83, which is a maize plant.
85. A transgenic plant according to any one of claims 77 to 84, which is a hybrid plant.
86. Plant propagating material of a plant according to any one of claims 77 to 84 treated with a seed protectant coating.
87. A microorganism transformed with an expression cassette according to any one of claims 67 to 70 and/or a vector molecule according to any one of claims 71 or 72, wherein the said microorganism is preferably a microorganism that multiply on plants.
88. The microorganism of claims 87, which is a root colonizing bacterium.
89. An encapsulated insect-specific protein which comprises a microorganism of any one of claims 87 or 88 comprising an insect specific protein according to claims 18 or 23.
90. An entomocidal composition comprising a host organism of any one of claims 74-76 in an insecticidally-effective amount together with a suitable carrier.
91. An entomocidal composition comprising a purified *Bacillus strain* according to any one of claims 1 to 4 in an insecticidally-effective amount together with a suitable carrier.
92. An entomocidal composition comprising an isolated protein molecule according to any one of claims 5 to 36 and 66, alone or in combination with a host organism of any one of claims 74-76 and/or an encapsulated insect-specific protein according to claim 89 in an insecticidally-effective amount, together with a suitable carrier.
93. A method of obtaining a purified insect-specific protein according to any one of claims 5 to 36 said method comprising applying a solution comprising said insect-specific protein to a NAD column and eluting bound protein.
94. A method for identifying insect activity of an insect-specific protein according to any one of claims 5 to 36, said method comprising:

- (a) growing a *Bacillus* strain in a culture;
- (b) obtaining supernatant from said culture;
- (c) allowing insect larvae to feed on diet with said supernatant; and,
- (d) determining mortality.

95. A method for isolating an insect-specific protein according to any one of claims 5 to 36, said method comprising:

- (a) growing a *Bacillus* strain in a culture;
- (b) obtaining supernatant from said culture; and,
- (c) isolating said insect-specific protein from said supernatant.

96. A method for isolating a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein exhibiting the insecticidal activity of the proteins according to any one of claims 5 to 36, said method comprising:

- (a) obtaining a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein; and
- (b) hybridizing said DNA molecule with DNA obtained from a *Bacillus* species; and
- (c) isolating said hybridized DNA.

97. A method of increasing insect target range by using an insect specific protein according to any one of claims 5 to 36 in combination with at least one second insecticidal protein that is different from the insect specific protein according to any one of claims 5 to 36.

98. A method of increasing insect target range wherein an insect specific protein according to any one of claims 5 to 36 is expressed in a plant together with a at least one second insecticidal protein that is different from the insect specific protein according to any one of claims 5 to 36.

99. A method according to claim 97 or 98 wherein the second insecticidal protein is selected from the group consisting of *Bt* δ-endotoxins, protease inhibitors, lectins, α-amylases and peroxidases.

100. A method of protecting plants against damage caused by an insect pest comprising applying to the plant or the growing area of the said plant an entomocidal composition according to any one of claims 90 to 92.

101. A method of protecting plants against damage caused by an insect pest comprising applying to the plant a toxin protein according to any one of claims 5 to 36.
102. A method of protecting plants against damage caused by an insect pest comprising planting a transgenic plant expressing a insect-specific protein according to any one of claims 5 to 36 within an area where the said insect pest may occur.
103. A method of producing a host organism according to claim 74 to 76 comprising transforming the said host organism with a DNA molecule according to any one of claims 67 to 70 and 73 or a vector molecule according to claim 71 and 72.
104. A method of producing a transgenic plant or plant cell according to any one of claims 77 to 85 comprising transforming the said plant and plant cell, respectively, with an expression cassette according to any one of claims 70 or 73 or a vector molecule according to claim 72.
105. A method of producing an entomocidal composition according to any one of claims 90 to 92 comprising mixing a *Bacillus* strain according to any one of claims 1 to 4 and/or a host organism according to claim 74 to 76 and/or an isolated protein molecule according to any one of claims 5 to 36 and 66, and/or an encapsulated protein according to claim 89 in an insecticidally-effective amount with a suitable carrier.
106. A method of producing transgenic progeny of a transgenic parent plant comprising stably incorporated into the plant genome a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein according to any one of claims 5 to 36 and 66 comprising transforming the said parent plant with an expression cassette according to any one of claims 70 or 73 or a vector molecule according to claim 72, and transferring the pesticidal trait to the progeny of the said transgenic parent plant involving known plant breeding techniques.
107. An oligonucleotide probe capable of specifically hybridizing to a nucleotide sequence encoding an insect-specific protein isolatable during the vegetative growth phase of *Bacillus* spp. and components thereof, wherein said protein is not the mosquitocidal toxin from *B. sphaericus* SSII-1, wherein said probe comprises a contiguous portion of the coding sequence for the said insect-specific protein at least 10 nucleotides in length.

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108. Use of a oligonucleotide probe for screening of any *Bacillus* strain or other organisms to determine whether the insect-specific protein is naturally present or whether a particular transformed organism includes the said gene.
109. A DNA molecule comprising a nucleotide sequence which encodes the protein of any one of claims 8, 11, 16-18 and 23 to 36 obtainable by a process comprising
 - (a) obtaining a DNA molecule comprising a nucleotide sequence encoding an insect-specific protein; and
 - (b) hybridizing said DNA molecule with an oligonucleotide probe according to claim 107 obtained from a DNA molecule comprising a nucleotide sequence as given in SEQ ID NO: 28, SEQ ID NO: 30, or SEQ ID NO: 31; and
 - (c) isolating said hybridized DNA.

INTERNATIONAL SEARCH REPORT

In Application No
PCT/EP 95/03826

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 6 C12N15/32 C07K14/32 C07K14/325 C12N15/62 C12Q1/68
 C12N15/82 A01N63/00 A01H5/00 C12N1/21 G01N33/00
 //C07K16/12, C12N15/84, (C12N1/21, C12R1:07, 1:19, 1:085, 1:91)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 IPC 6 C07K C12N A01N A01H C12Q G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

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- *&* document member of the same patent family

Date of the actual completion of the international search

16 January 1996

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Hix, R

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INTERNATIONAL SEARCH REPORT

Int. Application No.
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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International Application No
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Information on patent family members

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